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RESEARCH ARTICLE

The incidence of Norovirus compared with Rotavirus in Baghdad City

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Norovirus are the commonest cause of outbreaks of non-bacterial acute gastroenteritis, and it is the most commonly recognized food borne viral infection and second only to *Rotavirus* as a cause of severe diarrhea in children. This study describes, for the first time incidence of *Norovirus* in Baghdad Between May 2012 and May 2013, 252 stool samples were collected from children 31 (12.30%) cases of males and 27 (10.71%) cases of females) in the age group under 5 years from Baghdad Hospitals. *Norovirus* genogroups detected in fecal specimens using real time reverse transcription PCR. 81 samples were identified as *Norovirus*, and 62 as *Rotavirus*. Recurrence of *Norovirus* in males were more than in females. Study appeared high incidence *Norovirus* during August 66.6%, Mars 55%, June 50% and January 40.9% and June, and dominated NV GII genogroup on VN GI genogroup. *Norovirus* NVGI and NVGII co-infections was represented (3.5%) and *Rotavirus*–*Norovirus* co-infection was represented (3.5%) of all cases, with more severe symptoms and long duration in *Norovirus*–*Rotavirus* co-infection.

Noroviruses are the commonest cause of

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Introduction

Noroviruses (NoVs) are not only the leading causative agents of outbreaks of acute viral gastroenteritis worldwide in people of all ages, but also the second most common viral etiological agents of severe childhood gastroenteritis after *Rotavirus* (Patel *et al.*, 2008; Glass *et al.*, 2009), and the principle cause of foodborne illness in Europe and the United States (Kroneman *et al.*, 2009; Philips *et al.*, 2010). The number of estimated cases in the United States was recently revised to 5.5 million annually (Scallan *et al.*, 2011), while England has an estimated 2 million cases per year (Philips *et al.*, 2010). Children aged under 5 years residing in resource-rich countries, NoVs cause ~900 000 episodes of gastroenteritis necessitating a clinic visit, compared with resource poor countries in which NoVs may cause more than one million hospitalizations and up to 200 000 deaths each year (Patel *et al.*, 2008). As enteric viruses, the NoVs are spread via the feces or vomitus of infected individuals. *Norovirus* illness is contracted through contaminated food and water and direct person-to-person transmission. Contamination often arises during the handling and preparation of foods (Jiang *et al.* 1990). *Norovirus*, belonging to the family of Caliciviridae, NoVs are non-enveloped viruses with a single-stranded, positive-sense, polyadenylated RNA genome of about 7500 nucleotides (nt) in length. Three overlapping ORFs encode the nonstructural (ORF1) and structural (ORF2 and ORF3) viral Proteins (Green *et al.*, 2000). NoV in nature is genetically and antigenically highly diverse (Kageyama *et al.*, 2004). NoV is tentatively divided into five genogroups (GI to GV). Most NoVs infecting humans belong to genogroups GI and GII. GI genogroup is further subdivided into at least eight genotypes and GII genogroup into 17 genotypes. Is a highly contagious virus and has been found to be one of the most important causes of nonbacterial acute gastroenteritis in all ages in developing as well as in developed countries (Jiang *et al.*, 1990; Lopmann *et al.*, 2004). While most of the outbreaks are known to have a seasonal pattern, sporadic cases of disease throughout the year are described (Pianget *et al.*, 2000). Specific diagnosis of *Norovirus* is routinely made by

polymerase chain reaction (PCR) assays or real-time PCR assays, which give results within a few hours. These assays are very sensitive and can detect as few as 10 virus particles (Marshall and Bruggink 2006).

Material and Methods

Sample collection

Stool samples were collected from children under 5 years with clinical symptoms of non-bacterial or parasitic acute gastroenteritis: nausea, vomiting and/or three or more loose stools in 24 hours during the acute phase of the infection in sterile plastic water proof container labeled with patient name, patient number, hospital, and date of collection Figure 1; Samples were transported to the laboratory on ice in sealed bag stored at +4°C in refrigerator until processing. After examination, samples were stored at -20°C. (Hansman, 2007; Anbazhagi *et al.*, 2011).



Figure 1. Sample collection

RNA Extraction

For viral RNA extraction 30% (w/v) stool suspensions were made in phosphate-buffered saline (PBS; 7.2pH). Extractions were performed using the QIAamp Viral RNA Mini kit (Qiagen, Germany) according to the manufacturer's instructions.

Real-time quantitative reverse transcription-PCR (RT-PCR) (Lazaro, 2010).

