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RESEARCH ARTICLE

Ecological Observation on Phytoplankton Species Composition in Wastewater Treatment Plant / Iraq

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Abstract

The present study focuses on phytoplankton species composition in wastewater treatment plant. Semi-monthly sampling was collected from three selected sites to implement in the study plant from November, 2012 to April, 2013. Phytoplankton and physicochemical parameters investigated in the present study.

The results showed the dominance of non-diatom algae qualitatively and quantitatively. The non diatom algae represented 55.7% of the total identified algae, where Cyanophyceae, Chlorophyceae and Euglenophyceae represented 31.2%, 11.5% and 6.1%, while each of Xanthophyceae and Chrysophyceae represented 2.04% and 0.68% for Rhodophyceae. Diatom algae represented 44.2% of the total identified phytoplankton during the study period. The total number of phytoplankton ranged 2512-26649 cell x 10³/l, while chlorophyll-a ranged 7.57-44.88 mg/l.

Nine species of phytoplankton recorded in the present study as a new record in Iraq. These are *Anabaena naviculoides*, *Arthrospira jenneri*, *Camptylonemopsis*, *Lyngbya polysiphoniae*, *Echinospaerella limnetica*, *Excentrosphaera viridis*, *Sphaeroplea annulina*, *Trachelomonas dybowskii*, *Hemidinium nasutum*.

The physicochemical parameters were temperature (air and water), electric conductivity, salinity, pH, TDS and TSS. The study included dissolved oxygen, biochemical oxygen demand, total alkalinity, total hardness, calcium, magnesium, chloride, ammonia, nitrite, nitrate and phosphate. The study concluded that the major constituent of Almamierh wastewater treatment plant has level between medium to strong domestic wastewater.

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Introduction

The water scarcity is a real threat to many regions worldwide due to the huge usages of water. That increased threefold compared with 1950, As one out of six people cannot get safe drinking water, and the lack of potable water and sewage disposal affect the health of 1.2 billion people annually (WHO and UNICEF, 2000). In treatment plant, there are diverse organisms such as viruses, bacteria, phytoplankton, protozoa, Helminths, Fungi and invertebrates that play a major role in the recycling of energy and nutrients flow in these systems (Hoogenboezem, 2007).

Algae are the base of the food web in all aquatic ecosystems and larger organisms produced and stabilized energy. In addition to its role in providing a significant portion of the oxygen to the water and supporting the food web, which includes a wide range of consumers (Zooplankton, larvae, pupae insects and the different types of animals) food and energy needed to sustain it (Stefels and Boekel, 1993). Despite the availability of a large number of studies on the temporal and spatial variation of the presence of phytoplankton, the density and composition of the

community's rivers and lakes, but the information is still few about the density of phytoplankton in contaminated (sewage) water (Ramirez and Bicudo, 2005). Many authors recognized the importance of algal composition of the wastewater treatment either using them as treatment tools or to use the wastewater as media for getting mass algal culture (Moreno et al., 1990; Wong and Chan, 1990). While other authors investigated tolerance of different types of algae to pollution in diverse aquatic systems (Al- Saadi et al., 1989, Erganshev and Tajiev, 1986, Palmer, 1974). Pereira et al. (2001) found the wastewater treatment plant in northern Portugal as optimal habitat for Cyanophyceae. Silva (1998) found Chlorophyceae, Euglenophyceae, and Bacillariophyceae in addition to Cyanophyceae in wastewater treatment plant. Mahapatra et al. (2013) investigated the phytoplankton community and its effect on treatment efficiency in Vidyaranyaapuram (India) sewage treatment plant.

Phytoplankton is the source of the primary production in aquatic systems and it plays an important role in self treatments in sewage water (Gani et al. 2011) , but the intensive growth of these algae, particularly blue-greens algae, releases dangerous toxins (Neurotoxin & Hepatotoxin) that affect the liver and nervous system of humans if they consume this water for drinking (Humpage et al., 2000, Hassan et al., 2004) . The study aims to focus on the dominant algae in domestic water treatment; moreover, the qualitative and quantitative study of phytoplankton and their relation to environmental factors.

Materials and Methods

The present study has chosen the main domestic treatment plant in Al-Hilla Province, Iraq that locally called Almamierh. Almamierh located just 1 km from Al Hilla River and 5 km from the centre of the Al-Hilla City.

Three sites were selected in treatment plant (inlet effluent, aeration and out effluent), at a rate of 12 samples in semi-monthly sampling for the period from November 2012 to April 2013 (Fig.1).

Phytoplankton samples collected by phytoplankton net (20 μ m mesh size). The sample preserved with Lugol's solution (VollenWeider, 1974). A non-diatom alga was identified by using the following references; Prescott (1982) and Desikachary (1959). While for diatom algae identified after cleaning and mounting process (Hadi, 1984). The identification was made by following the references (Foged, 1976, Hustedt, 1985, Hadi et al., 1984, Germain, 1981, Al-Handal, 1994, Hassan et al., 2012, Al-Hassany and Hassan 2014).

