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RESEARCH ARTICLE

Detection Norovirus and Rotavirus using Transmission Electronic Microscope

¹Nadira S.Mohamed,²Faisal G. Nasser and ³Khaled A.Habeb

1. AL-Nahrain Research & Training Center for forensic DNA, Baghdad.

2. Central public Health Laboratory, Baghdad.

3. College of Science for Women, University of Baghdad.

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Abstract

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Transmission Electronic Microscope (TEM) continues to play a role in the understanding of viral gastroenteritis.

Ten samples proven to be positive *Norovirus* by real time PCR and two ELISA *Rotaviruse* positive sample were selected and examine under Transmission Electrone microscope magnified 80000-100000 time .we found that *Rotaviruse* easy detected because the viruses shedding in large quantities and has a circular shape size of 65-70 nm ranges. We found that *Norovirus* diagnosis still difficult and needs to concentrate the sample to increase the number of virus particles and its size ranged from approximately 38-40 nm. Because there is not a previous study of *Norovirus* in Iraq, we decided to take a micrograph of the virus to find out size and shape of the virus we previously isolated.

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Introduction

It was only with the development of Transmission Electron Microscopy (TEM) that virus structure could be studied in detail (Finner and White, 1976), and TEM has had a significant impact on the development of virology in general (Palmer and Martin, 1988; Marshall, 2005). The accumulated knowledge on virus ultrastructure formed the grounds for the first classification of viruses based on virus intrinsic properties rather than on host characteristics and diseases produced by them (Finner et al., 1974).

The technique of transmission electron microscopy (TEM) was crucial in the discovery of the major viral causes of gastroenteritis in humans (*Norovirus*, *Rotavirus*, *Astrovirus*, *Sapovirus*, *Adenovirus*) and subsequently played a valuable role in the detection of these viruses. In the 21st century, TEM continues to play a role in the understanding of viral gastroenteritis but chiefly in a research role rather than in a diagnostic context (Marshall, 2012).

TEM continues to play a valuable research role in the identification and characterization of viruses as associated with the gastrointestinal tract in both humans and animals. (Marshall, 2012)

Negative staining TEM has proved to be a valuable approach for the detection of fecal viruses in a variety of animals (13-15) and this approach remains relevant today. Negative staining TEM was also used to identify *Rotavirus* in an English red squirrel (19) and has also been used to identify *Norovirus*-like particles in monkeys (Wang et al., 2007).

Negative staining TEM is a valuable technique to monitor the presence of recombinant *Norovirus* virus-like particles (Wang et al., 2007; Taube et al., 2005).

The method is also useful in evaluating the sensitivity and specificity of commercial virus detection kits (Curry et al., 2006). The family *Reoviridae* includes the genus *Rotavirus*. Human and animal rotaviruses are known. Structure. *Rotavirus* was first identified by electron microscopy in 1973 from duodenal biopsies of children with diarrhea (Vale et al., 2010). Rotaviruses are non-enveloped viruses with icosahedral symmetry and a double capsid. Their electron microscopic appearance shows a 60-80 nm wheel with radiating spokes (Latin, Rota = wheel). The rotavirus genome contains double stranded (ds) RNA in 11 segments. (Narayan and Albercht, 2013).

Rotaviruses are found worldwide, causing major diarrhea-associated hospitalization and 600,000-850,000 deaths per year. Seroprevalence studies show that antibody is present in most infants by age 3 years.

