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RESEARCH ARTICLE

Protective effects of date palm extract as natural antioxidants on hepatotoxicity induced by *Cerastes cerastes* venom in albino rats.

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Abstract

..... The present study was designed to determine the possible protective effects of date palm extract against Cerastes cerastes venom induced oxidative stress and biochemical changes in liver of albino rats. A patch of 104 male Wister albino rats averaged weights (130±10g) at the beginning of the experiment were divided into 6 main groups according to the treatment and requirements of the experiment. Control rats group, DPE group treated orally with DPE at dose 1.2 mg/kg body weight, venoms groups: Rats injected with a single dose of *Cerastes cerastes* for one day, this group divided into three subgroups injected by a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD₅₀ for one day also for two weeks groups, venoms treated with DPE groups treated orally with DPE for two weeks before /and after injecting of a single dose of venom at dose $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day. Each group contains 6 rats except venom groups of LD₅₀ were 8 rats and six rats were sacrificed after the 1st day, 2nd and 4th week from each group of treatment.

The results showed a significant raise of some hepatic parameters (ALAT, ASAT, ALP, TG, CHO and LDL-C), while a significant reduction in some other parameters (Total protein, albumin and HDL-C), on the 1st day and 2nd weeks in rats injected intraperitoneal with *Cerastes cerastes* venom as compared to the control groups. The administration of the DPE ameliorated the side effects of the poisonous changes of *Cerastes cerastes* venom.

In conclusion, according to the results obtained revealed that the administration of DPE had hepatoprotective effects against envenomation with *Cerastes cerastes* venom in male Wister albino rats by inhibiting oxidative stress through ROS scavenger and improvement of the biochemical markers.

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Introduction:-

Venomous animal bites are a significant health problem for rural populations in many parts of the world (Warrell, 1992). Venom of snakes consists of a complex mixture of many substances, such as toxins, enzymes, growth factors, activators and inhibitors with a wide spectrum of biological activities and non-enzymatic proteins with pro and anticoagulant activities, causing hemorrhage, hypovolemic shock, thrombosis and tissue damage of vital organs (White, 2005; Al-Sadoon *et al.*, 2013; Cherifi and Laraba-Djebari, 2013 and Berling and Isbister, 2015).

Viperidae toxicity is including cytotoxicity, haemotoxicity and neurotoxicity (Masood, 2012 and WHO, 2012). Viper venoms are slowly absorbed and reach maximum serum levels within 6-24 hours (Warrell, 1995).

The most common venoms snake bite was of the horned viper, *Cerastes cerastes* (Al-Sadoon, 2015), It contains different enzymes displaying proteolytic activity and causes many toxicities (Warrell, 2004). Also Boumaiza *et al*, (2016) suggested that significant physiopathological changes in liver, heart and skin. Enzyme in *Cerastes cerastes* such as, phospholipases A_2 (PLA₂) has been associated with multiple toxicities including lung toxicity (Uma *et al.*, 2000), nephrotoxicity (de Castro *et al.*, 2004), cardiotoxicity (Cher *et al.*, 2005), neurotoxicity and hepatotoxicity (Mukherjee and Maity, 1998).

The lethal effects of snake venom were largely attributed to its active ingredients as phospholipase A_2 (PLA₂). Phospholipid hydrolysis by PLA₂ releases arachidonic acid whose metabolism results in the formation of potentially toxic ROS and lipid peroxides (Adibhatla *et al.*, 2003 and Abdel-Rahman *et al.*, 2013).

The rise in the activities of liver enzymes indicate the damage of liver, heart and other organs brought about by the venom (Al-Sadoon and Fahim, 2012 and Al-Quraishy, *et al.*, 2014). Salman (2009) reported that intraperitoneal injection of the venom of *Echis coloratus* induced a significant increase in plasma cholesterol level in rabbits.

Antioxidants play an important role in inhibiting and scavenging radicals, thus providing protection to infection and degenerative diseases (**Reddy**, *et al.*, **2010**). Nutritional antioxidants are important in controlling and improving the harmful effects of oxidative stress, High eating of fruits with high antioxidant content contributes to reduced risk of oxidative stress-mediated diseases such as liver disease and cancer (**Lindquist**, *et al.*, **2000**).

Date palm (*Phoenix dactylifera*), is a good source of energy, vitamins, and a group of elements like phosphorus, iron, potassium and calcium (**El-Gazzar** *et al.*, 2009). Dates contain several vitamins including a small amount of vitamin C, and vitamins B1 (thiamine), B2 (riboflavin), nicotinic acid (niacin) and vitamin A (Al-Shahib and Marshall, 1993) Many researchers have also documented the antioxidant property of dates (Mohamed and Al-Okbi, 2004; Allaith and Abdul, 2005 and Al-Qarawi *et al.*, 2008).

Dates are considered to be one of the most significant commercial crops and also have been documented in Holy Quran and modern scientific literature. Earlier studies have shown that constituents of dates act as potent antioxidant, anti-tumour as well as anti-inflammatory, provide a suitable alternative therapy in various diseases cure (**Rahmani**, *et al.*, **2014**).

Also, date palm extract (DPE) has anticancer (**Ishurd** *et al.*, **2004**) and antiviral (**Vayalil**, **2002**), antimutagenic properties (**Hasan** *et al.*, **2010**) and immunomodulatory benefits to health but also has diverse medicinal values, including antihyperlipidemic, gastroprotective, hepatoprotective, and nephroprotective properties (**Tang** *et al.*, **2013**).

Date palm could be considered as a functional protective food for liver toxicity (**Abdu. 2011**). Aqueous DPE have strong antioxidant activity it's attributed to the wide range of phenolic compounds in dates including p-coumaric, ferulic and sinapic acids, flavonoids and procyanidins (**Kelm**, *et al.*, **2003**; **Al-Farsi**, *et al.*, **2005**).

Date palm characteristic contents of a high percentage of carbohydrates, dietary fibers, fats, proteins, vitamins and minerals gave it the superiority over other types of dates (Al-Shahib and Marshall, 2003 and Abdu, 2011). Melatonin is found to be capable of protector of DNA (LopezBurillo *et al.*, 2003), protein and lipids in cellular membranes (Cuzzocrea and Reiter, 2001). It also acts as an antagonist and suppresses of a number of endogenous and exogenous free radicals generated during the cellular process (Zang *et al.*, 1998 and Guo *et al.*, 2003).

Abdelrahman *et al.* (2012) Studied pretreated dogs with DPE found that significantly decrease the levels of liver enzymes at 6 and 14 days post CCL_4 injection. Palm date syrup showed significant hepatoprotective activity in carbon tetrachloride (CCl_4) induced hepatotoxicity among New Zealand rabbits (Mallhi *et al.*, 2014); El Arem *et al.* (2014) demonstrated a hepatoprotective effect of date palm fruit extract on oxidative damage induced by dimethoate, trichloroacetic, and carbon tetrachloride (CCl_4).

Aim of the work:-

The present study aimed to evaluate the ameliorative effects of date palm extract in reducing the hazards resulted from venom injection.

Materials and Methods:-

Experimental Design:-

One hundred and four male Wister albino rats (*Rattus norvegicus*) of about 130 ± 10 g body weight were conducted in accordance with the criteria of the investigations and Ethics Committee of the Community Laws governing the use of experimental animals. The rats placed in regular designed cages and maintained in conditions of good ventilation, normal temperatures, and humidity range. Six rats were placed into each cage. Food and water were provided *adlibitum* to the animals.