Viral RNA was extracted from 140 μ L of supernatant with a QIAamp viral RNA mini kit (Qiagen, Germany). Real-time quantitative RT-PCR was performed using Go Tag 1 step RT-qPCR reaction kit from (Promega). Three sets of specific primers, QNINF4, NV1LCR, and NVGG1P probe were used to detect NV GI and QNIF2, COG2R and QNIFS probe were used to detect NVGII, respectively with using IACP (Applied Biosystem UK). Real-time RT-PCR was performed with an ABI Prism 7500 sequence detector (Applied Biosystem) under the following conditions: (1) Reverse transcription $\geq 37^{\circ}\text{C}$ for 15 min, (2) Hot start activation 95°C for 10 min, (3) steps qPCR 40 cycles of amplification with denaturation at 95°C for 10 sec, annealing at 60°C for 30 sec and extension at 72°C for 30 sec.

ROTA-VIRUS LATEX TEST KIT

It is a rapid latex agglutination assay for the detection of Rotavirus in fecal samples. Stool suspensions were prepared as a 10% suspension of the fecal sample by adding 0.1 ml/0.1 g of sample to 1 ml of extraction buffer in a screw capped vial. Mix well. Stand at room temperature for 2 minutes, then Examine according to the manufacturer's instructions.

Results

Table 1. The incidence of human Norovirus and Rotavirus in five age groups (%) from 252 samples of different gender.

Age groups year	NV GI Positive sample		NV GII positive sample		Rotavirus positive sample		Negative sample	
	♂(%)	♀(%)	♂(%)	♀(%)	♂(%)	♀(%)	♂(%)	♀(%)
0-≤1	9 (69.23)	7 (70.0)	22 (70.96)	15 (55.56)	21 (67.74)	20 (64.52)	36 (59.01)	26 (54.16)
1-≤2	4 (30.76)	2 (20.0)	2 (6.45)	7 (25.93)	6 (19.36)	9 (29.03)	10 (16.39)	10 (20.01)
2-≤3	0 (0.00)	1 (10.0)	3 (9.67)	4 (14.81)	0 (0.00)	1 (3.23)	7 (11.47)	2 (4.16)
3-≤4	0 (0.00)	0 (0.00)	1 (3.23)	1 (3.70)	1 (3.23)	1 (3.23)	2 (3.27)	6 (12.5)
4-≤5	0 (0.00)	0 (0.00)	3 (9.67)	0 (0.00)	3 (9.68)	0 (0.00)	8 (13.11)	4 (8.33)
Chi-square	11.54 **	10.94 **	10.78 **	8.63 **	10.03 **	9.64 **	8.36 **	8.12 **
Total	13 (5.1)	10 (3.96)	31 (12.30)	27 (10.71)	31 (12.3)	31 (12.3)	61 (24.2)	48 (19.04)
252	23		58		62		109	

** (P<0.01).

Table 2. The number or Rotavirus and Norovirus positive samples and co-infection between May 2012 and May 2013 in Baghdad (%).

Month	No of Rotavirus Positive Sample (%)	No of Norovirus Positive Sample (%)	No of Norovirus Rotavirus co- infection (%)	No of negative cases (%)	No of total cases	Chi-square value
May	2 (10.5)	2 (10.5)	0 (0)	15 (78.9)	19	4.051 *
June	5 (20)	12 (50)	0 (0)	8 (33.3)	24	10.54 **
July	9 (45)	4 (40)	0 (0)	8 (40)	20	10.82 **
August	6 (28.57)	14 (66.6)	2 (9.5)	4 (19)	21	9.46 **
September	5 (23.8)	1 (4.8)	0 (0)	15 (71.4)	21	6.84 **
October	7 (30.4)	5 (21)	2 (8.7)	13 (56.5)	23	7.72 **
November	3 (13.6)	3 (13.6)	0 (0)	17 (77.3)	22	3.68 *
December	4 (20)	7 (35)	0 (0)	9 (45)	20	7.91 **
January	7 (31.8)	9 (40.9)	4 (17.4)	12 (52.2)	23	8.52 **
February	5 (26.3)	5 (26.3)	0 (0)	10 (52)	19	7.39 **
March	4 (20)	11 (55)	1 (5)	8 (40)	20	9.43 **
April	5 (25)	8 (40)	2 (10)	10 (50)	20	9.18 **
Total	62 (24.60)	81 (32.14)	11 (4.36)	129 (51.19)	252	