In the U.S., about 2.7 to 3.5 million people affected by *Rotavirus* infections each year and 20 - 40 deaths per year with 50-70,000 hospitalizations and 500,000 physician visits per year. In about 1-2.5% of case there is severe dehydration. In all, there are probably about 2.7 to 3.5 million people in the US affected by *Rotavirus* infections each year. *Norovirus* belong to Human caliciviruses were first described in 1976. Norwalk virus was first detected in stools of patients with gastroenteritis (winter vomiting disease) in Norwalk, Ohio in 1968. They cause 40 per cent of non-bacterial gastroenteritis epidemics. Forty five per cent are food-borne and 52 percent are raw shell-fish associated. They tend to cause rapid (explosive) epidemics in places of close contact such as cruise ships, nursing homes, hospitals and camps (Eckardt & Baumgart, 2011). *Noroviruses* are found world-wide and cause more than 23 million cases of gastroenteritis very year in the United States and are non-enveloped, single strand, positive sense RNA viruses. They appear round in shape with icosahedral symmetry and ragged surface contain a single capsid protein. (Narayan and Albercht, 2013). The virus particles demonstrate an amorphous surface structure when visualized using electron microscopy and are between 27-38 nm in size (Prasad et al., 2001). *Norovirus* affects around 267 million people and causes over 200,000 deaths each year; these deaths are usually in less developed countries and in the very young, elderly and immunosuppressed (Debbink et al., 2012; Marshall, 2011).

Material and Methods

About 20% of stool count for 10 *Norovirus* positive results by Real Time PCR positive, and only two ELISA *Rotavirus* were suspended phosphate – buffered saline (PBS).

Fecal suspension was clarified at 2000g for 10 min twice. For negative staining, a drop of about 10 µl of the virus suspension to be studied is applied to the hydrophobic surface of a parafilm square in a Petri dish. A formvar-coated grid is floated onto this drop for one minute to hold the small particles, with the formvar side of the grid in contact with the liquid. The excess liquid is removed from the grid by touching its border with a cut piece of filter paper. The grid is immediately floated in a drop of 1.5% phosphotungstic acid, and 1% aqueous uranyl acetate. For a better assessment of the samples, three grids should be prepared, each stained with one of

these stains. After staining for one minute, the excess stain is removed with filter paper and the grid left to dry for a few minutes, before insertion into the microscope column Transmission Electron Microscopy (Zeiss EM10 OCR from Germany) (Goldsmith and Miller, 2009).

Concentration methods for *Norovirus* detection using Ammonium sulphate precipitation.

Fecal suspension 30% PBS were centrifuged at 2500 g for 20 min at 2 °C, then 5 ml of supernatant were added to equal volume of 5 ml of pre-cooled saturated ammonium sulphate. After incubation on ice for 90 min, the mixtures were centrifuged at 2300 g for 30 min at 2 °C. The pooled precipitates were dissolved in 0.5 ml of 10 mM-tris-HCl buffer, pH 7.2, 0.1 M-NaCl, 1 mM EDTA (TNE buffer) (Tamura, 1978).