The total number of phytoplankton done by a hemocytometer method for non-diatom algae and the micro-transect method for diatom algae (Stein, 1973; Hadi 1984). Chlorophyll-a measured by following the method described by Aminot and Rey (2001). All the studied physicochemical factors followed APHA (2003) except nitrite; nitrate and phosphate followed Parsons et al. (1984). The statistical analysis was conducted by applying Canonical Corresponded Analysis (CCA) using the program Canoco Ver.4.5



Figure 1: The study area

Results

A total of 147 species and 58 genera identified during the present study period. A total of 82 species of non-diatom algae were predominate and represented by 40 genera in this study. Higher number of taxon observed in site 3(129 taxa and 49 genera) while other sites (1 and 2) were 87 and 89 species (belonged to 39 genera for each site), respectively (Table 1).

Nine species of phytoplankton in this study were new recorded to Iraqi algal flora according to Maulood and Toma (2004). These species are as follows (Plate 1a and b):

A. Cyanophyceae:

- *Anabaena naviculoides* Fritsch
(Desikachary 1959. P:397,410 . Figures: 2 , 3.5-5 broad , as long as or shorter than broad)
- *Arthrospira jeneri* Stizenberger ex Gomont (Prescott 1973 . p:481 , pl:108, figures:22,23 . 6-8/ μ in diameter; 4-5/ μ long; distance between turns 12-14/ μ)
- *Camptyloneopsis minor* Desikachary
(Desikachary 1959. P:444 . figs:1,5 . 2.6-5.2 μ broad , 3.9-6.6 μ long)
- *Leptolyngbya polysiphoniae* (Frémy) Anagonostidis (*Lyngbya polysiphoniae* Frey)
(Desikachary 1959. P:306, figures: 5 , p:287 , up tp 200 μ long , cells about to 2 μ broad)

B. Chlorophyceae:

- *Echinospaerella limnetica* G.M. Smith
(Prescott 1973 . pl:51, figures:1,2. Cells 9-12/ μ in diameter, spines 2.5-3/ μ wide at the base, 20-25/ μ long.)
- *Excentrosphaera viridis* Moore
(Prescott 1973, pl:46, figures:15,16 . cells 22-55/ μ in diameter)
- *Sphaeroplea annulina* (Roth) C.A.Agardh
(Prescott 1973, PI. 12, figures: 5-8, Cells 27-72/ μ in diameter, up to 20 times longer than wide)

C. Euglenophyceae:

- *Trachelomonas dybowskii* Drezepolski
(Prescott 1973. PI. 83, Figure: 21; PI. 84, fig. 6. 10-18/ μ in diameter, 16-32/ μ long)

D. Pyrrophyceae :

- *Hemidinium nasutum* Stein
(Prescott 1973. PI. 90, Figures: 4-6. Cell 16-20/ μ in diameter, 24-28/ μ long)

Many dominant species observed in sites 1 and 2 (Table 2). There were: *Lyngbya epiphytica* , *L. nordgardii* , , *L. lagerheimmii* , *L. limnetica* , *L. versicolor* , *Oscillatoria acutissima* , *O. angutissima* , *O. formosa* , *O. hamelli* , *O. limnetica* , *Oscillatoria sp.* , *Phormidium fragile* , *P. tenue* and *Spirulina laxissima* .

While, at the site 3, the most dominant species were filamentous species such as; *Lyngbya* spp. Which appeared most of the months and in large numbers, and the most prominent types: *Lyngbya epiphytica* , *L. lagerheimmii* , *L. limnetica* , *L. versicolor* , *O. acutissima* , *O. angutissima* , *O. articulate* , *O. Formosa* , *O. limnetica* , *O. prolific* , *Oscillatoria sp.* and *Spirulina sp.* .

Total number of phytoplankton ranged 2542.4-26649.6 cell/ml at sites 2 and 3 in January 2013 and December 2012, respectively. Cyanophyceae was the great majority and represented 72.3% ($247895.2 \text{ cell} \times 10^3 / \text{l}$) of the total number of phytoplankton. While Bacillariophyceae and Chlorophyceae represented 11% and 6.67%, respectively. Other algal groups represented as follows: 6.34, 3.46%, 0.07%, 0.07% and 0.01 for Chrysophyceae, Euglenophyceae, Xanthophyceae, Pyrrophyceae and Rhodophyceae, respectively. Some alga obtained high density at study sites. *Oscillatoria acutissima* recorded high total number (6479 cells/ml) at site 1 in November, 2012. Also, this species is present in most of the study months at sites 2 and 3. *Oscillatoria angutissima* and *Phormidium tenue* were observed at all study sites while *Oscillatoria limnetica* obviously existed in site 1. The present study recorded predominant of alga *Spirulina* at site 3. The most common diatomic taxa were *Cyclotella meneghiniana*, *Cymbella tumida* and *Nitzschia palea*.

Ankistrodesmus falcatus appearance of a few at site 1, but it was increased in number in November, 2012 at site 1. While *Chlamydomonas* noticeably found in winter months at site 3.

Mallomonas sp. (Chrysophyceae) recorded a dense existing (20741 cell/ml) at site 3, while *Euglena elastic* (Euglenophyceae) found at site 3 frequently and reached 186.39cell/ml in April 2013. *Batrachospermum moniliforme* (Rhodophyceae) found at site 3.

