

Journal homepage: http://www.journalijar.com

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH

RESEARCH ARTICLE

Potential Anti-inflammatory Effect of Telmisartan In Experimentally Induced Acute Pancreatitis in Rats.

SAMIA M. M. ELSHIATY¹, HANAN T. EMAM², THANAA BELAL³

1, 2.Department of Pharmacology, Faculty of Medicine, Benha University, Egypt.

3. Department of Biochemistry, Faculty of Medicine, Benha University, Egypt.

Manuscript Info

Abstract

Manuscript History:

Received: 12 April 2014 Final Accepted: 23 May 2014 Published Online: June 2014

Key words:

Acute pancreatitis (AP) -Peroxisome proliferators activated receptor (PPARs) -Thiazolidinediones (TZDs) -Rosiglitazone (Ros)-**Telmisartan (Tel).**

Corresponding Author HANAN.T. EMAM Background: Acute pancreatitis (AP) is an acute inflammatory disorder of the pancreas. The destruction of pancreatic parenchyma induces a systemic activation of coagulation, kinin, complement and fibrinolytic cascades with liberation of cytokines and reactive oxygen metabolites.

.....

Peroxisome proliferator–activated receptors (PPARs) play key roles in the processes of fat metabolism, adipocyte differentiation, tumorigenesis, inflammation and variety of immune processes. Telmisartan is the only angiotensin receptor blocker (ARB) that may modulate PPAR γ activation at physiologic plasma concentrations. Telmisartan may reduce markers of inflammation, such as interleukin-6 and C-reactive protein, oxidative stress and improves markers of vascular function.

The aim of the current article was to investigate the possible antiinflammatory, antioxidant effects, and peroxisome proliferator-activated receptor- γ (PPAR γ) properties of the angiotensin type 1 receptor blocker telmisartan in experimentally induced acute pancreatitis.

Materials and Methods:Fifty male albino rats were divided randomly into 5 groups; control normal group and 4 acute pancreatic groups group (n=40). Acute pancreatitis were induced by i.p. injection of 20% L-arginine hydrochloride in saline (2x250 mg/100 g at 1 h interval) and subdivided equally into acute pancreatic group (no treatment); AP treatedt with Tel or Ros alone or their combination groups. Serum amylase and lipase activity were determined. Also TNF- α , IL-6, MDA and CAT were estimated in addition to histopathological examination.

Results: Ros alone or in combination with Tel induced significant attenuation of serum amylase and lipase activity. Also TNF- α , IL-6, MDA and CAT were improved. While Tel induced insignificant improvement of previous parameters

Conclusion: Use of rosiglitazone associated with a reduced risk of AP by improving inflammatory status and oxidative stress reflecting the important role of PPAR- γ in AP. Furthermore, telmisartan does not ameliorate acute pancreatic inflammation. Both agents have affinity for PPAR- γ , telmisartan binds to the receptor in a different manner, resulting in distinct pharmacological actions. *Copy Right, IJAR, 2014, All rights reserved.*

864

INTRODUCTION

Acute pancreatitis (AP) still remains an enigmatic clinical problem. No specific treatment is available to treat AP. Many therapies and medical management is aimed to control the sign and symptoms of AP, using steroids, analgesics and anti-inflammatory agents. These compounds are very expensive and not reliable. Hence, there is need of potential antioxidant & anti-inflammatory agents which are cost effective and have several advantages than the others^[1].

Acute pancreatitis (AP) can sometimes, though not always, is a life-threatening disease with a significant impact on patient health. Up to 25% of patients with AP suffer a severe form which accounts for the high mortality rate of AP^[1,2]. Autoactivation of digestive enzymes in the pancreatic acinar cells and the subsequent destruction of pancreatic parenchyma induces a systemic activation of coagulation, kinin, complement and fibrinolytic cascades with liberation of pro- inflammatory cytokines, such as interleukin-1 β (IL-1 β), IL-6, tumor necrosis factor- α (TNF- α) and reactive oxygen metabolites. In addition, micro- circulatory disturbances and leukocytes activation play a crucial role in development of AP and its associated serious sequels ^[3,4,2].