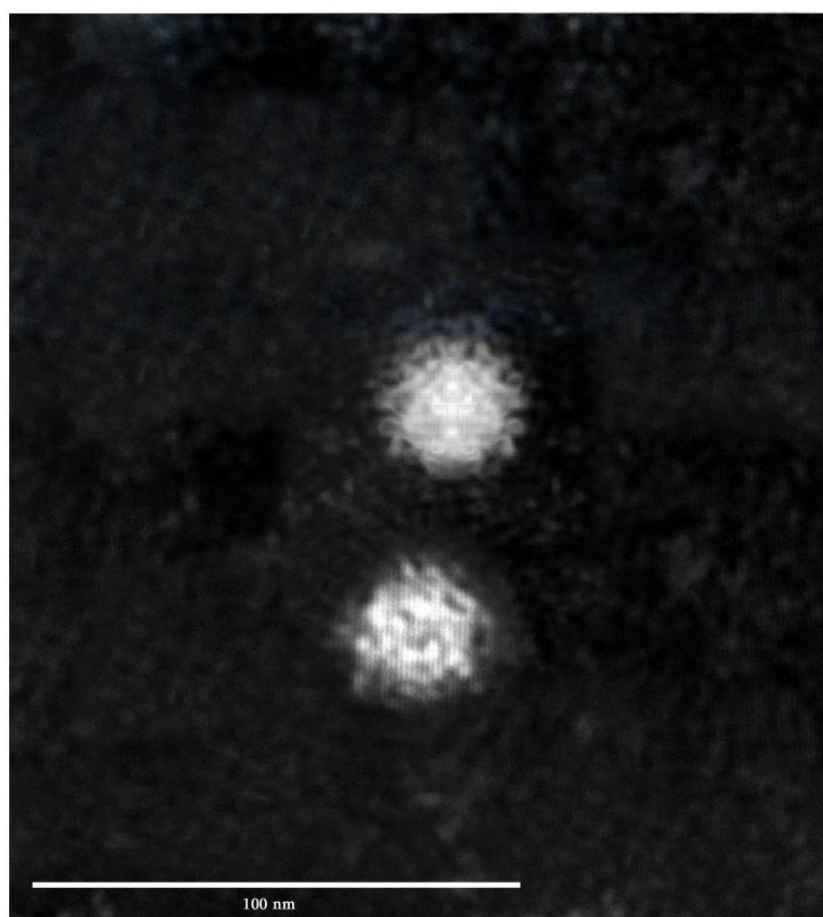
Result and Discussion

The minimum virus concentration necessary for successful detection by TEM is about 10^6 particles per ml. This is a rather concentrated suspension and viruses are often present in diseased tissues and organic fluids at considerably lower concentrations. Another problem in fecal samples is the presence of other structures like bacteria and cell debris that may make virus identification difficult. In these cases, viruses must be separated from contaminating debris using centrifugation twice and concentrated of *Norovirus* samples using ammonium sulphate because of the low shedding number of virus to get more than 10^6 particles which is a workable preparation for electron microscopy. Negative staining is a rapid procedure used for viewing small particles, such as viruses, in fluids. Phosphotungstic acid (PTA) sometimes outlines fringe/spikes on enveloped viruses better and does not cause positive staining. Uranyl acetate acts as a fixative as well as a stain, and viruses can be viewed intact long after the initial diagnosis. PTA actively degrades some viruses, and while immediate visualization is possible, viruses fall apart in just a few hours after staining as mentioned in (Hayat and Miller, 1990). Samples selected from positive ELISA and PCR *Rotaviruses* and *Noroviruses* previously diagnosed. Molecular or immunological methods, such as Real time PCR and ELISA, are very effective in detecting known viruses and can be used to scrutinize a large amount of sample. Diagnostic electron microscopy has two advantages over enzyme-linked immunosorbent assay and nucleic acid amplification tests. After a simple and fast negative stain preparation, the undirected, “open view” of electron microscopy allows rapid

morphologic identification and differential diagnosis of different agents contained in the specimen. Electron Microscopy remains the only method that can provide a quick assessment of all pathogens present in a sample, providing an "open view." Electron microscopy is still a major and indispensable inspection technique for Noroviruses, which cannot still be cultured as mentioned in (Gurry, et al 2006; Utagawa, 2004; Vieler and Herbst, 1995). TEM also plays a valuable role in the identification and characterization of both Noroviruses and

Rotaviruses in stool sample which collected from Iraqi children. As shown in figure 1, 2, and 3 the Norovirus particle has icosahedral symmetry and a ragged surface. It contains a single capsid protein 32-37 nm. We found in figure 4 and 5 the Rotaviruses particles have icosahedral symmetry and a double capsid with spikes 76 nm. Wheel with radiating spokes comparison with similar results of (Marshall 2005; Narayan and Albercht, 2013).