Chlorophyll-a ranged 7.570- 44.884 mg/l at sites 2 (both numbers) in October 2012 and January 2013, respectively. No variation found in chlorophyll –a concentration among sites and the study months.

The values of studied parameters ranged as follows (Table 2): 12-36°C, 12-33 °C, 1351-4221 $\mu\text{s/cm}$ and 0.864-2.324‰ for temperature (air and water), electric conductivity (EC) and salinity. While 7.0-7.8, 1920-2851mg/l, 21-1788mg/l recorded for pH, TDS and TSS. Dissolved oxygen and biochemical oxygen demand ranged 0.6-5.9mg/l and 20-673 mg/l. Total alkalinity, total hardness, calcium, magnesium and chloride ranged 15-190 mg CaCO_3 /l, 860-3100 mg CaCO_3 /l, 164.321-641.286mg/l, 26.549-629.126mg/l and 340.329-721.323mg/l. Nutrients ranged 0.194-19.891mg/l, 0.165- 5.339mg/l, 10.415-100.544mg/l and 22.379-196.6mg/l for Ammonia, nitrite, nitrate and phosphate.

CCA analysis presented in fig 2 showed a correlation between the studied factors in site 1 (part 1 of axis) and both Cyanophyceae and Bacillariophyceae. Cyanophyceae presence was related to Mg, BOD₅, TSS, Nitrite, Salinity and Alkalinity while Bacillariophyceae associated with pH, temperature, Chloride and phosphate. In part 2 axes, Euglenophyceae and Chlorophyceae associated with Ammonia and dissolved oxygen while Chrysophyceae and Xanthophyceae were correlated with Nitrate, TDS, Hardness and Ca. In site 2 (aeration site) Cyanophyceae associated closely with most of the studied factors, while Chlorophyceae and Chrysophyceae correlated with pH, dissolved oxygen, nitrate, Phosphate and Ca (first part of axes). Bacillariophyceae and Euglenophyceae associated with phosphate, BOD₅, temperature, TSS and ammonia (part 2 axes). Different manner occurred in site 3 (effluent site); Cyanophyceae, Bacillariophyceae, Chlorophyceae, Xanthophyceae, Pyrrophyceae and Rhodophyceae were related to TSS and chloride closely in part 2axes. In part 1 axes in site 3; Chrysophyceae correlated with pH, nitrite, phosphate and Mg, while Euglenophyceae correlated with temperature, nitrate, alkalinity and BOD₅.

Table 1: Numbers of species, genera and the percentage of phytoplankton at Almamierh plant during the study period.

Class	Site1			Site 2			Site 3			Total		
	G	sp	%	G	sp	%	G	sp	%	G	sp	%
Cyanophyceae	12	14	38.2	9	26	29.2	14	38	29.4	15	46	29.4
Chlorophyceae	7	7	7.8	9	10	11.2	8	9	7	14	17	7.0
Euglenophyceae	4	6	6.7	3	4	4.4	3	8	6.2	4	9	6.2
Pyrrophyceae	-	-	-	-	-	-	2	3	2.3	2	3	2.3
Rhodophyceae	-	-	-	-	-	-	1	1	0.7	1	1	0.8
Chrysophyceae	1	1	1.1	2	2	2.2	2	3	2.3	2	3	2.3
Xanthophyceae	1	1	1.1	-	-	-	2	3	2.3	2	3	2.3
Bacillariophyceae												
Centrales	2	6	6.7	2	5	5.6	2	9	7	2	9	7
Pennales	38.2	34	38.2	14	42	47.1	15	55	42.6	16	56	42.6
Total	40	89		39	89		49	129		58	147	

Table 2: Identified wastewater phytoplankton during study sites.

* Species recorded for the first time in Iraq

Taxa	Sites		
	1	2	3
Cyanophyceae			
<i>Anabaena naviculoides</i> Fritsch*	+	-	-
<i>A. sp.</i>	+	-	-
<i>Arthrospira jenneri</i> Kuetz. *	+	-	-
<i>Calothrix sp.</i>	+	+	+