The Peroxisome proliferators activated (PPAR) family consists of at least three different isoforms; PPAR α , PPAR δ , PPAR $\gamma^{[5]}$. The modulatory role of PPAR receptor has been proposed in the inflammatory response of different organs ^[6].

PPAR γ is a member of the nuclear hormone receptor superfamily originally reported to be expressed at high levels in adipose tissue and to play a critical role in adipocyte differentiation, glucose metabolism, lipid storage and suppress inflammatory gene expression^[7,8].

PPAR γ is expressed in both endocrine and exocrine pancreatic cell types^[9]. Its role in pancreas is not restricted to insulin signaling pathway, it has been shown to be imported in cell growth, metabolism (in particular response to altered energy homeostasis), apoptosis and inflammation^[10].

Activation of PPAR γ receptors leads to anti-inflammatory and antiproliferative effects as well as increased insulin sensitivity, improvement in dyslipidemia and regression of athrosclerosis ^[11].PPAR γ has powerful synthetic ligands, the thiazolidinediones (TZDs) also called glitazones; torglitazone, ciglitazone, pioglitazoner, and rosiglitazone. TZDs are used to treat type2 diabetes because of their efficacy in controlling blood glucose secondary to enhancing insulin action through a mechanism (s) that is yet to be completely elucidated^[12]. Currently, PPAR- γ agonists have also been found to have excellent antioxidant activity^[13,14]. Newer studies have also reported that PPAR- γ agonists exhibit anti-inflammatory properties which are due to negative regulation of the expression of pro-inflammatory molecules such as interleukin-1b (IL-1b), IL-6 and TNF- α ^[15,16]. Considerable evidence indicates that PPAR γ agonists inhibit inflammatory responses during inflammatory diseases^[17,18].

Rosiglitazone, PPAR γ agonist, is a widely used drug for the treatment of type 2 diabetes mellitus. It increases insulin sensitivity of peripheral tissues. In addition, there is evidence that rosiglitazone has anti-inflammatory effects [19,20].

Telmisartan is an orally active, long-acting, non-peptide angiotensin type 1 (AT₁) receptor blocker (ARB) with high selectivity for the AT₁ receptor. Telmisartan is a potent antihypertensive drug^[21].

In addition, telmisartan is the only ARB that may modulate PPAR γ activation at physiologic plasma concentrations, an effect that is likely to be related to telmisartan high lipophilicity. Several studies have found that telmisartan can influence PPAR γ gene expression. In treatment hypertensive patients, telmisartan treatment significantly increased PPAR γ mRNA levels in peripheral monocytes ^{[22].}

Telmisartan has also been shown in animal models to reduce the levels of several markers of inflammation, such as interleukins and TNF α . In a rat model, telmisartan protected against experimental autoimmune myocarditis, partly by suppressing inflammatory cytokines and oxidative stress^[23].

Despite various experimental and clinical testing of potential therapeutic drugs, no specific therapeutic strategy has been shown to be uniformly effective in controlling AP and its lethal complications. Therefore, there is a necessary need for development of more effective therapeutic alternatives.

This study aimed to determined the anti-inflammatory effect of telmisartan compared with rosiglitazone in experimental model of acute pancreatitis

MATERIALS AND METHODS

Animals:

Adult male albino rats, with initial body weight ranging from (150-200 g) were used. Rats were purchased from Experimental Animal Breeding Farm, Helwan. All animals were housed in a controlled laboratory conditions at 20-

25°C in a 12h light /dark cycle and had free access to food and water. They were allowed for one week acclimatization period before to their use in the experiment at Pharmacology Department, Faculty of Medicine, Benha University.