Fig.1:- Norovirus two particles stained with negative staining under TEM



Noro Virus

Amount of High Voltage used : 80 kV

Electronic Microscope Magnification : 100000

Total Magnification : 386360

Fig. 2:- Noroviruse particles stained with negative staining under TEM

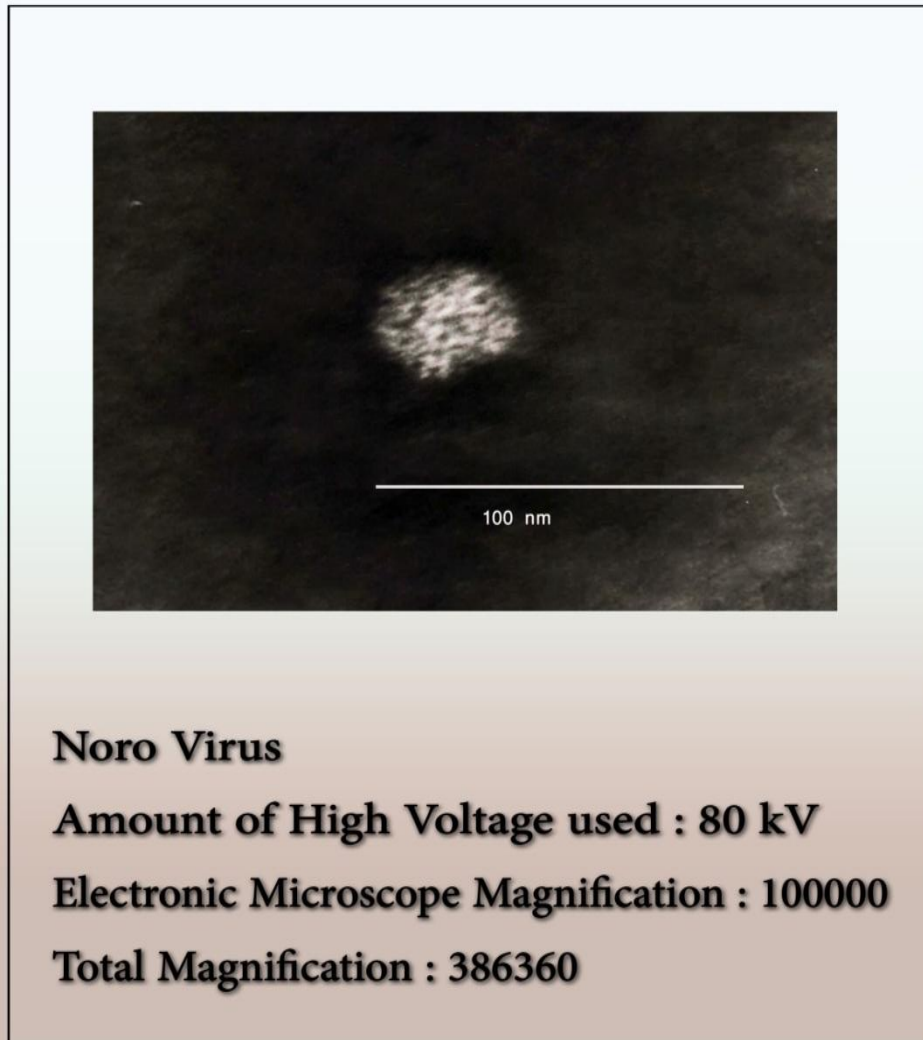
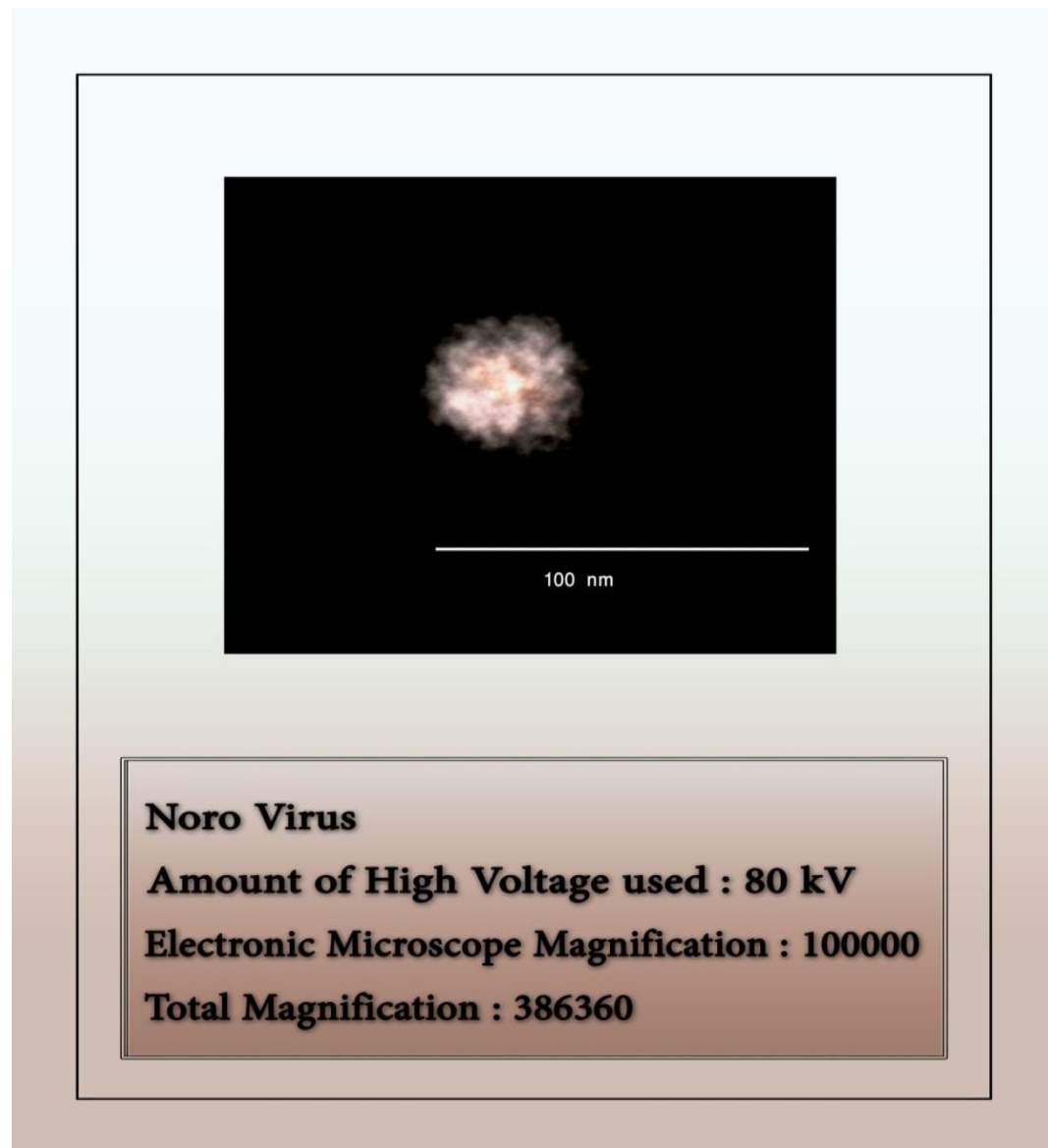


