

RESEARCH ARTICLE

CONTAMINATION OF THE SHELLS OF LEBANESE FREE RANGE CHICKEN EGGS WITH POTENTIAL PATHOGENS.

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Manuscript Info	Abstract		
Manuscript History	Local free-range eggs in Lebanon are favored by consumers, but are often sold without prior cleaning or processing. Eggs were proved to		
Received: 04 april 2017	be a major source of many enteric pathogens that can cause varying		
Final Accepted: 06 june 2017 Published: june 2017	episodes of gastroenteritis, this study was designed to investigate the surface contamination of free range egg shells. The tested eggs were purchased from 11 stores located in different regions of Lebanon. The		
Key words:- Enterobacteriaceae, Fecal contamination, Free range-chicken eggs, Lebanese poultry market	results revealed that many members of the family <i>Enterobacteriaceae</i> , that are present in chicken feces, including <i>Escherichia coli</i> , <i>Enterobacter cloacae</i> and others, were isolated, however it was noted that <i>Salmonella</i> spp. were not among the isolates. The presence of these organisms, in the eggs obtained from different areas, confirmed that there was a trend not to clean or process these eggs in any way before sending them to the stores for sale. The mere isolation of fecal		
	organisms, indicated that the eggs can, at any time, be contaminated with <i>Salmonella</i> spp. and other dangerous enteric bacteria, and thus endanger the health of the customers. It is required that public health		

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protective measures, be enforced immediately to protect the

consumers from what is currently an obvious health hazard.

Introduction:-

Birds have a common excretory and reproductive system. As such, it is known that chicken egg shells are often contaminated with a wide spectrum of different enteric Gram positive and Gram negative bacteria. Thus, fecal contamination of eggs results in the presence of many members of the family *Enterobacteriaceae*, including potentially pathogenic species such as *Salmonella spp.* and *Escherichia spp.* (Gole et al., 2013). The contamination of eggs with *Salmonella* Enteritidis (*Salmonella enterica*, subspecies *enterica*, serovar Enteritidis), has been of major concern, being the leading cause of Salmonellosis in both

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humans and animals in the USA, Europe and many other countries around the world including Lebanon (JAMA, 1992; Mishu et al., 1994; Barbour et al., 1999; Poppe, 1999; WHO, 2000; FDA, 2016a). SE infections can be very serious, even life-threatening, especially to the very young, the elderly, and people with weakened immune systems. Infected people may experience general enteric symptoms, but some infected people may suffer from severe illness, arthritis, or even death (FDA, 2016a). Eggs can become contaminated on the farm because a laying hen can become infected with SE and pass the bacteria into the egg before it is laid. If the egg was not refrigerated, the bacteria can grow inside the uncracked, whole egg (FDA, 2016a). In fact, *Salmonella spp.* such as SE, have shown the ability to

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penetrate egg shells through pores on their surface and enter to the egg yolk where they can multiply significantly faster (Gast et al., 2005). For this reason, the FDA imposed a set of strict regulations on egg producers to prevent and reduce SE infections (FDA, 2017). Today, modern commercial processing successfully decreased microbial contamination of egg shells in many countries all over the globe (Musgrove et al., 2005).

In Lebanon, some stores proudly advertise that they sell local free-range eggs, transported "fresh" from a farm. These eggs are perceived by the public as being healthier and less subject to the chemicals used during commercial egg production, and are thus desired by a large segment of the Lebanese society. Since there are no standard washing or processing procedures performed by the farmers, these eggs are often not cleaned and present a higher risk of contamination by numerous members of the family *Enterobacteriaceae* (Hannah et al., 2011). The estimated incidence of Salmonellosis in Lebanon is reported at 13.34 cases/100,000 individuals mostly in the age group of 20-39 years old (Malaeb et al., 2016). The incidence is, however, believed to be higher for many reasons including: the general symptoms in most infection (diarrhea, fever, abdominal cramps, headache, nausea and vomiting) that, often, go unreported, in addition to, the lack of regular reporting, especially for the less developed areas and the absence of a central office that compiles the information (Malaeb et al., 2016). *Salmonella* infections in Lebanon have been, however, noted to become increasingly resistant to antibiotics (Moubareck et al., 2005). This study was designed to gather information about the contamination of the surfaces of the shells of free-range chicken eggs, obtained from stores located in different areas of Lebanon, with members of the family *Enterobacteriaceae* and specifically *Salmonella* spp.