<i>Camptylonemopsis minor</i> Desikachary *	-	+	-
<i>Camptylonemopsis</i> sp	+	+	+
<i>Chroococcus limnaticus</i> var. <i>distans</i> G.M.Smith	-	-	+
<i>C. Turgidus</i> Kuetz.	-	-	+
<i>Dactylococcopsis fascicularis</i> Lemmermann	+	+	+
<i>Gloeocapsa</i> sp.	+	+	+
<i>Lyngbya epiphytica</i> Hieron	+	+	+
<i>L. lagerheimmii</i> (moeb) Gomont	+	+	+
<i>L. limnetica</i> Lemmermann	+	+	+
<i>L. nordgardii</i> Wille	+	+	+
<i>L. polysiphoniae</i> Frey *	-	+	-
<i>L. versicolor</i> (wartmman) Gomont	+	+	+
<i>Merismepdia glauca</i> (Ehr.) Naegeli	+	-	+
<i>Nostoc carneum</i> Agardh	+	-	+
<i>Nostoc</i> sp.	+	+	+
<i>Oscillatoria acutissima</i> Kufferath	+	+	+
<i>O. amoena</i> (Ktz) Gomont	+	+	-
<i>O. amphibian</i> agardh	+	-	+
<i>O. amphigranulate</i> var Goor	+	-	+
<i>O. anguina</i> (Bory) Gomont	-	+	-
<i>O. angutissima</i> West and West	+	+	+
<i>O. articulate</i> Gardner	+	+	+
<i>O. chalybea</i> Mertens	-	+	-
<i>O. formosa</i> Bory	+	+	+
<i>O. hamelli</i> Frey	+	+	+
<i>O. limnetica</i> Lemmermann	+	+	+
<i>O. minima</i> Gicklhorn	-	+	+
<i>O. prolific</i> (Grev.) Gomont	+	-	+
<i>O. splendid</i> Greville	+	-	+
<i>O. willei</i> Gardner	+	-	+
<i>Oscillatoria</i> sp.	+	+	+
<i>Phormidium fragile</i> Gomont	-	+	+
<i>P. mucicola</i> (Huber-pest and Naumann)	-	+	-
<i>P. tenue</i> (Menegh) Gomont	+	+	+
<i>P. molle</i> Gomont	-	-	+
<i>Rivularia haematites</i> (D.C.) C.A. Agardh	+	-	-
<i>Spirulina laxa</i> G.M. Smith	+	-	-
<i>S. laxissima</i> G.S. West	+	+	+
<i>S. major</i> Ktz.	+	-	-
<i>S. subsalsa</i> Orested	+	-	-
<i>Spirulina</i> sp.	+	-	-
<i>Synechococcus aeruginosus</i> Naegeli	+	-	+
Chlorophyceae			
<i>Ankistrodesmus falcatus</i> (Cord.) Ralfs	+	-	+
<i>Chlamydomonas epiphytica</i> G.M. Smith	-	+	-
<i>C. snowii</i> Printz	-	+	+
<i>C.</i> sp.	+	-	-

<i>Chlorococcum humicola</i> Neag.	-	+	+
<i>Closteridiopsis longissima</i> lemmerman	+	+	+
<i>Echinosphaerella limnetica</i> G.M. Smith *	-	+	-
<i>Excentrosphaera viridis</i> Moore *	-	+	-
<i>Sphaeroplea annulina</i> (Roth) C.A.Agardh *	+	-	-
<i>Scendesmus acuminatus</i> Var. minar G.M. Smith	+	-	-
<i>Schizomeris leibleinii</i> Kuetzing	-	-	+
<i>Spirogyra proticalis</i> (Muell.) petit	-	-	+
<i>Stigeoclonium subceccandum</i> Ktz.	-	+	-
<i>Tetradron regular</i> Ktz.	+	+	-
<i>Treubaria setigerum</i> (Archer) G.M. Smith	+	+	-
<i>Ulothrix zonata</i> (Weber & Mohr) Kuetzing	+	+	+
<i>U. tenerrima</i> ktz.	+	-	-
Euglenophyceae			
<i>Euglena elastic</i> prescott	+	-	+
<i>E. polymorpha</i> Dangeread	+	-	+
<i>E. proxima</i> Dangeread	+	-	-
<i>Euglena</i> sp.	+	+	+
<i>Lipocinalis shagnophila</i>	-	-	+
<i>Phacus caudataus</i> Huebner	+	+	-
<i>Phacus</i> sp.	+	+	+
<i>Trachelomonas dybowskii</i> drezepolskii *	+	-	-
<i>Trachelomonas volvocina</i> Ehrenbergo	+	-	-
Pyrrophyceae			
<i>Hemidinium nasutum</i> Stein *	+	-	-
<i>Peridinium pussilum</i> (Penard) Lemmerman	+	-	-
<i>Peridinium</i> sp.	+	-	-
Rhodophyceae			
<i>Batrachospermum moniliforme</i> Roth	-	-	+
Chrysophyceae			
<i>Dinobryon bavaricum</i> Imof.	+	+	-
<i>Dinobryon</i> sp.	+	-	-
<i>Mallomonas</i> sp.	+	+	+
Xanthophyceae			
<i>Characiopsis acuta</i> (A.Braun) Borzi	+	-	-
<i>C. Longiceps</i> (Rabenh.) Borzi	+	-	-
<i>Chlorochromonas minuta</i> lewis	+	-	+
Bacillariophyceae			
Centrales			
<i>Cyclotella comta</i> (Ehr.) Kuetzing	+	+	+
<i>C. kuetzingiana</i> var. <i>radiosa</i> Pricke	+	-	-
<i>C. meneghiniana</i> Kützing	+	+	+
<i>C. ocellata</i> Pantocsek	+	+	+
<i>C. striata</i> (ktz) Grunow	+	-	-
<i>Melosira italic</i> (Ehr.) Kuetzing	+	-	+
<i>M. granulate</i> (Ehr.) Ralfs	+	+	+