** (P<0.01).

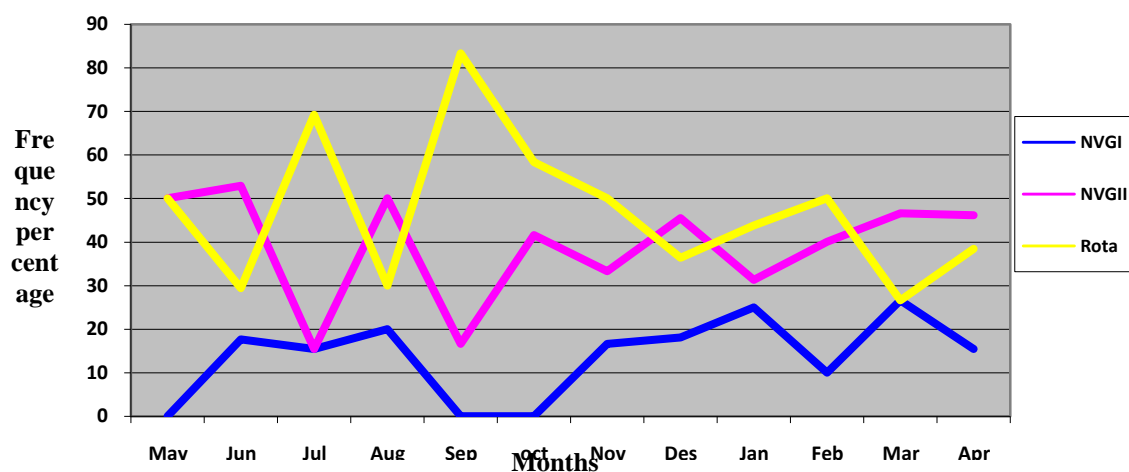


Figure 2. The Frequency percentage of Norovirus GI, Norovirus GII, and Rotavirus positive samples between May 2012 and May 2013

Table 3 .The number of *Norovirus GI* ,*Norovirus GII* ,and Co-infection positive samples between May 2012 and May 2013 (%).

Month	No of Norovirus G1 positive Sample(%)	No of Norovirus G2 positive Sample(%)	No of Co- infection NVG1& NVG2(%)	No of total cases	Chi-square value
May	0 (0)	2 (10.5)	0 (0)	19	12.56 **
June	3 (12.5)	9 (37.5)	1 (4.2)	24	8.63 **
July	2 (10)	2 (10)	1 (50)	20	9.12 **
August	4 (19)	10 (47.6)	2 (9.5)	21	9.05 **
September	0 (0)	1 (4.8)	0 (0)	21	11.38 **
October	0 (0)	5 (21.7).	0 (0)	23	9.62 **
November	1 (4.5)	2 (9.1)	0 (0)	22	12.85 **
December	2 (10)	5 (25)	0 (0)	20	9.63 **
January	4 (17.4)	5 (21.7)	1 (4.3)	23	9.24 **
February	1 (5.3)	4 (21)	1 (5.2)	19	10.63 **
March	4 (20)	7 (35)	2 (10)	20	9.21 **
April	2 (10)	6 (30)	1 (5)	20	10.78 **
Total	23 (9.13)	58 (23)	9 (3.57)	252	10.32 **

** (P<0.01).

Table 4. The Gastrointestinal symptoms in Norovirus in 81 positive cases

Gastrointestinal symptoms	Percentage (%)
Abdominal pain	83.93
Vomiting	91.07
Diarrhea	98.21
Severe dehydration	1.80
Some dehydration	33.90
No dehydration	46.42
Nausea	51.80
Normal temperature	80.30
Mild temperature	16.07
fever	3.57
Chi-square value	13.287 **

** (P<0.01).

Table 5. The co-infection and mono-infection in relation to duration ,vomiting and diarrhea episodes among 143 Rotavirus and Norovirus positive cases

Disease Manifestation	Norovirus Mono-infection	Rotavirus Mono-infection	NVGI& NVGII mix- infection	NV & Rotavirus Mix-infection
Duration				
<4 day	50 (81.96)	20 (39.21)	6 (66.6)	1 (9.09)
4-7 day	10 (16.39)	27 (52.94)	2 (22.2)	4 (36.36)
<7 day	1 (1.63)	4 (7.84)	1 (11.1)	6 (54.54)
Chi-square	13.58 **	9.43 **	10.45 **	9.27 **
Vomiting episode/day				
<3/d	22 (36.06)	14 (27.45)	2 (22.22)	1 (9.09)
3-7/d	31 (50.81)	22 (43.13)	2 (22.22)	3 (27.27)
>7/d	8 (13.11)	15 (29.41)	5 (55.56)	7 (63.63)
Chi-square	9.63 **	11.49 **	9.54 **	11.43 **
Diarrhea episode/day				
<3/d	0 (0)	0 (0)	0 (0)	0 (0)
3-7/d	39 (63.93)	35 (68.62)	4 (44.44)	4 (36.36)
>7/d	22 (36.06)	16 (31.37)	5 (55.55)	7 (63.63)
Chi-square	11.67 **	11.75 **	10.43 **	11.94 **
Total	61	51	9	11