Materials and Methods:-

Samples tested:-

From 11 different stores located in different parts of Lebanon (Figure 1), 2 "free-range" and "fresh" eggs were purchased from each. The sample eggs were purchased between January and March, 2017, from the stores and not the producers themselves, to ensure that the eggs were received just as a regular consumer would. The "freshest" eggs were chosen and transported to the microbiology laboratory at room temperature and in the shortest period of time not exceeding 24 hours. The eggs were transported to the laboratory in the same bag that they were packed in, simulating what a consumer would actually take home.



Figure 1:- A map of Lebanon, showing the sites from where the samples, in this study, were purchased

Processing of samples:-

Upon arrival to the microbiology laboratory, separate sterile swabs were dipped in Selenite Cystine Broth (SCB) and used to swab the entire shell surface of each egg. Each of the swabs was used to inoculate: Tryptic Soy Agar (TSA), MacConkey Agar (MCA), and Salmonella-Shigella Agar (SSA) plates in that order. Following inoculation, the swabs were re-inserted into the tubes and the plates were properly streaked. All plates and tubes were then incubated at 35°C for 20-24 hours. After the incubation period, the surfaces of the plates were checked for bacterial growth and all colonies that appeared on the different media were isolated, checked for purity and preserved on TSA slants at 4°C for the next step of identification. The SCB tubes were further used to inoculate the same set of plates again. The new plates were incubated for 20-24 hours at 35°C. The purpose of using SC broth was to inhibit the growth of Gram positive bacteria and aid in the recovery of *Salmonella* spp. and/or *Shigella* spp. in case they were present in small amounts (Hammack et al., 1999).

Identification of isolates:-

After Gram staining, as only Gram-negative bacilli were sought, the following preliminary tests were performed on each isolate: Oxidase production using oxidase test strips and Glucose and lactose fermentation and gas and H2S production using Kligler's Iron Agar (KIA) done as recommended (Jorgensen and Pfaller, 2015). Definitive identification of the isolates, however, was done using the API 20E kits (Biomerieux-France) as recommended by the manufacturer.

Results:-

Table 1, shows that, except for the eggs purchased from two stores in Zahle and coastal Akkar (numbered 7 and 10), the eggs from the other locations showed positive bacterial growth. Of the 18 tested eggs, 16 grew Gram negative bacilli, 15 of which were members of the family *Enterobacteriaceae* while the last was a non-fermenter from the family *Moraxellaceae*.

Sample Number	Source Location	Egg	Growth on study	Organisms isolated
1	Maidalyoun	1	ilicula	
1	Majueryoun	2	-	-
-		2	+	Enterobacier cloacae
2	Bchamoun	1	+	Escherichia coli
		2	+	Pantoea sp.
3	Kfarshima	1	+	Citrobacter freundii
		2	+	Citrobacter braakii
4	Hamra	1	-	-
		2	+	Pantoea sp.
5	Jdaide	1	+	Escherichia coli
		2	+	Citrobacter freundii
6	Mazraat Yachouh	1	+	Enterobacter cloacae
		2	+	Enterobacter cloacae
7	Zahle	1	-	-
		2	-	-
8	Koura	1	+	Citrobacter freundii
		2	+	Pantoea sp.
9	Tripoli	1	+	Enterobacter cloacae
		2	+	Enterobacter cloacae
10	Akkar (coast)	1	-	-
		2	-	-
11	Akkar (inland)	1	+	<i>Moraxella</i> sp.
		2	+	Escherichia coli

Table 1:-The isolates from the shells of the eggs; reported by location of stores they were purchased from.