<i>M. varains</i> Agradh	+	-	-
<i>M. distance</i> (Ehr.) Kuetzing	+	+	-
Pennales			
<i>Amphiprora alata</i> kutz.	+	-	-
<i>Asterionella</i> sp.	+	-	-
<i>Cocconeis placentula</i> Ehrenberg	+	+	+
<i>Cymbella affinis</i> (Kuetzing)	+	+	+
<i>C. cuspidata</i> Ktz. Var. <i>anglica</i> lagst	+	+	+
<i>C. cymbiformis</i> (Ktz.) Van Heurck	+	+	+
<i>C. hustedtii</i> Krasske	+	-	-
<i>C. tumida</i> (Breb.) van. Heurck	+	+	+
<i>C. ventricosa</i> Kuetzing	+	+	+
<i>Cymatopleura elliptica</i> (Breb.) W.Smith	+	+	+
<i>Diatoma elongatum</i> (Lyngb.) Agardh	+	+	-
<i>D. hiemala</i> (Roth) Heiberg	+	+	+
<i>D. vulgare</i> Bory	+	+	+
<i>Diploneis Ovalis</i> (Hilse) Cleve	+	+	+
<i>D. pseudovalis</i> Hustedt	+	-	+
<i>Eunotia praerupta</i> Ehrenberg	-	+	+
<i>Fragilaria brevistriata</i> var. <i>inflata</i> (Pant.) Hustedt	+	+	+
<i>F.capucina</i> var. <i>gracilis</i> (Oster.) A.Cleve	+	+	+
<i>F. crotonensis</i> Kitton	+	+	+
<i>Fragilaria</i> sp.	+	+	+
<i>Gomphonema constrictum</i> Ehrenberg	+	+	+
<i>G. longiceps</i> Ehr.	+	+	-
<i>Gyrosigma scalproides</i> (Rabenhorst) Cleve	+	+	+
<i>Gyrosigma spenceri</i> (Quek.) Griff et Henfr	+	-	-
<i>Gyrosigma</i> sp.	+	-	-
<i>Meridion circulare</i> Agardh	+	+	-
<i>Navicula anglica</i> Ralf	+	-	-
<i>N. bacillum</i> Ehrenberg	+	+	+
<i>N. cryptocephale</i> Kuetzing	+	+	+
<i>N. crptocephalae</i> var. <i>veneta</i> (Ktz.) Cleve	+	+	+
<i>N. cuspidata</i> (Ktz.) Kuetzing	+	+	-
<i>N.dicephala</i> var. <i>neglecta</i> (Krasske) Hust.	+	-	-
<i>N. halophila</i> (Grun.) Cleve	+	+	+
<i>N. lanceolata</i> (Ag.) Kuetzing	+	+	-
<i>N. psuedotuscula</i> Hustedt	+	+	-
<i>N. virtea</i> (Oester.) Hustedt	+	+	+
<i>Nitzschia acuta</i> Hantzsch	+	+	-
<i>N. amphibia</i> Grunow	+	-	-
<i>N. angustata</i> (W.Smith) Grunow	+	+	+
<i>N. dissipata</i> (Ktz.) Grunow	+	+	+
<i>N. hantzschiana</i> Rabenhorst	+	+	-
<i>N. hungarica</i> Grunow	+	+	-
<i>N. linearis</i> W.Smith	+	+	-
<i>N. lottoralis</i> Grunow	+	+	-

<i>N. longissima</i> (Breb.) Ralfs	+	-	-
<i>N. palea</i> (Ktz.) W.Smith	+	+	+
<i>N. punctata</i> (W.Smith) Grunow	+	-	+
<i>N. recta</i> Hantzsch ex Rabenh.	+	+	+
<i>N. sigma</i> (Ktz.) W.Smith	+	-	-
<i>N. sigmoidea</i> (Ehr.) W.Smith	+	+	+
<i>N. vermicularis</i> (Ktz.) Hantzsch	+	+	-
<i>Surirella ovate</i> Kützing	+	+	+
<i>Syndra acus</i> Kützing	+	-	+
<i>S. tabulate</i> var. <i>fasciculate</i>	+	+	+
<i>S. ulna</i> (Nitz.) Ehrenberg	+	+	+
<i>S. ulna</i> var. <i>longissima</i> (W.Smith)	+	-	+
Brun			

Table 3: Range (Mean \pm SD) of Physicochemical parameters for Almamierh plant.