The rats were distributed into 6 main groups and 16 subgroups according to the treatment and requirements of the experiment as the following: **Group I:** Control rats **Group II:** treated orally with DPE at dose 1.2 mg/kg body weight **Group III:** Rats injected with a single dose of *Cerastes cerastes* for one day, this group divided into three subgroup injected by a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group IV:** Rats injected with a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group IV:** Rats injected with a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group IV:** Rats injected with a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for two weeks, this group divided into three subgroup injected by a single dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group VI:** treated orally with DPE for two weeks and injected by a single dose of venom at dose $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group VI:** treated orally with DPE for two weeks and injected by a single dose of venom at dose $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day **Group VI:** treated orally with DPE for two weeks and injected by a single dose of venom at dose $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day and completely treated by DPE for two weeks.

The animals were observed daily for signs of toxicity. The body weights were recorded weekly during the period of the experiment. Six rats from each group were sacrificed on the 1st and 15th days for control and treatments.

Preparations of date palm extract (Phoenix dactylifera L.):-

Dates palm (*Phoenix dactylifera L.*) fruits were washed with tap water and the seeds were removed. The extract was prepared by removing the nuclei of the fruit and grounded into fine Powder. The flesh of the fruits was left in distilled water (1:3) for 48 hours in 4° C (**Al-Qarawi** *et al.*, **2005**). The whole solution was ground, then centrifuged at 4° C for 20 min at 4000 rpm. The supernatant was collected and stored at - 80°C till use (**Vayalil**, **2002**).

Dose calculation:-

The method described by **Sutken** *et al.*, (2007) the dose level of 1.2 mg/kg/day of DPE is equivalent to 7 DP for each person per day, the human date fruit recommended antioxidant dose was 35g/day. Rat dose (mg/kg) = 1.2 g/kg body weight, according to studies of Al-Qarawi *et al.*, (2004).

Determination of the half lethal dose (LD50):-

The approximate median lethal dose (LD_{50}) of the crude venom (*Cerastes cerastes*) was calculated according to the method described by **Meier and Theakston (1986).** The Lethal Dose (LD_{50}) of venom of *Cerastes cerastes* viper equal 3.8 mg/kg.

Induction of hepatotoxicity:-

Hepatotoxicity was induced by intraperitoneal injection of *Cerastes cerastes* venom at different doses ($1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50}).

Sample preparation:-

Blood samples were collected without anti-coagulants and centrifuged at 4000 r.p.m for 10 minutes to harvest serum. The serum was frozen at -20 °C until used.

Biochemical parameters:-

The serum levels of transaminases, alkaline phosphatase, total protein and albumin were estimated using kits from Egyptian Company for biotechnology spectrum, Egypt. The concentrations of transaminases (ALAT and ASAT) were determined using the method of **Reitman and Frankel** (1975). Serum ALP was determined according to the method described by **Tietz** *et al.*, (1983). Serum total protein was determined according to the method described by **Doumas** *et al.* (1971). Lipid profile was estimated using kits from Egyptian Company for biotechnology spectrum, Egypt. The concentrations of triglycerides were determined according to the method described by **MGowan** *et al.* (1983). Serum cholesterol level was determined according to the method described by **MGowan** *et al.* (1983). Serum cholesterol level was determined according to the method described by **Tietz** (1976). Serum HDL-C was determined according to the method of **Burstein** *et al.* (1970). The LDL-C uses estimated according to the formula of **Wieland and Seidel** (1982).

Statistical analysis:-

The statistical package for social sciences SPSS/PC computer program (version 20) was used for statistical analysis of the results. Data were analyzed using one-way analysis of variance (ANOVA). The data were expressed as mean \pm S.E. Differences were considered statistically significant at P < 0.05.

Results:-

The present results showed thatrats injected with a single dose of *Cerastes cerastes* venoms at $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} exhibited biochemical disorders established by significant increase (p \leq 0.05) in some hepatic parameters (ALAT, ASAT and ALP) post-injection with a single dose of venom for one day as well as for two weeks groups as compared to the control group. On the other hand, insignificant differences with recorded in DPE groups when compared to control groups. Also, the present study demonstrated that the rats treated with DPE at a dose level of 1.2 mg/kg/day before/ and after injection with a single dose of venom at doses $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} showed significantly decrease (P \leq 0.05) in liver function enzymes activities (ALAT, ASAT and ALP) when compared to venom groups. As shown in Table and figure (1, 2 and 3).

Serum total protein and albumin level in rats injected with a single dose of venom for one day as well as for two weeks groups recorded a significant decrease (p < 0.05) as compared to the control group. On contrast insignificant differences with recorded in DPE when compared to control groups. Rats treated with DPE at a dose level of 1.2 mg/kg/day before and /or after injection with a single dose of venom at doses 1/4 LD₅₀, 1/2 LD₅₀ and LD₅₀ observed a significant increase amelioration (p < 0.05) when compared with venom for two and four weeks. On the other hand, the rats treated with DPE for four week at a dose level of 1.2 mg/kg/day before and after injection with a single dose of venom at doses 1/4 LD₅₀, 1/2 LD₅₀ and LD₅₀, 1/2 LD₅₀ and LD₅₀ observed to control groups. As shown in Table and figure (4 and 5).

Resulted data which found in table and figure (6, 7, 8 and 10) showed a significantly increase (p < 0.05) in lipid profile (triglycerides, cholesterol and LDL-C) level, also a significantly decrease (p < 0.05) in HDL-C level in in rats injection with a single dose of venom for one day as well as for two weeks groups as compared to the control group. On the other hand, insignificant differences with recorded in DPE groups when compared to control groups. Rats treated with DPE at a dose level of 1.2 mg/kg/day before and /or after injection with a single dose of venom at doses 1/4 LD₅₀, 1/2 LD₅₀ and LD₅₀, observed a significant increase for (triglycerides, cholesterol and LDL-C) level and increase for HDL-C level (p < 0.05) when compared with venom for two and four weeks.

	1	Non veno	om group	DS						Venom g	roups					
Groups						Venom g	groups wit	thout dat	e palm			Venon	n groups	with da	te palm	
Groups	Сог	ntrol	Da	ate	Vei	nom for 1	day	Ven	om for 2v	veek	Date	1te 2w +venom 12 1/2 1/2 LD50 LI 2W	om	Da	ate2w+ve + Date 2	nom w
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Doco	Sa	line	1.20/	z/dov	1/4	1/2	1 D50	1/4	1/2	1 D50	1/4	1/2		1/4	1/2	
Dose			1.2g/	K/uay	LD50	LD50	LD30	LD50	LD50	LD30	LD50	LD50 LD50 LD5			LD50	LD50
Period	2W	4W	2W	4W		1 Day			2W			2W			4W	
Mean	45.63 ^a	45.75 ^a	45.79 ^a	45.39 ^a	104.92 ^b	108.06 ^C	d 115.66	99.8 ^e	f 102.46	110.33 ^c	77.23 ^g	85.32 ^h	i 88.03	46.02 ^a	46.8 ^a	48.28 ^a
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.56	0.71	1.12	0.57	1.19	0.46	0.98	0.68	0.85	0.87	0.58	1.2	1.68	0.39	0.56	0.6
%			0.35	0.25	59.29	62.43	70.03	54.17	56.83	64.69	31.6 39.69 42.3			0.28	1.1	2.5
Each value	e represe	nts mean	of 6 reco	rds ± S.E,	, Except LD	0 ₅₀ groups a	re 4 record	ls, Means	with dissin	nilar supe	rscript lett	er are sig	nificantly	different	at (p<0.0	5),

Table (1): Effect of pre-and post-treatment of date palm extract on serum alanine aminotransferase (ALAT) activity (U/ml) of rats injected with cerastes cerastes venom.

W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (1): serum alanine aminotransferase (ALAT) activity (U/ml) in different animal groups.