The frequencies of the *Enterobacteriaceae* isolates from the 15 eggs were: *Enterobacter cloacae* (33.3%), *Escherichia coli, Pantoea* spp. *Citrobacter freundii* (20 % each) and *Citrobacter braakii* (6.7 %). No *Salmonella* spp. were isolated from any of the egg shells.

Discussion and Conclusion:-

Checking the surface contamination of the shells of the free-range eggs tested in this study, was done using media that usually enhances the growth of Gram negative bacteria. The broth used to collect the samples from the surface of the shells was selenite cysteine broth, as the cystine formulation was first proposed by the US Food and Drug Administration (FDA), as an enrichment medium for detecting *Salmonella* in food materials. It was proved useful in detecting *Salmonella* when low numbers of organisms were present in stools (Jorgensen and Pfaller, 2015), and it was also recommended for use in detecting *Salmonella* in food and water (Speck, 1984; Greenberg, 1985). The sodium selenite in the medium is an inhibitor of Gram positive bacteria.

No Salmonella spp. were isolated from the samples in this study, for reasons, that for the investigators, were obvious and will be discussed later. The results in Table 1, however, clearly show that the organisms isolated indicate that the surfaces of the eggs were contaminated with fecal material. Moreover, all the isolated organisms are described to be potential pathogens. They may not be primary pathogens, but pose a higher risk of infection for the young, elderly and immunocompromised members of the community. Enterobacter and Pantoea species for instance, were reported to cause several hospital-acquired infections and were noted to have become increasingly multi-resistant (Davin-Regli and Pagã"s, 2015; Dutkiewiez et al., 2016). E. coli is also known to cause infection upon ingestion, even in small numbers (Mayo Clinic, 2014). Shiga toxin-producing E. coli strains which were detected in living hens (CDC, 2015), and the dangerous O157:H7 strain were linked to many outbreaks of gastrointestinal infections (Dipineto et al. 2006). Moreover, it was also proved that E. coli is capable of facilitating the penetration of Staphylococcus aureus into the egg content (Al-Natour et al., 2011), another reason for the increased interest in controlling fecal contamination, of the surface of egg shells. Although the Citrobacter isolates are generally not considered as primary pathogens as some of the other isolates in the study, yet some of the members of this genus, have been reported to cause urinary tract infections, upper respiratory tract infections (Mossad, 2013) otitis media in children (Doran, 1999) and infant meningitis (Samonis et al., 2008), but mostly in patients who have at least 1 underlying illness (Mossad, 2013). The isolate from the genus Moraxella belongs to a genus that is also known to have many members that act as opportunistic pathogens that cause many infections including upper respiratory tract infections (Bernhard, 2012).

The WHO predicts that opportunistic infections will become more dangerous in the 21^{st} century, further increasing the importance of restricting the spread of the bacteria that cause these infections. What adds to the problem, is the fact that most members of the family *Enterobacteriaceae* have become increasingly multi-resistant to antibiotics, as seen through the emergence of the Extended Spectrum β -Lactamase producing (ESBL) bacteria all over the world and the isolation of the first ESBL *Salmonella* Typhimurium in Lebanon in 2006 (Moubareck et al., 2005), and around the world (Mishu et al., 1994; WHO, 2000).

The absence of *Salmonella* spp. from the isolates is instigated by many factors. It is a known for a fact that *Salmonella* spp. are much more sensitive to external conditions than most other fecal organisms, and their survival is influenced by several different factors (Kim et al., 1989; Garcia et al., 2010; CDC, 2016). The samples tested in this study were collected in the winter (between January and March) from the different regions in Lebanon, which are known to have different weather conditions. The request from the merchant to provide the investigators with the "freshest free range eggs" in the market, may not have been respected and the samples may have been older eggs that were present in the store. Another factor is the long distances that these eggs had to be transported through, from the farm to the market and later to the laboratory, without refrigeration; This probably led to the loss of any *Salmonella* spp. or any other sensitive organism that may have initially been present on the shells. This is confirmed by noting that the two samples (7 and 10 in Table 1) purchased from the markets that are farthest away from the microbiology laboratory, located in the capital Beirut, did not grow any organism.