Parameters	Sites		
	St.1	St.2	St.3
Air temperature C°	13 – 36 (21.5 \pm 6.06)	15 – 36 (22 \pm 6.24)	12 – 36 (22.07 \pm 6.32)
Water temperature C°	12-33 (19.08 \pm 6)	12 – 28 (18.797 \pm 5.20)	12 – 29 (18.53 \pm 5.48)
EC μ s/cm	1911 – 4221 (3625.3 \pm 673.30)	1817- 3941 (3319.1 \pm 596.1)	1351–4061 (3174.5 \pm 762.7)
Salinity ‰	1.22 – 2.77 (0.44 \pm 2.32)	1.164-2.53 (2.13 \pm 0.39)	0.86-2.59 (2.03 \pm 0.49)
pH	7.3 – 7.8 (7.68 \pm 0.12)	7.2 – 7.8 (7.60 \pm 0.17)	7 – 7.8 (7.69 \pm 0.18)
TDS (mg/l)	2452 – 3651 (2821.75 \pm 293.80)	2241–2785 (2524.42 \pm 152.97)	1920-2851 (2431.58 \pm 261.47)
TSS (mg/l)	52– 407 (179.08 \pm 79.29)	448- 1788 (960.75 \pm 346.09)	21 – 64 (35 \pm 12.00)
DO (mg/l)	0.6 – 2.1 (1.2 \pm 0.48)	1.3 – 4.9 (2.87 \pm 1.14)	1.8 – 5.9 (3.93 \pm 1.3)
BOD ₅ (mg/l)	170 – 673 (373.33 \pm 182.97)	120 – 317 (197.83 \pm 48.11)	20 – 63 (38.50 \pm 11.45)
Total alkalinity (mg CaCO ₃ /l)	30 – 190 (112.64 \pm 42.3)	15 – 170 (109.83 \pm 43.40)	23 – 162 (98.33 \pm 42.81)
Total hardness (mg CaCO ₃ /l)	1010 – 2500 (1455.4 \pm 400.90)	900- 3100 (1592.5 \pm 569.23)	860 – 2400 (1431.66 \pm 450.43)
Calcium (mg CaCO ₃ /l)	188.38–360.72 (253.93 \pm 57.90)	188.38-641.29 (279.87 \pm 123.60)	164.32-480.96 (272.59 \pm 88.42)
Magnesium (mg CaCO ₃ /l)	33.81 - 480.89 (199.63 \pm 105.65)	26.55-629.13 (199.99 \pm 47.83)	31.47-485.84 (187.39 \pm 83.07)
Chloride (mg/l)	382.86-719.77 (552.26 \pm 109.21)	382.42-719.13 (550.07 \pm 113.99)	340.33-721.32 (551.76 \pm 120.29)
Ammonia (mg/l)	4.65 –19.89 (12.33 \pm 4.59)	0.76- 16.92 (8.84 \pm 5.45)	0.19- 12.93 (6.02 \pm 2.12)
Nitrate (mg/l)	10.42 - 84.2 (35.68 \pm 16.13)	27.59- 97.96 (63.06 \pm 24.01)	10.49-100.54 (56.57 \pm 20.62)
Nitrite (mg/l)	0.17 – 2.86 (1.75 \pm 0.67)	0.28- 5.75 (3.23 \pm 1.50)	0.20-5.34 (2.81 \pm 1.05)
Reactive Phosphate (mg/l)	80.25- 196.60 (155.67 \pm 42.73)	60.74-162.80 (90.92 \pm 31.17)	22.38-75.17 (44.78 \pm 17.91)
Chlorophyll-A (mg/l)	10.66 – 37.12 (18.19 \pm 7.11)	7.57 – 44.88 (20.71 \pm 9.81)	8.32-39.89 (18.61 \pm 9.10)
Total Phytoplankton cell*10 ³ /L	5038- 18729(10264 \pm 3909)	2512-26557 (8139 \pm 3488)	5323 -26650 (10154 \pm 4696)