	N	on veno	om grou	ps						Venom	groups					
Groups						Venom	groups wi	ithout dat	e palm			Veno	m groups	with date	palm	
Groups	Con	trol	Da	ate	Vei	nom for 1	day	Ven	om for 2v	veek	Date	e 2w +ver	nom	Dat -	te2w+vend + Date 2w	om
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Doco	Sal	ine	1.20/	k/dov	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2		1/4	1/2	
Dose			1.2g/	к/цау	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀
Period	2W	4 W	2W	4 W		1 Day			2W			2W			4 W	
Mean	119.3 ^a	119.3 ^a	119.4 ^a	a 119.6	213.77 ^b	226.54 ^C	237.53 ^d	201.63 ^e	228.72 ^c	226.48 ^c	f 160.75	162.03 ^f	166.95 ^g	121.15 ^a	122.28 ^a	124.53 ^a
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.7	0.46	0.63	0.3	1.46	0.62	1.08	1.11	0.93	0.54	0.85	0.55	0.46	0.74	0.99	1.05
%			0.27	0.47	94.6	107.37	118.36	82.47	109.55	107.3	41.58	42.87	47.78	1.98	3.12	5.36
Each value	represei	nts mean	of 6 reco	ords ± S.E	, Except L	D ₅₀ groups	are 4 recor	ds, Means	with dissin	nilar super	script lette	r are signi	ficantly dif	ferent at (p	<0.05), W:	Weeks,

Table (2): Effect of pre-and post-treatment of date palm extract on serum aspartate aminotransferase (ASAT) activity (U/ml) of rats injected with *cerastes cerastes* venom.

Each value represents mean of 6 records ± S.E, Except LD₅₀ groups are 4 records, Means with dissimilar superscript letter are significantly different at (p<0.05), W:Wee Venom: single dose, % Percent of changes from control values.



Fig (2): Serum aspartate aminotransferase (ASAT) activity (U/ml) in different animal groups

	Ν	Non vei	10m grou	ps						Venom	groups					
Groups						Venom	groups w	vithout da	ate palm			Veno	m group	os with dat	e palm	
Groups	Cont	trol	Da	ate	Ver	om for 1	dav	Ven	om for 2	week	Dat	e 2w +ve	nom	Dat	e2w+veno	m
					ven		uuy	ven		week	Dui	Date 2w +venon 1 12 1/4 1/2 050 LD50 L 2W 2W	nom	+	- Date 2w	
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Dece	Sali	ne	1.2~/	1.2g/k/day		1/2	ID	1/4	1/2	ID	1/4	1/2		1/4	1/2	
Dose			1.2g/I	K/uay	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀
Period	2W	4 W	2W	4 W		1 Day			2W			2W			4 W	
Mean	116.82 ^a	117 ^a	117.15 ^a	117.23 ^a	263.33 ^b	265.64 ^b	277.67 ^C	223.3 ^d	220.88 ^d	219.91 ^d	201.5 ^e	209.52 ^f	213.33 ^f	151.77 ^g	h 160.5	i 167.55
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	1.08	1.28	0.8	0.59	0.83	0.66	1.06	0.81	0.77	0.62	3	1.97	0.83	2.33	2.51	2.17
%			0.33	0.42	146.52	148.82	160.85	106.48	104.06	103.1	84.68	92.7	96.51	34.95	43.68	50.73
Each value	represent	ts mean	of 6 record	ds ± S.E. F	xcept LD	groups a	are 4 recor	ds. Means	with dissi	imilar supe	erscript le	tter are s	ignificant	lv different	at (p<0.05)).

Table (3): Effect of pre-and post-treatment of date palm extract on serum alkaline phosphatase (ALP) activity of rats injected with *cerastes cerastes* venom.

Each value represents mean of 6 records ± S.E, Except LD₅₀ groups are 4 records, Means with dissimilar superscript letter are significantly different at (p<0.05), W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (3): serum alkaline phosphatase (ALP) activity (IU/L) in different animal groups.

Venom groups											ups	om gro	on ven	N		
palm	ith date p	groups wi	Venom		L	ate palm	ithout d	groups w	Venom g						Cround	
te2w+venom + Date 2w	Dat +	om	te 2w +ven	Dat	week	om for 2v	Ven	day	om for 1	Ven	ate	D	ntrol	Con	Groups	
15 16	14	13	12	11	10	9	8	7	6	5	4	3	2	1	Sub Gp.	
1/2 LD	1/4	ID	1/2	1/4	ID	1/2	1/4	ID	1/2	1/4	1.2g/k/day		line	Sal	Dogo	
LD_{50}	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	K/uay	1.4g/			Dose	
4 W			2W			2W			1 Day		4 W	2W	4 W	2W	Period	
6.2 ^{h,i} 6.06 ^{g,I}	6.31 ^h	5.89 ^f	5.75 ^g	5.96 ^g	5.27 ^c	5.54 ^{b,f}	5.69 ^e	4.96 ^d	5.14 ^c	5.2 ^b	6.65 ^a	6.61 ^a	6.61 ^a	6.64 ^a	Mean	
± ±	±	±	±	±	±	±	±	±	±	±	±	±	±	0.11	±	
0.11 0.12	0.11	0.12	0.11	0.11	0.12	0.11	0.12	0.12	0.11	0.12	0.12	0.11	0.11		SE	
-0.41 -0.55	-0.29	-0.75	-0.89	-0.68	-1.36	-1.1	-0.95	-1.68	-1.5	-1.44	0.02	0.03			%	
	LD ₅₀ 6.31 ^h ± 0.11 -0.29	5.89 ^f ± 0.12 -0.75	LD ₅₀ 2W 5.75 ^g ± 0.11 -0.89	LD ₅₀ 5.96 ^g ± 0.11 -0.68	5.27 ^c ± 0.12 -1.36	LD ₅₀ 2W 5.54 ^{b,f} ± 0.11 -1.1	LD ₅₀ 5.69 ^e ± 0.12 -0.95	4.96 ^d ± 0.12 -1.68	LD ₅₀ 1 Day 5.14 ^c ± 0.11 -1.5	LD ₅₀ 5.2 ^b ± 0.12 -1.44	4W 6.65 ^a ± 0.12 0.02	2W 6.61 ^a ± 0.11 0.03	4W 6.61 ^a ± 0.11	2W 6.64 ^a ± 0.11	Period Mean ± SE	

Table (4): Effect of pre-and post-treatment of date palm extract on serum total protein concentration (g/dl) of rats injected with cerastes venom.

Each value represents mean of 6 records \pm S.E, Except LD₅₀ groups are 4 records, Means with dissimilar superscript letter are significantly different at (p<0.05), W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (4): Serum total protein concentration (g/dl) in different animal groups.

	N	on veno	om grou	ps						Venom	groups					
Croups						Venom	groups v	without da	te palm			Venon	n groups	with dat	e palm	
Groups	Con	trol	D	ate	Vei	nom for 1	day	Veno	m for 2w	veek	Dat	e 2w +ve	nom	Dat	te2w+ver + Date 2v	iom w
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Dece	Sal	ine	1.2~	/lr/dow	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2		1/4	1/2	
Dose			1.2g/	K/uay	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀
Period	2W	4 W	2W	4W		1 Day			2W			2W			4 W	
Mean	4.42 ^a	4.43 ^a	4.41 ^a	4.43 ^a	3.36 ^b	3.27 ^b	3.5 ^c	3.88 ^d	3.68 ^e	3.7 ^e	4.01 ^g	3.93 ^{d,g}	3.93 ^{d,g}	4.3 ^a	4.31 ^a	4.21 ^a
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.03	0.3	0.3	0.3	0.05	0.017	0.025	0.082	0.063	0.07	0.04	0.02	0.05	0.03	0.03	0.04
%			-0.07	-0.02	-1.06	-1.15	-0.92	-0.55	-0.74	-0.71	-0.41	-0.49	-0.48	-0.12	-0.11	-0.21
Each value	represen	ts mean	of 6 reco	rds + S.E	Except	LD ₅₀ grou	ns are 4 r	ecords. Me	ans with	dissimila	r supersc	rint letter	are signif	ïcantly di	ifferent af	ł

Table (5): Effect of pre-and post-treatment of date palm extract on serum albumin concentration (g/dl) of rats injected with *cerastes cerastes* venom.