** (P<0.01).

Discussion

Through this study, which investigated children under five years with acute gastroenteritis (AGE) attending hospitals in Baghdad. A number of 252 samples were collected and information was extracted from patient using Diarrhea case report form. All samples were Examined using RT-PCR (the genomic region utilized to detect and genotype HuCV by RT-PCR codify to the polymerase gene (RdRp)), which is relatively conserved in both genera. Table (3.1) shows that the overall number of males was higher than female children in AGE by 136 (53.9%) males and 116 (46.03%) females. Also the result revealed a higher incidence of Norovirus infection in males compared to females. As we have noted the infection with genogroup NVGI was affecting 13 males (5.1%) of AGE and effecting only 10 females (3.96%) of AGE, and the occurrence of infection with genogroup NVGII was effecting 31 males (12.30%) of AGE in comparison to only 27 females (10.71%) of AGE. While the study found through equal readiness of Rotavirus infection between males and females and by 31 (12.30%) cases for both of them with a high significant value as shown in Table (1). The recurrence Norovirus infection under one year and in males were more than in females. The result showed increase in Norovirus infection for the same age group and superiority over Rotavirus. The lower incidence in older ages for both viruses, is probably due to their past infection in early childhood which might earn immunity against infection in the future and also using *Rotavirus* vaccine.

These results were expected by Gómez-Santiago *et al.* (2012) in Brazil and with Mandellet *et al.* (2010) which refers to the age group most affected with HuCV infection (7-18 months) corresponded to the group most commonly affected by *Rotavirus*, with a median age of 12 months. Throughout the study appeared emergence of NVG infection during all months throughout the different seasons, (Table 2). This correspond with Beeket *et al.* (2013) who get the same pattern throughout the year. Also this study showed a high incidence of NVG during August 66.6%, March (55%), June (50%) and January (40.9%), and dominated NV GGI genogroup on NVGGI genogroup. These results was correspond with Chan and Chan (2013) in Hong Kong, throughout 2012, refers to *Norovirus* activity peaked in August and subsided in October, in addition to Glass and Parashar (2009) also observed a higher incidence in the summer months compared to the winter months. Our results was not correspond with studies in many of countries indicate that epidemics of NVG outbreaks tend to occur each year with a peak in colder months in northern hemisphere countries as observed by Goller and Dimiriadis (2004), and a peak in warmer months in southern hemisphere countries, such as Australia as mentioned by Lazaro (2010) and Bruggink and marshal (2009). Also Gómez-Santiago *et al.* (2012) was refers to the distribution of human caliciviruses (HuCV) infection occurred mainly during two periods: November-December, and April-June, in contrast with Brazil, where infection has been reported along the entire year without a seasonal pattern (Soares *et al.*, 2007). But in Chennai city of southern India, Anbazhagiet *et al.* (2011) referred in their study to that prominent numbers of NoV infection was observed on April 2006 followed by January 2006, whereas *Norovirus* is generally referred to as "winter-vomiting disease", In comparison to this study (in Iraq) a contrary picture observed as we see that is high in the summer season especially during August. On the other hand Rotavirus infection spread throughout the year and a significant increase in the July (45%) in the summer months and in January (31%) in the winter months as can be noted in (Figure 2). The results demonstrates a high prevalence of NoV infections in neonates. Mixed infections and their distribution according to months are shown in Tables 2, and 3. *Norovirus* and *Rotavirus* co-infection was seen in 11 cases presented 17.4%, 10%, 9.5%, 8.7%, and 5% of positive cases in January, April, August, October, and March respectively. This indicates that co-infection with *Rotavirus* may result in significantly more gastrointestinal disease in this age group. NVGI, and NVGII co-infection were found in 9 cases presented 3.5% of all collected samples and appear through highly prevalence months in January, February, March, April, June, July, and August. Mixed infections were predominantly detected in the under 1 years age group and this is consistent with observations seen by Romanet *et al.* (2003) who explains that virus-virus co-infections were significantly more frequent in 7-18 months age group. Also the majority of these co-infection were combinations of *Rotavirus* with other viruses (*Astrovirus*, *Adenovirus* or *Calicivirus*). Detection of more than two infectious agents was infrequent. As it has been noted in this study that mixed infections are less frequent than mono-infections but with more severe and prolong duration time, and this correspond with the result done by Pratset *et al.* (1997) in their study In Barcelona, Spain. The Statistical comparison of single virus infection with double infections is complex because most of the co-infection groups are small. (Table 5), it has been shown that the most important exhibitors of patients were vomiting, and diarrhea, a high proportion 91.01%, 98.21% respectively. A high rate of patient 83.93% were suffering from abdominal pain and moderate nausea 51.8%. Also severe dehydration and fever were occurred only in 1.80% and 3.57% of patients on the other hand most patient have normal body temperature and very low