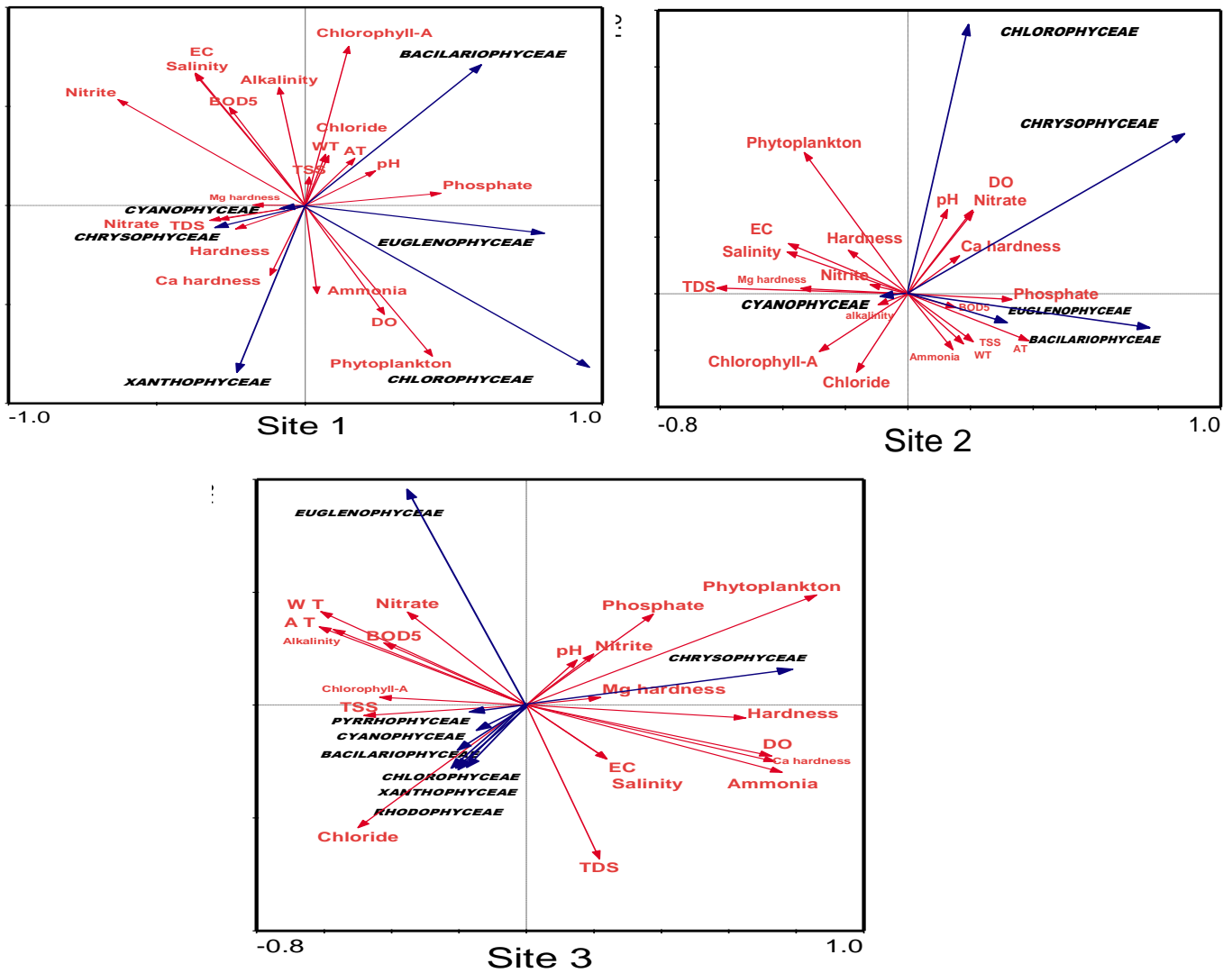


Figure2: Correlations between water quality parameters and phytoplankton groups in study sites according to CCA.