Each value represents mean of 6 records \pm S.E, Except LD₅₀ groups are 4 records, Means with dissimilar superscript letter are significantly different at (p<0.05), W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (5): serum albumin concentration (g/dl) in different animal groups.

		Non ven	om groups							Venor	n groups					
Groups						Venom	groups w	ithout dat	e palm			Veno	m groups	with dat	e palm	
Groups	Con	trol	Da	ate	Ver	nom for 1 (dav	Ven	om for 2w	eek	Dat	e 2w +ven	om	Ι	Date2w+ven	om
							v								+ Date 2w	V
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Dece	Sali	ine	1.20/	k/dov	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID
Dose			1.2g/	K/uay	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD_{50}
Period	2W	4 W	2W	4 W		1 Day			2W			2W			4 W	
Mean	114.55 ^a	114.5 ^a	114.52 ^a	114.45 ^a	164.32 ^b	147.97 ^c	183.5 ^d	143.67 ^e	147.11 ^f	155.6 ^g	129.85 ^h	138.22 ⁱ	139.68 ⁱ	118 ^{j.k}	119.35 ^{j.k}	121.32 ^k
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.36	0.58	0.25	0.3	1.12	1.48	1.48	0.79	1.31	1.03	0.75	0.62	1.28	0.37	0.33	0.4
%			0.03	0.1	49.77	60.42	68.95	29.12	32.56	41.04	15.3	23.67	25.13	3.45	4.8	6.77
Each valu	e represen	ts mean	of 6 record	ls ± S.E, E	xcept LD ₅	o groups a	re 4 recor	ds, Means	with dissi	milar sup	oerscript le	tter are sig	nificantly	differen	t at (p<0.05	i) ,

Table (6): Effect of pre-and post-treatment of date palm extract on serum Triglycerides (TG) concentration (g/dl) of rats injected with *cerastes cerastes* venom.

W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (6): serum triglycerides (TG) concentration (mg/dl) in different animal groups.

	I	Non veno	om group	s						Venom	groups					
Groups						Venom	groups wi	thout da	te palm			Venon	n groups	with dat	e palm	
Groups	Con	itrol	Da	ate	Vei	nom for 1	day	Ven	om for 2	week	Date	te 2w +venom 12 13 1/2 LD LD ₅₀ LD 2W	nom	Dat	te2w+ven + Date 2w	0 m
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Dece	Sal	ine	1.20/	l/dov	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID
Dose			1.2g/	к/uay	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	$\begin{array}{c c} \mathbf{D}_{50} & \mathbf{L}\mathbf{D}_{50} \end{array} \begin{array}{c c} \mathbf{L}\mathbf{D}_{50} & \mathbf{L}\mathbf{D}_{50} \end{array}$		LD ₅₀	LD ₅₀	LD ₅₀
Period	2W	4 W	2W	4W		1 Day			2W			2W			4W	
Mean	114.14 ^a	114.06 ^a	114.36 ^a	114.3 ^a	203.26 ^b	214.96 ^c	218.77 ^d	178.88 ^e	184.72 ^f	195.43 ^g	137.23 ^h	143.88 ⁱ	158.05 ^j	120.36 ^k	123.78 ^l	125.06 ^l
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.54	0.48	0.54	0.86	1.13	0.67	0.72	0.86	0.97	0.85	1.17	0.73	1.24	0.69	0.44	1.85
%			0.21	0.16	89.12	100.82	104.63	64.74	70.57	81.29	23.09	29.73	43.9	6.21	9.64	10.92
Each value	represent	s mean of	6 records	$s \pm S.E, Ex$	cept LD ₅₀	groups ar	e 4 records	, Means v	vith dissiı	nilar supe	rscript let	ter are sig	gnificantly	y different	at (p<0.05	5),

Table (7): Effect of pre-and post-treatment of date palm extract on serum cholesterol concentration (mg/dl) of rats injected with *cerastes cerastes* venom.

Each value represents mean of 6 records ± S.E, Except LD₅₀ groups are 4 records, Means with dissimilar superscript letter are significantly different at (p<0.05). W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (7): serum cholesterol (CHO) concentration (mg/dl) in different animal groups.

	N	on venc	om grou	ps						Venom	groups					
Groups						Venom	groups v	vithout da	ate palm			Ven	om grou	ps with	date palm	
Groups	Cor	ntrol	Da	ate	Vor	om for 1	day	Vor	om for 2	vool	Det	2	nom	D	0ate2w+ve	nom
					vei		uay	v en		WEEK	Date	Date 2w +venor 11 12 1 1/4 1/2 L .D ₅₀ LD ₅₀ L	nom		+ Date 2	W
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Doso	Sa	line	1.2g/k/dav		1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID
Dose			1.2g/	к/uay	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD_{50}	LD ₅₀	LD ₅₀	LD_{50}
Period	2W	4 W	2W	4 W		1 Day			2W			2W			4W	
Mean	27.26 ^a	27.11 ^a	27.03 ^a	27.08 ^a	145.06 ^b	152.97 ^c	158.57 ^d	124.48 ^e	130.9 ^f	136.06 ^g	69.76 ^h	77.23 ⁱ	93.11 ^j	32.62 ^k	46.18 ^l	52.04 ^m
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.78	0.39	0.47	0.68	1.09	1.71	1.25	1.99	1.84	2.31	1.01	1.03	1.78	0.82	0.65	1.27
%			0.22	0.18	117.8	125.71	131.31	97.22	103.64	108.81	42.5	49.97	65.85	5.36	18.92	24.78
Each value	represen	ts mean	of 6 reco	rds ± S.F	E, Except I	D ₅₀ group	os are 4 re	cords, Me	ans with di	issimilar sı	perscrip	t letter a	re signifi	icantly di	fferent at (p<0.05),

Table (8): Effect of pre-and post-treatment of date palm extract on serum low-density lipoprotein cholesterol (LDL-C) concentration (mg/dl) of rats injected with *cerastes cerastes* venom.

W:Weeks, Venom: single dose, % Percent of changes from control values.



Fig (8): Serum low-density lipoprotein cholesterol (LDL-C) concentration (mg/dl) in different animal groups.

	1	Non ven	om grou	ps						Venom	groups					
Croups					-	Venom g	groups w	vithout da	te palm			Venon	ı groups	s with dat	e palm	
Groups	Co	ntrol	Da	ate	Veno	om for 1	day	Veno	m for 2v	veek	Dat	e 2w +vei	nom	Date +	e2w+venc Date 2w)m
Sub Gp.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Doco	Sa	line	1.2 a/	1.2g/k/day		1/2	ID	1/4	1/2	ID	1/4	1/2	ID	1/4	1/2	ID
Duse			1.2g/	к/uay	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀	LD ₅₀
Period	2W	4 W	2W	4 W		1 Day			2W			2W			4 W	
Mean	67 ^a	67 ^a	67.33 ^a	67.25 ^a	25.33 ^{b,d}	27 ^{b,d}	23.5 ^b	25.67 ^{b,d}	24.4 ^b	28.25 ^d	41.5 ^e	39 ^{e,f}	37 ^f	56.17 ^g	50.67 ^h	45.5 ⁱ
±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±	±
SE	0.58	0.58	0.67	0.81	1.12	1.05	0.65	1.5	1.21	3.09	0.92	0.58	1.08	0.83	1.43	2.25
%			0.33	0.25	-41.67	-40	-43.5	-41.33	-42.6	- 38.75	-25.5	-28	-30	-10.83	-16.33	-21.5
Each value (p<0.05), W	represe V:Week	ents mean s, Venom	n of 6 rec 1: single d	ords ± S. lose, % P	E, Except I ercent of c	LD ₅₀ grou hanges fr	ips are 4 om contr	records, M ol values.	eans with	ı dissimila	r supers	cript lette	r are sigr	nificantly d	ifferent at	

Table (9): Effect of pre-and post-treatment of date p	palm extract on serum high-densit	y lipoprotein cholesterol (HDL	-C) concentration (mg/dl) of rats
injected with cerastes cerastes venom.			