Fig. 3:- Noroviruse particles stained with negative staining under TEM.



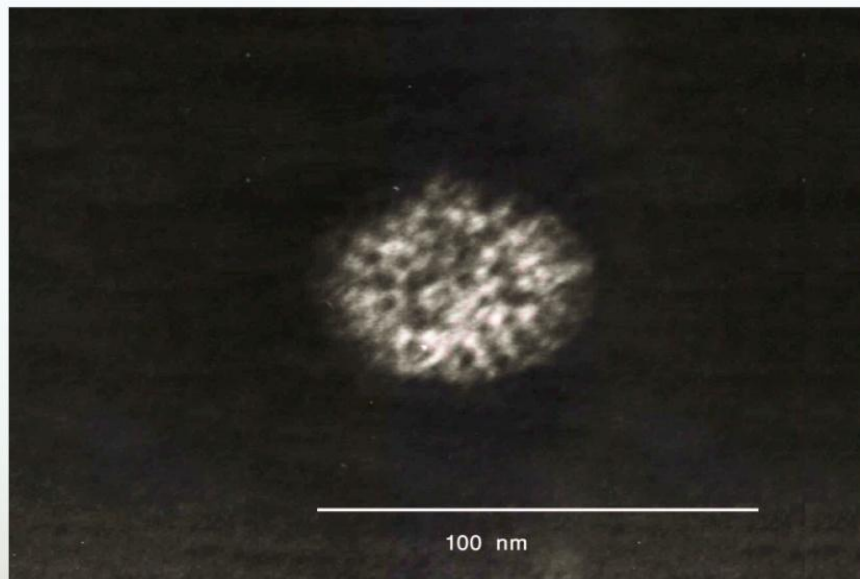
**Rota Virus****Amount of High Voltage used : 60 kV****Electronic Microscope Magnification : 100000****Total Magnification : 386360**

Fig. 4:- Rotaviruse particles stained with negative staining under TEM.

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