Drugs and Chemicals:

- L-arginine (Sigma, USA), it was supplied as pure powder and was dissolved in sterile saline (0.9%) , intraperitoneal (i.p.) administration .
- Rosiglitazone (Avandia; GlaxoSmithKline, USA) was administered after being suspended in 1ml of sterile saline (0.9%).
- Telmisartan (Boehringer Ingelheim, Germany) freshly prepared, was administered after being suspended in 1ml of distilled water.
- TNF-α detection ELISA kits was obtained from Biosource, Belgium.
 Drugs were given by the oral route through an orogastric catheter.
- All the other chemicals used were purchased from Sigma Co., USA

Experimental groups:

Fifty male albino rats were divided randomly into 5 groups; (1)control normal group received 1ml of physiologic saline by IP injection; and 4 acute pancreatic groups Acute pancreatitis were induced by two ip injection of 20% L-arginine hydrochloride in saline (2x250 mg/100 g at 1 h interval) causes pancreatitis without mortality ^[24] then, subdivided equally into; (2) acute pancreatic group(no medication); (3) acute pancreatic with rosiglitazone administration group, at the dose of 10mg/kg of bw after suspended in 0.9% NaCl to the volume of 1 ml, by an orogastric catheter ^[25], group (4) acute pancreatic with telmisartan administration group, at the dose of 10mg/kg of bw after suspended to the volume of 1 ml of distilled water by an orogastric catheter ^[23] and group (5) acute pancreatic with rosiglitazone plus telmisartan administration group.

Treatment started one hour prior to induction of AP, The rats were scarified after 24 hours from AP.

Experimental Parameters and design:

At the end of the experimental period, the animals were anesthetized by ether. Blood samples were obtained from retro-orbital venous plexus using capillary pipette. The blood clot and the samples were centrifuged for 15 minutes at 3000 rpm. Sera were separated and stored at -20C until being used for determination of serum amylase (U/L) according to the method of ^[26] and lipase activity (U/L) according to the method of ^[27]. Serum tumor necrosis factor- α (TNF- α) (pg/ml) and interleukin-6 (IL-6) (pg/ml) were determined in serum using the commercially available ELISA kits according to the methods of ^[28] and ^[29] respectively. Malondialdhehyde (MDA) (nmol/ ml) as a marker of lipid peroxidation following the method described by ^[30] and catalase (CAT) activity (U/ml) was determined following the method described by ^[31].

In addition, pancreas was isolated from animal after scarification. Pancreatic tissue was stored at -80C until further examination.

Histopathological Examination:

Samples of pancreatic tissue were fixed in 10% buffered formalin solution, embedded in paraffin using standard methods, cut into 5 um sections, stained with hematoxylin-eosin, and then assessed under light microscopy and examined for grading the histopathological alterations. Pancreatic edema, leukocyte infiltration, acinar vacuolization, and necrosis were described with scores ranging from 0 to 3 (0 being normal and 3 being severe) as previously described ^{[32, 33].}

Statistical analysis:

All data were expressed as mean \pm S.E.M. and analyzed with statistical package SPSS (Version 10, 2002) for Windows. One-way analysis of variance (ANOVA) was used to determine statistically significant differences among the groups, and means of every two different groups were detected with Student's t-test. p<0.05 was considered statistically significant.

RESULTS

In the present study, ip injections of L arginine induced acute pancreatitis as evidenced by significant increase (P<0.05) in serum levels of amylase and lipase (Tab.1, Fig.1, 2).

Pretreatment with rosiglitazone at the dose of 10 mg/kg at the same time of induction of AP, produced marked improvement of acute pancreatitis manifested by the significant decreases (P<0.05) in serum levels of amylase and lipase compared with acute pancreatitis. While, there was a significant increase (P<0.05) in serum amylase and lipase in rosiglitazone pretreated group when compared with control group (Tab.1, Fig.1, 2).

Pretreatment with telmisartan at the dose of 10 mg/kg of bw produced insignificant decrease (P>0.05) in serum amylase and lipase level as compared to AP group. While, there was a significant increase (P<0.05) in serum amylase and lipase in telmisartan pretreated group when compared with control group (Tab.1, Fig.1, 2).

Pretreatment with rosiglitazone plus telmisartan produced marked improvement of acute pancreatitis manifested by the significant decreases (P<0.05) in serum levels of amylase and lipase compared with acute pancreatitis. While, there was a significant increase (P<0.05) in serum amylase and lipase when compared with control group (Tab.1, Fig.1, 2).