Fig (9): Serum high-density lipoprotein cholesterol (HDL-C) concentration (mg/dl) in different animal groups.

Discussion:-

Liver enzymes (ALAT and ASAT) are markers for cellular damage, although ALAT is more specific to detect liver injury the liver than is ASAT.

The increases of ALAT, ASAT and ALP enzyme activities in rats injected with a single dose of venom (*Cerastes* cerastes) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day as well as two weeks showed significant increase (p<0.05) when compared to the corresponding values in control group. This increase could be attributed to venom induced oxidative stress on the liver cells and distortion of the architecture pattern of the hepatocytes by the interaction of released free radicals with cell membrane and this may be indicated to necrosis or hepatocellular injury; These results are in agreement with (Al-Jammaz 2001; Al-Sadoon and Fahim, 2012 and Al-Sadoon *et al.*, 2013).

Otherwise their activities in the groups pre/or post-treated with DPE and injected with a single dose of venom (*Cerastes cerastes*) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for two weeks as well as four weeks showed significant decrease (p<0.05) when compared to the corresponding values in venom groups. Also showed insignificant change in the rats treated with DPE for four week plus a single dose of venom at doses $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} , showed insignificant change in liver function enzymes activities (ALAT and ASAT) when compared to control groups. This reduction in the serum enzymes activities by DPE may be due to reduction in the lipid peroxidation of hepatocellular membrane induced by the venom. While, it is may be due to the accelerated regeneration/ repairing of damaged hepatocytes.

El-Gazzar *et al.*, (2009) suggested that aqueous DPE can be ameliorated hepatotoxicity induced by CCl_4 , also **Araak and Abdulhussein** (2012) who demonstrated that treatment of rabbits with date palm pollen aqueous suspension decreased serum levels of ALT, AST and ALP induced by administration of CCl_4 .

DPE had been found to contain high amount of melatonin and vitamin E (Al-Qarawi et al., 2005), and this may reduce the hydroxyl and superoxide anion radicals (Vayalil, 2002). The decrease in ALAT, ASAT and ALP activities may improve the liver activity and this in agreement with other research worked on CCL4, Trichloracetic acid and Ochrotoxin A, El-Mougy et al., 1991; Zang et al., 1998; Cuzzocrea and Reiter, 2001; Al Qarawi, et al., 2004; Al-Farsi et al., 2005; Bastway et al., 2008; Rock et al., 2009; Saafi et al., 2010; Abd el-rahman et al., 2012; Azadbakht et al., 2012; Ragab et al., 2013; Sangi et al., 2014 and Sheikh et al., 2014.

The results of serum total protein and albumin concentrations in the groups injection with a single dose of venom (*Cerastes cerastes*) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day as well as two weeks showed a significant decrease (p<0.05) when compared with the corresponding values in control group. This decrease may be due to inhibition of oxidative phosphorylation which leads to decrease in protein synthesis, increase in catabolic processes and reduction of protein absorption. As well as the albumin levels in hepatic were found to be decreased in envenomed rats.

Meier and Stocker 1991 and March *et al.*, 1997 described reduction of total protein and albumin in envenomated rats was observed by the increasing in vascular permeability and hemorrhages in vital organs, and this is agreement with (Al-Sadoon, 1991; Abdul- Nabi *et al.*, 1997; Fahim 1998; Al-Jammaz *et al.*, 1998 and 1999; Al-Sadoon and Fahim, 2012). Also, this is in accordance with the results of Salman, (2009) and (2014) who showed the levels of serum albumin, globulin and total protein were significantly decreased in the envenomated guinea pigs by viper. Also, the level of total protein indicated a significant decrease in the envenomated male albino rats after injected with *cerastes viper* snake venom (Al Jammaz, *et al.*, 2002). The results also are in agreements with the finding of abdou *et al.* (2015) who indicated that plasma total protein which is an indicator for hepatic function showed a significant decrease in male albino mice injected interaprotenial with a single dose.

Otherwise their activities in the groups pre/and post treated with DPE and injection with a single dose of venom (*Cerastes cerastes*) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for two weeks as well as four weeks revealed significant increase (p<0.05) in serum albumin, globulin and total protein when compared to the corresponding values in venom group. On the other hand, the rats treated with DPE for four weeks plus injection with a single dose of venom at doses $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ showed insignificant change in albumin when compared to control groups.

Okwuosa et al. (2014) demonstrated that treatment of rats with aqueous DPE and methanolic DPE, respectively, improves thioacetamide-induced liver damage represented by alterations in liver function parameter. Dates seeds significantly increased serum level of albumin (Paranthaman et al., 2013). While, the present results are this

disagreement with the findings of **Orabi and Shawky**, 2014 reporting that Date seed causes a significant decrease in total protein and ALAT when compared with the control groups.

The results of the serum total cholesterol, triglycerides and low-density lipoprotein cholesterol concentrations showed significant increase as well as a significant decrease of high-density lipoprotein cholesterol concentration in the groups injection with a single dose of venom (*Cerastes cerastes*) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for one day and for two weeks when compared to the corresponding values in control group. The results may be due to the hepatocytes damage rendering them unable to phosphorylate the increasing amounts of fatty acids, hence leading to fatty liver and alteration of cell membranes of tissues. Coronary heart disease is strongly related to decrease in the concentrations of HDL-cholesterol and increase in LDL-cholesterol (**Salah and Al-Maiman, 2005**). **Al-Jammaz** (**2001**) and **Salman** (**2009**) observed that the envenomated rats and guinea pigs showed an increase in serum cholesterol levels and triglycerides.

Results decreasing protein synthesis mainly, low density lipoprotein (LDL) that is responsible for carrying capacity of triglycerides from the liver to extra hepatic tissues, thus lipids tends to accumulate inside the hepatocytes giving the characteristics of fatty liver or steatosis (**Clawson et al, 1987**).

Al-Sadoon *et al.*, 2013 suggested that the increases in serum total cholesterol, triglycerides and low-density lipoprotein cholesterol concentrations in envenomated guinea pig could be due to the hepatocytes damage rendering them unable to phosphorylate the increasing amounts of fatty acids, hence leading to fatty liver and alteration of cell membranes of tissues and this is in agreement with the finding of Al-Jammaz, (2002); Lewis and Gutmann, (2004); Salman, (2009) and Al-Sadoon and Fahim, (2012). And disagreement with Abd El-Aal and Ezzat (1997) found that a sub-lethal dose of *Naja haje* venom decreased serum total cholesterol in rabbits.

On the other hand, those levels in the groups of pre-or post-treated with DPE and injected with a single dose of venom (*Cerastes cerastes*) in TG,CHO and LDL-C at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD_{50} for two weeks revealed significant decrease when compared to the corresponding values in venom groups. while, the groups of pre-or post-treated with DPE and injection with a single dose of venom (*Cerastes cerastes*) at dose of $1/4 \text{ LD}_{50}$, $1/2 \text{ LD}_{50}$ and LD₅₀ for four weeks revealed a very high significant decrease as compared to the corresponding values in venom groups.