Knowing that not all chicken carry *Salmonella* as part of their microflora, the fecal contamination that was obvious on the shells of the tested eggs, however, reveals a health hazard to the consumers. Considering these findings, it is recommended that Lebanese egg producers implement the accepted international guidelines in order to reduce contamination by these potentially dangerous bacteria. The FDA (2016b, 2017), introduced a set of prevention methods and recommendations which could be easily implemented by the Lebanese producers, although there were previous recommendations to that effect (Barbour et al., 2001). Moreover, pasteurization and dry heat treatment can also be used as they were proved to be effective in decreasing SE contamination (Arnold et al., 2014), in addition to that vaccination against certain strains of *Salmonella* spp., also showed effectiveness in decreasing SE contamination on egg shells (Barker et al., 2003). However, perhaps the most important recommendation is for the

consumer, as the risk of infection by SE or any of the *Enterobacteriaceae* can be tremendously decreased by making sure that eggs are not eaten raw, as is common in the Lebanese countryside, and that they are well-cooked. Consumers must also wash all kitchen utensils, benches, and hands exposed to these eggs using proper detergents to diminish any chance of cross-contamination (Barker et al., 2003). The role of public health authorities in adopting cleaning and processing regulations and monitoring their implementation, should also be affirmed.

References:-

- 1. Al-Natour, M.Q., Alaboudi, A.R., Al-Hatamelh, N.A. and Osaili, T.M. (2011). *Escherichia coli* O157:H7 Facilitates the Penetration of Staphylococcus aureus into Table Eggs. Journal of Food Science. 77: M29–M34. doi:10.1111/j.1750-3841.2011.02483.x
- Arnold, M.E., Gosling, R.J., Ragione, R.M., Davies, R.H. and Martelli, F. (2014). Estimation of the impact of vaccination on faecal shedding and organ and egg contamination for *Salmonella* Enteritidis, *Salmonella* Typhiumurium and monophasic *Salmonella* Typhimurium. Avian Pathology. 43(2): 155-163. doi:10.1080/03079457.2014.896990
- Barbour, E K., Jurdi, L., Talhouk, R., Qatanani, M., Eid, A., Sakr, W., Bouljihad, M. and Spasojevic, R. (1999). Emergence of *Salmonella enteriditis* outbreaks in broiler chicken in the Lebanon: epidemiological markers and competitive exclusion control. Revue Scientifique et Technique de l'OIE. 18(3): 710-718. doi:10.20506/rst.18.3.1184
- 4. Barbour, E.K., Jurdi, L.E., Issa, C. and Tannous, R. (2001). Preliminary attempts towards production of table eggs free from *Salmonella enteritidis*. Journal of Cleaner Production. 9(1): 69-73. doi:10.1016/s0959-6526(00)00033-0
- Barker, J., Naeeni, M. and Bloomfield, S. (2003). The effects of cleaning and disinfection in reducing Salmonella contamination in a laboratory model kitchen. Journal of Applied Microbiology. 95(6): 1351-1360. doi:10.1046/j.1365-2672.2003.02127.x
- 6. Bernhard, S., Spaniol, V. and Aebi, C. (2012). Molecular pathogenesis of infections caused by *Moraxella catarrhalis* in children. Swiss Medical Weekly. 142, w13694. doi: 10.4414/smw.2012.13694
- 7. Centers for Disease Control and Prevention. (2015). *Escherichia coli* Fact sheet. CDC. USA. Available online at: https://www.cdc.gov/ecoli/general/index.html
- 8. Centers for Disease Control and Prevention. (2016). *Salmonella* Fact sheet. CDC. USA. Available online at: https://www.cdc.gov/salmonella/outbreaks.html
- 9. Davin-Regli, A. and Pagã^{*}s, J. (2015). *Enterobacter aerogenes* and *Enterobacter cloacae*; versatile bacterial pathogens confronting antibiotic treatment. Frontiers in Microbiology. 6: 392. doi:10.3389/fmicb.2015.00392
- Dipineto, L., Santaniello, A., Fontanella, M., Lagos, K., Fioretti, A. and Menna, L. (2006). Presence of Shiga toxin-producing *Escherichia coli* O157:H7 in living layer hens. Letters in Applied Microbiology. 43(3): 293-295. doi:10.1111/j.1472-765x.2006.01954.x
- 11. Doran, T. (1999). The Role of *Citrobacter* in Clinical Disease of Children: Review. Clinical Infectious Diseases. 28(2): 384-394. doi:10.1086/515106
- 12. Dutkiewicz, J., Mackiewicz, B., Lemieszek, M. K., Golec, M. and Milanowski, J. (2016). *Pantoea agglomerans*: a mysterious bacterium of evil and good. Part III. Deleterious effects: infections of humans, animals and plants. Annals of Agricultural and Environmental Medicine. 23(2): 197-205. doi:10.5604/12321966.1203878
- 13. Food and Drug Administration. (2016a). FDA Improves Egg Safety. U.S. Food and Drug Administration (FDA). Available online at:

https://www.fda.gov/ForConsumers/ConsumerUpdates/ucm170640.htm

- 14. Food and Drug Administration. (2016b). Eggs Guidance for Industry: Prevention of *Salmonella* Enteritidis in Shell Eggs During Production, Storage, and Transportation. Center for Food Safety and Applied Nutrition. U.S. Food and Drug Administration. Maryland. USA. Available online at: https://www.fda.gov/Food/GuidanceRegulation/GuidanceDocumentsRegulatoryInformation/Eggs/ucm285101.h tm
- 15. Food and Drug Administration. (2017). FDA. Eggs Egg Safety Final Rule. U.S. Food and Drug Administration (FDA). Available online at:
 - https://www.fda.gov/food/guidanceregulation/guidancedocuments regulatory information/eggs/ucm170615.htm
- Garcia, R., Baelum, J., Fredslund, L., Santorum, P. and Jacobsen, C. S. (2010). Influence of Temperature and Predation on Survival of *Salmonella enterica* Serovar Typhimurium and Expression of invA in Soil and Manure-Amended Soil. Applied and Environmental Microbiology. 76(15): 5025-5031. doi:10.1128/aem.00628-10

- 17. Gast, R.K., Holt, P.S. and Murase, T. (2005). Penetration of *Salmonella enteritidis* and *Salmonella heidelberg* into egg yolks in an in vitro contamination model. Poultry Science. 84(4): 621-625. doi:10.1093/ps/84.4.621
- Gole, V., Chousalkar, K. and Roberts, J. (2013). Survey of *Enterobacteriaceae* contamination of table eggs collected from layer flocks in Australia. International Journal of Food Microbiology. 164(2-3): 161-165. doi:10.1016/j.ijfoodmicro.2013.04.002
- 19. Greenberg A. (ed.). (1985). Standard Methods for the Examination of Water and Wastewater. 16th ed. APHA. Washington, D.C.
- Hammack, T.S., Amaguaña, R.M., June, G.A., Sherrod, P.S. and Andrews, W. H. (1999). Relative Effectiveness of Selenite Cystine Broth, Tetrathionate Broth, and Rappaport-Vassiliadis Medium for the Recovery of *Salmonella* spp. from Foods with a Low Microbial Load. Journal of Food Protection. 62(1): 16-21. doi:10.4315/0362-028x-62.1.16
- Hannah, J.F., Wilson, J.L., Cox, N.A., Cason, J.A., Bourassa, D.V., Musgrove, M.T., Richardson, L.J., Rigsby, L.L. and Buhr, R.J. (2011). Comparison of shell bacteria from unwashed and washed table eggs harvested from caged laying hens and cage-free floor-housed laying hens. Poultry Science, 90(7): 1586-1593. doi: 10.3382/ps.2010-01115
- 22. JAMA. (1992). Outbreak of *Salmonella enteritidis* Infection Associated With Consumption of Raw Shell Eggs. The Journal of the American Medical Association. 267(24): 3263-3264. doi:10.1001/jama.1992.03480240021012