Parameters	Serum amylase (U/L)	Serum lipase (U/L)	
Groups			
Control	18.36±0.79	663±73	
АР	863±15.8*	5965±513*	
Ros	298±36.2*#	1003±147*#	
Tel	795±18.2*	4978±394*	
Ros+Tel	199±19.5*#	963±89*#	

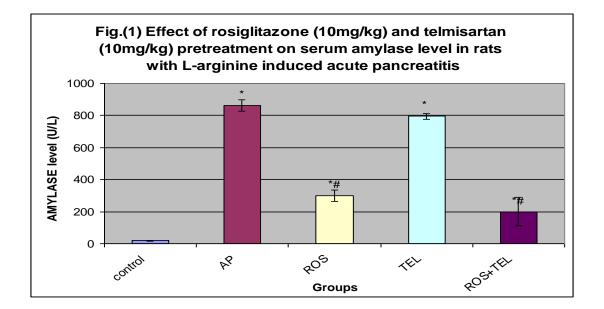
Table (1): Mean (±SE) Serum amylase (U/L), Serum lipase (U/L) reported in all studied groups (n=10)

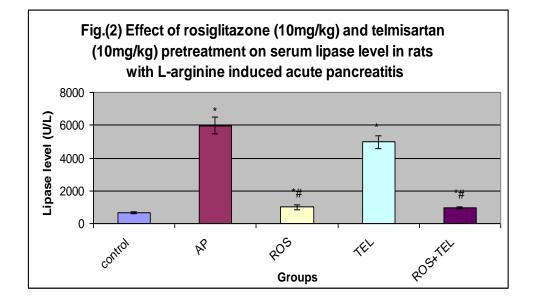
* : Significant difference versus control group.

: Significant difference versus AP.

AP : Acute pancreatitis

- Ros : Acute pancreatitis treated with rosiglitazone (single dose 10mg/kg of bw by an orogastric catheter).
- Tel : Acute pancreatitis treated with telmisartan (single dose 10mg/kg of bw by an orogastric catheter).





At 24hours after induction of AP, estimation of TNF- α and IL-6 serum levels show that L-arginine administration resulted in significant increase (P<0.05) in both parameters compared to control group (tab.2,Fig.3,4).

Pretreatment with rosiglitazone at the dose of 10mg/kg at the same time of induction of AP, produced marked improvement TNF- α and IL-6 serum levels (P<0.05) compared with AP group. While, there was a significant increase (P<0.05) in serum TNF- α and IL-6 in rosiglitazone pretreated group when compared with control group (tab.2, Fig.3, 4).

At the same time, Pretreatment with telmisartan at the dose of 10mg/kg of bw produced insignificant decrease (P>0.05) in improvement TNF- α and IL-6 serum levels (P>0.05) as compared to AP group. Also, there was a insignificant increase (P>0.05) in TNF- α and IL-6 serum levels in telmisartan pretreated group when compared with control group (tab.2, Fig.3, 4)

Furthermore, Pretreatment with rosiglitazone plus telmisartan, produced marked improvement TNF- α and IL-6 serum levels (P<0.05) compared with AP group. While, there was a significant increase (P<0.05) in serum TNF- α and IL-6 compared with control group (tab.2, Fig.3, 4).

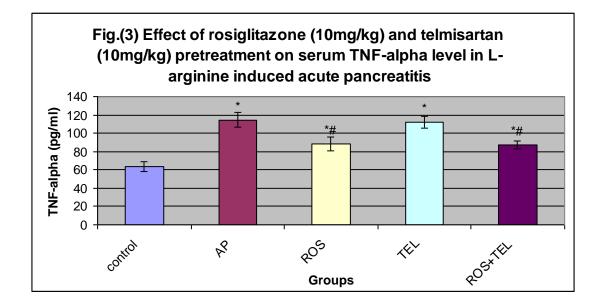
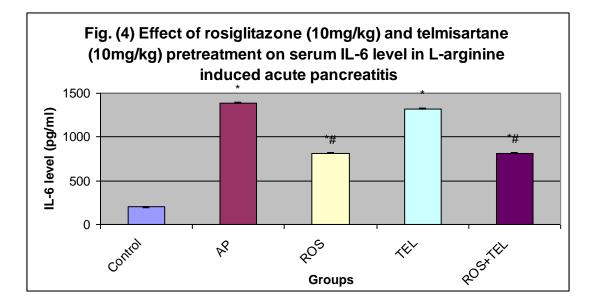