rate of dehydration in children and these finding consistent with Graham *et al.* (1994) results who explained that Norovirus can be detected in infected individuals with no major symptoms.

The symptoms between the mono-infection groups and mixed infection group were compared (Table 5), and found that rotavirus caused a higher frequency and longer duration of vomiting than did Norovirus. This can explain *Norovirus* RNA excretion can continue for long periods for up to 44.5 days following onset of symptoms in a naturally infected individual as mentioned by Menon *et al.* (2010). This may explain the causes behind the mixed infection with Rotavirus in asymptomatic patients in this study.

Reference

1. Anbazhagi, S. Kamatchiammal, S. and Jayakar Santhosh, D. (2011). Norovirus based viral gastroenteritis in Chennai city of southern India - An epidemiological study. *Journal of General and Molecular Virology*; Vol. 3(2), pp. 27-34.
2. Bruggink, L. D., and J. A. Marshall. (2009). Norovirus epidemics are linked to two distinct sets of controlling factors. *Int. J. Infect. Dis.* 13:e125–e126.
3. Glass RI, Parashar UD, Estes MK. (2009). Norovirus gastroenteritis. *New England Journal of Medicine*; 361:1776–1785.
4. Goller, J.L.; Dimitriadis, A.; Tan, A.; Kelly, H.; Marshall, J.A. (2004). Long-term features of *Norovirus* gastroenteritis in the elderly. *J. Hosp. Infect.* 58, 286-291.
5. Graham, D.Y.; Jiang, X.; Tanaka, T.; Opekun, A.R.; Madore, H.P.; Estes, M.K. (1994) Norwalk virus infection of volunteers: New insights based on improved assays. *J. Infect. Dis.* 170, 34-43.
6. Green KY, Ando T, Balayan MS, Berke T, Clarke IN, Estes MK, Matson DO, Nakata S, Neill JD, Studdert MJ, Thiel HJ. (2000). Taxonomy of the caliciviruses. *J Infect*, 181:S322-330.
7. Hansman. (2007). Norovirus infections in symptomatic and asymptomatic food handlers in Japan. *J. Clin. Microbiol.* 45:3996–4005.
8. Jiang X, Graham DY, Wang KN, Estes MK (1990). Norwalk virus genome cloning and characterization. *Science*, 250:1580-1583.
9. Kageyama, T., M. Shinohara, K. Uchida, S. Fukushi, F. B. Hoshino, S. Kojima, R. Takai, T. Oka, N. Takeda, and K. Katayama. (2004). Coexistence of multiple genotypes, including newly identified genotypes, in outbreaks of gastroenteritis due to Norovirus in Japan. *J. Clin. Microbiol.* 42:2988–2995.
10. Kroneman, A., Verhoef, L., Harris, J., Vennema, H., Duzier, E., van Duynhoven, Y., et al. (2008). Analysis of integrated virological and epidemiological reports of norovirus outbreaks collected within the foodborne viruses in Europe Network from 1 July 2001 to 30 June 2006. *Journal of Clinical Microbiology*, 46, 2959-2965.
11. Lazaro, D.R. (2010). Detection and quantification of Norovirus by real time PCR. SOP Vital 018.
12. Lopmann B, Vennema H, Kohli E, Pothier P, Sanchez A, Negro A, Buesa J, Schreier E, Reacher M, Brown D, Gray J, Iturriza M, Gallimore C, Bottiger B, Hedlund KO, Torvén M, von Bonsdorff CH, Maunula L, Poljsak-Prijatelj M, Zimsek J, Reuter G, Szűcs G, Melegh B, Svensson L, van Duynhoven Y, Koopmans M. (2004) Increase in viral gastroenteritis outbreaks in Europe and epidemic spread of new norovirus variant. *Lancet*. 363:682-688.
13. Maguire AJ, Green J, Brown DWG, Desselberger U, Gray JJ (1999). Molecular epidemiology of outbreaks of gastroenteritis associated with small round-structured viruses in East Anglia, United Kingdom during the 1996 – 1997 season. *J Clin Microbiol* 1999, 37:81-89.