A reduction in serum levels of triglyceride may be due to decreased lipogenesis and increased lipolytic activity by activation of the hormone-sensitive lipase or lipogenic enzymes (**Pari and Venkateswaran, 2004**). Also, the present results are in agreement with **Borochov-Neori** *et al.* **2013** who used ethanol and acetone extracts of DPE to inhibited LDL oxidation, and most extracts also stimulated total cholesterol removal from macrophages. Also, acetone extracts exhibited a significantly higher anti-atherogenic potency for most varieties. Our data are in agreement with **Abdelaziz and Ali, (2014) and Sheikh** *et al.* **(2014)** who found that treatment of rats with DP seeds and DPE, respectively, reduced the occurrence of liver injuries caused by CCl_4 intoxication.

Our data coincide with **Abdelaziz and Ali (2014)** and **Sheikh** *et al.* (2014) who found that treatment of rats with DP seeds and DPE, respectively, reduced the occurrence of liver injuries caused by CCl₄ intoxication.

Salah and Al-Maiman (2005) reported that feeding date seed flour to rats reduced the plasma triglycerides, total cholesterol and low density lipoprotein, and this in agreement with (El-Mougy *et al.*, 1991; Panahi and Asadi, 2009 and Mard *et al.* 2010).

Conclusion:-

The dates (DPE) showed improvement in all biochemical parameters of lives human after injection of venom at different doses.

References:-

- 1. Abdel-Aziz D. H. and Ali S. A. (2014): The protective effect of *Phoenix dactylifera* L. seeds against CCl4induced hepatotoxicity in rats, *Journal of* Ethnopharmacology, vol. 155(1), pp. 736–743.
- 2. Abdel-Rahman H. A., Fathalla S.I., Mohamed A. A., Jun H. K. and Kim D. H. (2012): Protective Effect of Dates (*Phoenix dactylifera L.*) And Licorice (*Glycyrrhiza glabra*) on Carbon Tetrachloride Induced Hepatotoxicity in Dogs, Global Veterinaria, vol. 9(2), pp. 184-191.
- **3. Abdel-Rahman M. A., Tashiro Y. and Sonomoto K. (2013):** Recent advances in lactic acid production by microbial fermentation processes, Biotechnology advances, vol. 31(6), pp. 877-902.

- 4. Abdou R. H., Fatma S., Abd El-Ghany., Ismail M., Abdel-Nabi. and Mohamed A., Abdel-Rahman. (2015): Toxicological Effects of the Horned Viper Venom *Cerastes Cerastes* and Their Neutralization With the Egyptian Scorpion *Androctonus Australis Heamolymph*, indian journal of applied research, vol. 5 (3), pp. 37-41.
- 5. Abdu S.B. (2011): Protective role of Ajwa date against the hepatotoxicity induced by Ochratoxin A, Egyption. J. Nat. Toxins, vol. 8(1, 2), pp. 1-15.
- 6. Abdul-Nabi I. M., Raafat A. and El-Shamy H. I. (1997): Biological effects of intraperitoneal injection of rats with the venom of the snake *Echis carinatus*, Egypt. J. Zool., vol. 29, pp. 195-205.
- 7. Adibhatla R.M., Hatcher J.F. and Dempsey R.J. (2003): Phospholipase A2, hydroxyl radicals, and lipid peroxidation in transient cerebral ischemia, Antioxid Redox Signal, vol. 5, pp. 647–654.
- 8. Al-Farsi M., Alasalvar C., Morris A., Baron M. and Shahidi F. (2005); Comparison of antioxidant activity, anthocyanins, carotenoids and phenolics of three native fresh and sun dried dates (*Phoenix dactylifera* L.) Varieties grown in Oman, J. Agri. Food Chem., vol. 53(19), pp. 7592-7599.
- 9. Al-Jammaz I. A. (2001): Effects of single doses of Bitis arietans crude venom on serum biochemical parameters in rats, Sci. J. King Faisal Univ. Saudi Arabia, vol. 2, pp. 103-112.
- 10. Al-Jammaz I., Al-Ayed M. I. and Al-Yahya H. (1998): Effect of acute envenomation with LD₅₀ of *B. arietans*, Ain. Shams. Sci. Bull., vol. 36, pp. 207-222.
- 11. Al-Jammaz I., Al-Sadoon M. K. and Fahim A. (1999): Effect of LD₅₀ dose of *Echis coloratus* venom on serum and tissue metabolites and some enzyme of male albino rats, J. King Saud Univ., vol. 11 (2), pp. 61-68.
- 12. Al-Jammaz, I. A. (2002): Efects of envenomation by *Cerastes vipera* crude venom on plasma and tissue metabolites of rats, Kuwait J. Sci. Eng, vol. 29(1), pp. 111-119.
- **13.** Allaith and Abdul A. A. (2005): In vitro evaluation of antioxidant activity of different extracts of *Phoenix dactylifera* L. fruits as functional foods, Deutsche Lebensmittel Rundschau, vol. 101, pp. 305-305.
- 14. Al-Qarawi A.A., Abdel-Rahman H., Mousa H.M., Ali B.H. and El-Mougy S.A. (2008): Nephroprotective action of *Phoenix dactylifera* in gentamicin-induced nephrotoxicity, Pharm Biol , vol. 4, pp. 227-230.
- 15. Al-Qarawi A.A., Mousa H.M., Ali B.E.H., Abdel-Rehman H. and El-Mougy S.A. (2004): Protective effects of extracts from date (Phoenix dactylifera L.) on CCl4 induced hepatotoxicity in rats, Intern. J. Appl. Res. Vet. Med, vol. 2(3), pp. 176-180.
- 16. Al-Quraishi S., Metwaly M. S., DkhilK M. A., Gewik M.M., Hassan A.S. and Zrieq R. (2014): Palm pollen as growth and metabolic enhancer during the course of murine intestinal eimeriosis, Pakistan J. Zool., vol. 46, pp. 423-430.
- 17. Al-Sadoon M. K. (1991): Metabolic rate-temperature curves of the horned viper, Cerastes cerastes gasperetti, the moila snake, Malpolon moilensis, and the adder, Virera berus, Comparative Biochemistry and Physiology Part A: Physiology, vol. 99(1), pp. 119-122.
- **18. Al-Sadoon M. K. (2015):** Snake bite envenomation in Riyadh province of Saudi Arabia over the period (2005–2010), Saudi Journal of Biological Sciences, vol. 22, pp. 198–203.
- **19. Al-Sadoon M. K. and Fahim A. (2012):** Possible recovery from an acute envenomation in male rats with LD₅₀ of *Echis coloratus* crude venom: IA seven days hematological follow-up study, Saudi journal of biological sciences, vol. 19(2), pp. 221-227.
- **20.** Al-Sadoon M.K., Abdel Moneim A.E., Bauomy A.A. and Diab M.M. (2013): Histochemical and Biolchemical effects induced by LD₅₀ of *Cerastes cerastes gasperetti* crude venom in mice, Life Sci J, vol. 10(4), pp. 810-817.
- 21. Al-Shahib W. and Marshall R. J. (2003): The fruit of the date palm: Its possible use as the best food for the future, International Journal of Food Science and Nutrition, vol. 54, pp. 247–259.
- 22. Araak J. K. and Abdulhussein M. A. A. (2012): The Protective Role of Date Palm Pollen (*Phoenix dactylifera L.*) on Liver Function in Adult Male Rats Treated with Carbon Tetrachloride, In Proceeding of the Eleventh Veterinary Scientific Conference, vol. 132, pp. 141.
- 23. Azadbakht, L., F. Haghighatdoost, G. Karimi and A. Esmaillzadeh, (2012): Effect of consuming salad and yogurt as preload on body weight management and cardiovascular risk factors: A randomized clinical trial. Int. J. Food Sci. Nutr., 10.3109/09637486.2012.753039.
- 24. Bastway. Ahmed M, Hasona N.A, and Selemain A.H, (2008): Protective Effects of Extract from Dates (*Phoenix Dactylifera* L.) and Ascorbic Acid on Thioacetamide-Induced Hepatotoxicity in Rats. Iranian Journal of Pharmaceutical Research, vol. 7 (3), pp. 193-201.
- 25. Berling I. and Isbister G. K. (2015): Prolonged QT risk assessment in antipsychotic overdose using the QT nomogram, Annals of emergency medicine, vol. 66(2), pp. 154-164.