- Jorgensen, J.H. and Pfaller, M.A. (editors-in-chief). (2015). Manual of Clinical Microbiology. Volumes 1 and 2. 11th ed. Carroll, K.C., Landry, M.L., Funke, G., Richter, S.S. and Warnock, D.W. American Society for Microbiology: ASM Press. Washington, D.C
- Kim, C.J., Emery, D.A., Rinke, H., Nagaraja, K.V. and Halvorson, D. A. (1989). Effect of Time and Temperature on Growth of *Salmonella enteritidis* in Experimentally Inoculated Eggs. Avian Diseases. 33(4): 735. doi:10.2307/1591153
- 25. Malaeb, M., Bizri, A. R., Ghosn, N., Berry, A., and Musharrafieh, U. (2016). Salmonella burden in Lebanon. Epidemiology and Infection. 144(08): 1761-1769. doi: 10.1017/s0950268815003076
- Mayo Clinic. E. coli Diseases and conditions: Causes. (2014). Mayo Foundation for Medical Education and Research (MFMER). Available online at: http://www.mayoclinic.org/diseases-conditions/ecoli/basics/causes/con-20032105
- Mishu, B., Koehler, J., Lee, L.A., Rodrigue, D., Brenner, F.H., Blake, P. and Tauxe, R.V. (1994). Outbreaks of Salmonella enteritidis infections in the United States, 1985-1991. J Infect Dis. 169: 547-52
- Mossad, S. B. (2013). Upper Respiratory Tract Infections. Cleveland Clinic: Center for continuing Education. Cleveland. USA. Available online at: http://www.clevelandclinicmeded.com/medicalpubs/diseasemanagement/infectious-disease/upper-respiratorytract-infection/
- Moubareck, C., Doucet-Populaire, F., Hamze, M., Daoud, Z. and Weill, F. (2005). First Extended-Spectrum--Lactamase (CTX-M-15) -Producing *Salmonella enterica* Serotype Typhimurium Isolate Identified in Lebanon. Antimicrobial Agents and Chemotherapy. 49(2): 864-865. doi:10.1128/aac.49.2.864-865.2005
- Musgrove, M.T., Jones, D.R., Northcutt, J.K., Harrison, M.A. and Cox, N. A. (2005). Impact of Commercial Processing on the Microbiology of Shell eggs. Journal of Food Protection, 68(11),:2367-2375. doi:10.4315/0362-028x-68.11.2367
- 31. Poppe, C. (1999). Epidemiology of *Salmonella enterica* serovar Enteritidis. In *Salmonella enterica* serovar Enteritidis in Humans and Animals. Saed A. (ed.). Ames, Iowa: Iowa State University Press, 3-18
- Samonis, G., Karageorgopoulos, D.E., Kofteridis, D.P., Matthaiou, D.K., Sidiropoulou, V., Maraki, S. and Falagas, M.E. (2008). *Citrobacter* infections in a general hospital: characteristics and outcomes. European Journal of Clinical Microbiology & Infectious Diseases. 28(1): 61-68. doi:10.1007/s10096-008-0598-z
- 33. Speck, M.L. (ed.). (1984). Compendium of Methods for the Microbiological Examination of Foods. 2nd ed. APHA. Washington, D.C.
- 34. World Health Organization Regional Office for Europe. (2000). 7th report of the WHO surveillance programme for the control of foodborne infections and intoxications in Europe (1993–1998). WHO.