- 14.Mandell A B., Gerald L.; Bennett, John E.; Dolin, Raphael (2004). Mandell's Principles and Practices of Infection Diseases (6th ed.). Churchill Livingstone. ISBN 0-443-06643-4 .
- 15.Marshall, J. A., A. Dimitriadis, and P. J. Wright. (2005). Molecular and epidemiological features of norovirus-associated gastroenteritis outbreaks in Victoria, Australia in 2001. *J. Med. Virol.* **75**:321–33116.
- 16.Marshall JA, Bruggink LD (2006). "Laboratory diagnosis of norovirus". *Clin. Lab.* 52 (11–12): 571–81. PMID 17175887.
- 17.Menon, V.K., George,S., Ramani,S., Illiayaraja,J., Sarkar,R.,Jana,A.K., Kuruvilla.A.K., Kang.G.(2010) Genogroup IIb Norovirus Infections and Association with Enteric Symptoms in a Neonatal Nursery in Southern India *Journal of Clinical Microbiology*. p. 3212–3215 Vol. 48, No. 9.
- 18.Mermelstein, N.H.(January 2013). Targeting Norovirus. *Food technology* ., Volume 67, No.1
- 19.Ozawa, K., T. Oka, N. Takeda, and G. S.(Aug. 2010). Diagnostic Accuracy and Analytical Sensitivity of IDEIA Norovirus Assay for Routine Screening of Human Norovirus, *Journal of Clinical Microbiology* p. 2770–2778 Vol. 48, No. 8.
- 20.Patel, M.M. Widdowson,M.A Glass,G.I. Akazawa,K Vinjé,J.and. Parashar,U.D.(2008). Systematic literature review of role of noroviruses in sporadic gastroenteritis. *Emerging Infectious Diseases*; 14: 1224–1231.
- 21.Phillips, G., Tam, C. C., Conti, S., Rodrigues, L. C., Brown, D.,Iturriza-Gomara, M., et al. (2010). Community incidence of Noroviruses-associated infectious intestinal disease in England: Improved estimates using viral load for Noroviruses diagnostics. *American Journal of Epidemiology*, 171, 1014–10221.
- 22.Piang XL, Honma S, Nakata S, Vesikari T.(2000).Human Calicivirus in acute gastroenteritis of young children in the community. *J Infect*, 181:288-294.
- 23.Roman,E. Wilhelmi,I. Colomina,J. Villar,J. Luz Cilleruelo,M. Nebreda,V. Del Alamo,M.and, Sa´nchez-Fauquier,A. (2003).Acute viral gastroenteritis: proportion and clinical relevance of multiple infections in Spanish children *Journal of Medical Microbiology* ; 52, 435–440.
- 24.Tu, E.T.V.; Bull, R.A.; Kim, M.J.; McIver, C.J.; Heron, L.; Rawlinson, W.D.; White, P.A. (2008).Norovirus excretion in an aged-care setting. *J. Clin. Microbiol.* **46**, 2119-2121.
- 25.Scallan, E., Hoekstra, R. M., Angulo, F. J., &Tauxe, R. V. (2011).Foodborne illness acquired in the United States—major pathogens. *Emerging Infectious Diseases*, 17, 7–15.
- 26.Souares, C.C., Santos ,N., Beard, R.S. Albuquerque, M.C. Maranhão, A.G., Rocha, L.N., Ramírez, M.L. Monroe ,S.S. Glass R,I. Gentsch ,J(2007).: Norovirus detection and genotyping for children with gastroenteritis, Brazil. *Emerg Infect*, 13:1244-1246.
- 27.Wilhelmi de Cal I, Revilla A, del Alamo JM, Román E, Moreno S,Sánchez-Fauquier A. (2007). "Evaluation of two commercial enzyme immunoassays for the detection of norovirus in faecal samples from hospitalised children with sporadic acute gastroenteritis". *Clin. Microbiol. Infect.* 13 (3): 341–3. doi:10.1111/j.1469-0691.2006.01594.x. PMID 17391396.