- 26. Borochov-Neori H., Judeinstein S., Greenberg A., Volkova N., Rosenblat M. and Aviram M. (2013): Date (Phoenix dactylifera L.) fruit soluble phenolics composition and anti-atherogenic properties in nine Israeli varieties, Journal of agricultural and food chemistry, vol. 61(18), pp. 4278-4286.
- 27. Boumaiza S., Oussedik-Oumehdi H. and Laraba-Djebari F. (2016): Pathophysiological effects of *Cerastes cerastes* and *Vipera lebetina* venoms: Immunoneutralization using anti-native and anti-60 Co irradiated venoms, Biologicals, vol. 44(1), pp. 1-11.
- 28. Burstein, M., Scholnick, H. R., and Morfin, R. (1970): Rapid method for the isolation of lipoproteins from human serum by precipitation with polyanions. J. lipid. Res., 11(6):583-595.
- 29. Cher C.D.N., Armugam A., Zhu Y.Z. and Jeyaseelan K. (2005): Molecular basis of cardiotoxicity upon cobra envenomation. Cell Mol, Life Sci., vol. 62(1), pp. 105-118.
- **30.** Chérifi F. and Laraba-Djebari F., (2013): Isolated biomolecules of pharmacological interest in hemostasis from *Cerastes cerastes* venom, Journal of Venomous Animals and Toxins including Tropical Diseases, vol. 19(1), pp. 1.
- **31.** Cuzzocrea S. and Reiter R.J. (2001): Pharmacological action of melatonin in shock, inflammation and ischemia/reperfusion injury, Eur. J. Pharmacol., vol. 426(1), pp. 1-10.
- **32. de Castro I., de A Burdmann E., Seguro A. C. and Yu L. (2004):** Bothrops venom induces direct renal tubular injury: role for lipid peroxidation and prevention by antivenom, Toxicon, vol. 43(7), pp. 833-839.
- 33. Doumas B.T., Watson W.A. and Biggs H.G. (1971): Albumin standard and the measurement of serum albumin with bromocresol green Clin Chim Acta, vol. 31, pp. 87 96.
- 34. El Arem A., Saafi E. B., Ghrairi F., Thouri A., Zekri M., Ayed A. and Achour L. (2014): Aqueous date fruit extract protects against lipid peroxidation and improves antioxidant status in the liver of rats subchronically exposed to trichloroacetic acid, Journal of physiology and biochemistry, vol. 70(2), pp. 451-464.
- 35. El-Aal A. A. and Ezzat A. R. (1997): Effects of three fractions obtained from *Naja haje* venom on hemolysis and lipid metabolism in rabbits. Journal of Venomous Animals and Toxins, vol. 3(2), pp. 311-323.
- **36.** El-Gazzar U.B., El-Far A.H. and Abel Maksoud H.A. (2009): Amerliorative effects of *Phoenix dactylifera* extract on CCl4 hepatotoxicity in New Zealand rabbits, J. Appl. Sci. Res., vol. 5(9), pp. 1082-1087.
- 37. El-Mougy S. A., Abdel-Aziz S. A., Al-Shanawany M. and Omar A. (1991): The gonadotropic activity of Palma in mature male rats, Alexandria Journal of Pharmaceutical Sciences, vol. 5, pp. 156–159.
- **38.** Fahim A. (1998): Biological effects of the viper *B. arietans*, crude venom on albino rats, Egypt. J. Zool., vol. 30, pp. 35-54.
- **39.** Gornall A.G., Bardawill C.J. and David M.M. (1949): Determination of serum protein by means of the biuret reagent, J Biol Chem, vol. 177(2), pp. 751-766.
- 40. Guo C., Yang J., Wei J., Li Y., Xu J. and Jiang Y. (2003); Antioxidant activities of Peel, Pulp and seed fractions of common fruits as determined by FRAP Assay. Nutr Res, vol. 23(12), pp. 1719-1726.
- **41. Hasan S, Amom H, Nor I, Mokhtarrudin N, Esa M, Azlan A. (2010):.**Nutritional composition and in vitro evaluation of the antioxidant properties of various dates extracts (*Phoenix dactylifera L*) from libya.Asian J.Clin.Nutr, vol. 2,pp. 208-214.
- 42. Ishurd O., Zgheel F., Kermagi A., Flefla M. and Elmabruk M. (2004): Antitumor activity of beta-Dglucan from Libyan dates, J. Med. Food, vol. 7, pp. 252-255.
- **43.** Kelm J. M., Timmins N. E., Brown C. J., Fussenegger M. and Nielsen L. K. (2003): Method for generation of homogeneous multicellular tumor spheroids applicable to a wide variety of cell types, Biotechnology and bioengineering, vol. 83(2), pp. 173-180.
- 44. Lewis R. L. and Gutmann L. (2004): Snake venoms and the neuromuscular junction, Seminars in neurology, vol. 24(2), pp. 175-179.
- **45.** Lindquist C.H., Gower B.A. and Goran M.I. (2000): Role of dietary factors in ethnic differences in early risk of cardiovascular disease and type 2 diabetes, Am J Clin Nutr, vol. 71, pp. 725–732.
- 46. López-Burillo S., Tan D. X., Rodriguez-Gallego V., Manchester L. C., Mayo J. C., Sainz R. M. and Reiter, R. J. (2003): Melatonin and its derivatives cyclic 3-hydroxymelatonin, N1-acetyl-N2-formyl-5-methoxykynuramine and 6-methoxymelatonin reduce oxidative DNA damage induced by Fenton reagents, Journal of pineal research, vol. 34(3), pp. 178-184.
- 47. Lopez-Burillo, S.; Tan, D.X.; Mayo, J.C.; Sainz, R.M.; Manchester, L.C. and Reiter, R.J. (2003): Melatonin, xanthurenic acid, resveratrol, EGCG, vitamin C and alpha-lipoic acid differentially reduce oxidative DNA damage induced by Fenton reagents: a study of their individual and synergistic actions. J. Pineal Res, vol. 34,pp. 269-277.