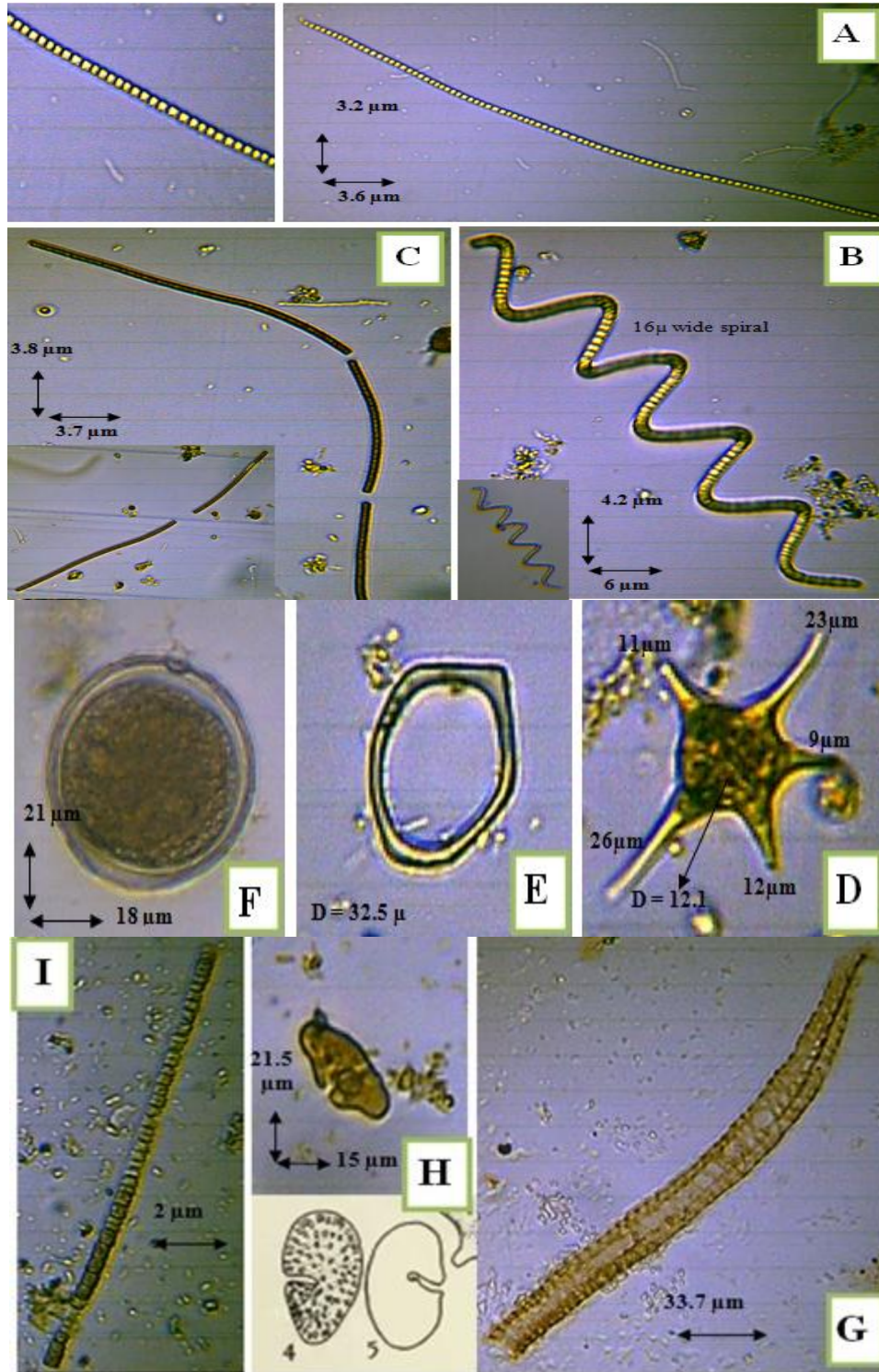


Plate1: Photos of new record to algal flora in Iraq during the present study.

A: *A. naviculoides* B: *A. jenneri* C: *C. minor* D: *L. polysiphoniae*
 E: *E. limnetica* F: *E. viridis* G: *S. annulina* H: *T. dybowskii*
 I: *H. Nasutum*

Discussion

The phytoplankton population structures are affected by surrounding environmental changes in aquatic systems (Zębek, 2004). Since the availability of nutrients and suitable water temperature led the growth of a diverse group of algae and may be suitable for a few algal groups, moreover some other abundant groups can effect the presence of another alga (Joseph and Joseph, 2002). These were appearing obviously during the current study. *Cyclotella meneghiniana*, *Cymbella tumida* and *Nitzschia palea* were found in this study area frequently, which indicated to inorganic nutrient enrichment in untreated sewage (Al-Hansan et al., 1990). Also, numerous studies have confirmed the possibility of the growth of algae effectively in the environment rich in organic matter and metals in the wastewater (Pittman et al., 2011). But the increase of organic pollutants led to increased toxic algal groups, despite that these toxins break down when adding chlorine for the purpose of sterilization by chlorination (Wang et al., 2007).

In the present study, it was observed that the non-diatom algae was predominate algae during the study period. That may be due to the effect of different pollutants (Palmer 1969; Bellinger and Sigeo, 2010). The total number of diatom algae in this study recorded less than the non-diatom algae. This finding differs from that recorded in Iraqi natural aquatic systems, in which the diatom algae are dominant algal groups (Sulaiman et al., 2000; Hassan et al., 2011, Maulood et al., 2013).

Different number of phytoplankton recorded in the study sites, where in site 1 (In-Effluent) and site 2 (Aeration) were 89 taxa while in site 3 (Out-Effluent) which represented the treated water that was 129 taxa. This difference was may be due to the physical and chemical properties as well as the quality of water for each site where the great similarity in the qualitative composition between the three study sites may be due to the main source of water (Al-Sarraf, 2006).

The abundance of Cyanophyceae in wastewater in this study was in accordance with other published manuscripts (Fulke et al., 2013; Ghughtai et al., 2013; Kaparapu and Geddada, 2013).

The occurrence of Bacillariophyceae in the wastewater represented its high ability to environmental change, which led to use some abundant diatom species as bioindicators for organic pollution or heavy metals such as *Cyclotella* and *Nitzschia* (Palmer, 1969). Moreover, the Iraqi aquatic systems contain high concentrations of silica (Hassan, 1997) which is necessary for the growth of this group in addition to the possibility of these algae to withstand changes in environmental conditions (Graham & Wilcox, 2000). The presence or absent of some phytoplankton species at different sites that may be as a response to their eco-physiological response to pollution (Guo et al., 2010).

Chlorophyceae found in the aeration tank where the high rate of dissolved oxygen helps to bloom phytoplankton. As well as the latter Chlorophyceae species that appeared in the treated water and this may be due to improved water quality after passing through the stages of treatment.

Other phytoplankton species such as; *Trachelomonas dybowskii* (Euglenophyceae) and *Hemidinium nasutum* indicated to wastewater pollution and to tolerant the salinity and pH (Al-Saadi et al., 2002).

The physicochemical characteristic of site 1 (Effluent) was showed medium to strong domestic wastewater composition according to typical guide of domestic wastewater (UN Department of Technical Cooperation for Development, 1985), also these results were in accordance with arid and semi-arid neighbouring countries (Al-Salem, 1987, Mohammed et al., 2012). High values of organic content, TDS, TSS, Chloride, nitrate and phosphate represented strength level in the present study at site 1. While the value of pH was typical values that may be due to buffer potential of the Iraqi water (Hassan et al., 2010). The aeration process in site 2 affected the physicochemical characteristic in this site. This process led to decrease of most of studied parameters except some of them that may be due to high concentration of EC and chlorides (Mohammed et al., 2012), which have role in treatment processes, on the other hand, the effect of phytoplankton obviously shown in site 2 and site 3.

The study recorded high density of phytoplankton in wastewater, which has a role in treatment plant. Cyanophyceae was dominant in the study treatment plant more than other classes of phytoplankton. Many phytoplankton species that have recorded may be used as bioindicators to pollution by wastewater and ecosystem health. The domestic wastewater of the Hilla city ranged between medium to strong.

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