- **48.** Mallhi T.H., Qadir M.I., Ali M., Ahmad B., Khan Y.H. and Rehman A.U. (2014): Ajwa Date (*Phoenix dactylifera*): An Emerging Plant in Pharmacological Research. Pak. J. Pharm. Sci, vol. 27(3), pp. 607-616.
- **49.** March J. S., Parker J. D., Sullivan K., Stallings P. and Conners C. K. (1997): The Multidimensional Anxiety Scale for Children (MASC): factor structure, reliability, and validity, Journal of the American academy of child & adolescent psychiatry, vol. 36(4), pp. 554-565.
- Mard S.A., Jalalvand K., Jafarinejad M., Balochi H. and Naseri M.K.G. (2010): Evaluation of antidiabetic and Antilipaemic Activities of Hydroalcoholic extract of *Phoenix dactylifera* Palm leaves and its fractions in Alloxan-Induced diabetic rats, Maylaysian J. Med. Sci., vol. 17(4), pp. 4-13.
- 51. Masood M. F. (2012) Ecological distribution of snakes' fauna of Jazan region of Saudi Arabia.
- 52. Meier J. and Stocker K. (1991): Effect of snake venoms on homeostasis, Toxicology, vol. 21 (3), pp. 1711-1820.
- **53.** Meier J. and Theakston R. D. Q. (1986): Approximate LD₅₀ Determinations of snake venoms using eight to ten experimental animals, Toxicon, vol. 24(4), pp. 395-401.
- 54. MGowan M.W., Artiss J.D., Standbergh D.R. and Zak B. (1983): A peroxidase-coupled method for colorimetric determination of serum triglycerides, Clin Chem, vol. 29:538, pp.452.
- 55. Mohamed D. A. and Al-okbi S.Y. (2004): In vivo evaluation of antioxidant and anti-inflammatory activity of different extract of date fruit in adjuvant arthritis, J food Nutr Sci, vol. 13, pp. 397-402.
- 56. Mukherjee A. K. and Maitt C. (1998): Effect of oral supplementation of vitamin E on the hemolysis and erythrocyte phospholipid-splitting action of cobra and viper venoms, Toxicon, vol. 36, pp. 657-664.
- 57. Muriel P., Garciapiña T., Perez-Alvarez V. and Mourelle M. (1992): Silymarin protects against paracetamol-induced lipid peroxidation and liver damage, Journal of Applied Toxicology, vol. 12(6), pp. 439-442.
- 58. Okwuosa C.N., Udeani T.K., Umeifekwem J.E., Onuba A.C., Anioke I.C. and Madubueze R.E. (2014): Hepatoprotective effect of methanolic fruit extracts of *Phoenix dactylifera* (Arecaceae) on thioacetamide induced liver damage in rats, A J P C T, vol. 2(3), pp. 290-300.
- **59. Orabi S. H. and Shawky S. M. (2014):** Effect of Date Palm (*Phoenix Dactylifera*) Seeds Extracts on Hematological, Biochemical Parameters and Some Fertility Indices in Male Rats, International Journal of Sciences: Basic and Applied Research, vol. 17(1), pp. 137-147.
- **60. Panahi A. and Asadi M. (2009):** Cholesterol lowering and protective effects of date fruit extract: An in vivo study, Toxicology Letters, vol. 189, S235.
- 61. Pari L. and Venkateswaran S. (2004): Protective role of Phaseolus vulgaris on changes in the fatty acid composition in experimental diabetes, Journal of Medicinal food, vol. 7(2), pp. 204-209.
- 62. Ragab A. R., Elkablawy M. A., Sheik B. Y. and Baraka H. N. (2013): Antioxidant and Tissue-Protective Studies on Ajwa Extract: Dates from Al-Madinah Al-Monwarah, Saudia Arabia, Journal of Environmental & Analytical Toxicology, vol. 3, pp. 2161-2175.
- 63. Rahmani A.H., Aly S.M., Ali H., Babiker A.Y., Srikar S. and khan A.A. (2014): Therapeutic effects of date fruits (*Phoenix dactylifera*) in the prevention of diseases via modulation of anti-inflammatory, anti-oxidant and anti-tumour activity, Int J Clin Exp Med, vol. 7(3), pp. 483-491.
- 64. Reddy C. V. K., Sreeramulu D. and Raghunath M. (2010): Antioxidant activity of fresh and dry fruits commonly consumed in India, Food Research International, vol. 43(1), pp. 285-288.
- 65. Reitman S. and Frankel S. (1975): Am .J.Clin.Path, vol. 28, pp. 65.
- 66. Rock W, Rosenblat M, Borochov-Neori H, Volkova N, Judeinstein S, Elias M, Aviram M. (2009): Effects of date (*Phoenix dactylifera L.*, Medjool or Hallawi Variety) consumption by healthy subjects on serum glucose and lipid levels and on serum oxidative status: a pilot study. J Agric Food Chem, Sep 9; 57(17): 8010-7.
- 67. Saafi EB, Louedi M, Elfeki A, Zakhama A, Najjar MF, Hammami M, Achour L. (2010): Protective effect of date palm fruit extract (*Phoenix dactylifera L.*) on dimethoate induced-oxidative stress in rat liver. Exp Toxicol Pathol,vol. Jul; 63(5):,pp. 433-441.
- **68.** Salah A. and Al-Maiman. (2005): Effect of date palm (*Phoenix dactylifera*) seed fibers on plasma lipids in rats, Journal of King Saud University, vol. 17, pp. 117–123.
- **69.** Salman M.M. A. (2009): Physiological effects of envenomation by two different doses of the viper *Echis coloratusis* crude venom on biochemical parameters in serum of Guinea pigs at different times, Egypt. Acad J. biolog. Sci., vol. 1(1), pp. 21-31.
- 70. Sheikh B. Y., Elsaed W. M., Samman A. H., Sheikh B. Y. and Ladin A. M. M. B. (2014): Ajwa dates as a protective agent against liver toxicity in rat, European Scientific Journal, vol. 3, pp.358 368.

- 71. Sutken E., Aral E., Ozdemir F., Uslu S., Alatas O. and Colak O. (2007): Protective role of melatonin and coenzyme Q10 in ochratoxin A toxicity in rat liver and kidney, Int. J. Toxicol., vol. 26, pp. 81-87.
- 72. Tang Z. X., Shi L. E. and Aleid S. M. (2013): Date fruit: chemical composition, nutritional and medicinal values, products, Journal of the Science of Food and Agriculture, vol. 93(10), pp. 2351-2361.
- **73.** Thibault R., Blachier F., Darcy-Vrillon B., De Coppet P., Bourreille A. and Segain J. P. (2010): Butyrate utilization by the colonic mucosa in inflammatory bowel diseases: a transport deficiency. Inflammatory bowel diseases, vol. 16(4), pp. 684-695.
- 74. Tietz N.W., Rinker A.D. and Shaw L.M. (1983): IFCC methods for the measurement of catalytic concentration of enzymes. Part 5. IFCC method for alkaline phosphatase, J Clin Chem Clin Biochem, vol. 21, pp. 731-748.
- **75.** Uma B. and Gowda T. V. (2000): Molecular mechanism of lung hemorrhage induction by VRV-PL-VIIIa from Russell's viper (Vipera russelli) venom, Toxicon, vol. 38(8), pp. 1129-1147.
- 76. Vayalil P.K. (2002): Antioxidant and antimutagenic properties of aqueous extract of date fruit (*Phoenix dactylifera* L. Arecaceae), J. Agri. Food and Chem., vol. 50(3), pp. 610-617.
- 77. Warrell D.A. (1995): Clinical toxicology of snake bite in Africa and the Middle East/ Arabian Peninsula. In: Meier J, White J, editors. Hand book of clinical toxicology of animal venoms and poisons. 5. ed. Boca Raton: CRC Press, 433-92.
- 78. Warrell, D. A. (1992): Venomous bites and stings in the tropical world. The Medical journal of Australia, vol. 159(11-12), pp. 773-779.
- **79.** Warrell, D.A. (2004): Epidemiology, clinical features and management of snake bites in Central and South America. In: Campbell, J., Lamar, W.W. (Eds.), Venomous Reptiles of the Western Hemisphere. Cornell University Press, Ithaca, pp. 709–761.
- **80.** White J. (2005): Snake venoms and coagulopathy, Toxicon, vol. 45(8), pp. 951-967.
- 81. Who U. (2012): UNFPA, the World Bank. Trends in maternal mortality: 1990 to 2010. World Health Organization, UNICEF, UNFPA, and the World Bank.
- **82.** Wieland H. and Seidel D. (1982): Improved assessment of plasma lipoprotein patterns. IV. Simple preparation of a lyophilized control serum containing intact human plasma lipoproteins, Clinical. Chemistry, vol. 28(6), pp. 1335-1337.
- 83. Zang L.Y., Cosma G., Gardner H. and Vallyathan V. (1998): Scavenging of reactive oxygen species by melatonin, Biochim. Biophys. Acta, vol. 1425, pp. 469-477.