Table (2): Mean (±SE) TNF-α (pg/ml), IL-6 (pg/ml) reported in all studied groups (n=10)

Parameters Groups	TNF-α (pg/ml)	IL-6 (pg/ml)	
Control	63.4±5.1	201.9±17.2	
АР	114.4±7.9*	1391.8±106.6*	
Ros	88.5±7.7*#	816.2±73.1*#	
Tel	112±6.1*	1322.1±145*	
Ros+Tel	87.7±4.3*#	814.9±80.3*#	

- * : Significant difference versus control group.
- # : Significant difference versus AP.
- AP : Acute pancreatitis
- Ros : Acute pancreatitis treated with rosiglitazone (single dose 10mg/kg of bw by an orogastric catheter).
- Tel : Acute pancreatitis treated with telmisartan (single dose 10mg/kg of bw by an orogastric catheter).



As regards malondialdhehyde (MDA) and catalase (CAT) activity, estimation of MDA and CAT serum levels show that L-arginine administration resulted in significance increase (P<0.05) in both parameters compared to control group (table 3, Fig.5,6).

Pretreatment with rosiglitazone at the dose of 10mg/kg at the same time of induction of AP, produced marked improvement MDA and CAT (P<0.05) compared with AP group. While, there was a significant increase (P<0.05) in serum MDA and CAT in rosiglitazone pretreated group when compared with control group (Table 3, Fig.5, 6). At the same time, Pretreatment with telmisartan at the dose of 10mg/kg of bw produced significant decrease (P>0.05) in improvement MDA and CAT serum levels

(P<0.05) compared to AP group. Also, there was a significant increase (P<0.05) in MDA and CAT in telmisartan pretreated group when compared with control group Table 3, Fig.5, 6).

Parameters	MDA (nmol/ml)	Catalase (U/ml)
Groups		
Control	2.42±0.29	3.46±0.16
AP	10.87±0.52*	0.38±0.06*
ROS	4.85±0.22*#	1.89±0.07*#
TEL	5.65±0.71*#	1.21±0.11*#
ROS+TEL	4.21±0.35*#	1.09±0.09*#

Table (3): Mean (±SE) MDA (nmol/ml) and CAT (U/ml) reported in all studied groups (n=10).

* : Significant difference versus control group.

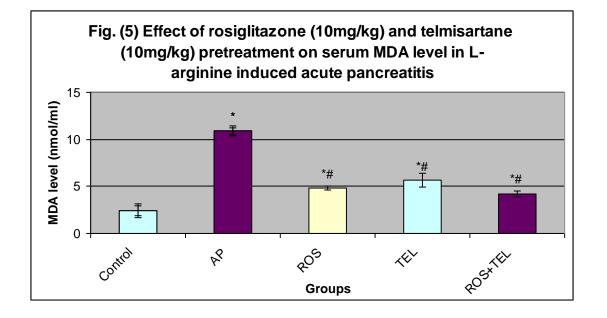
: Significant difference versus AP.

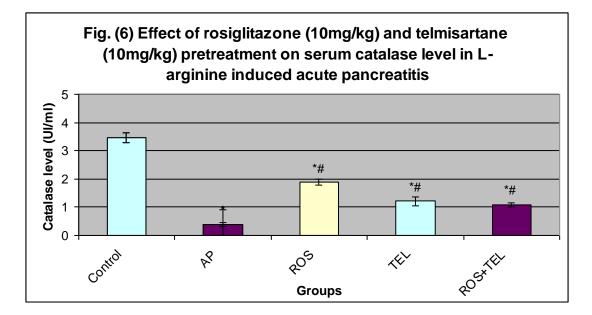
AP : Acute pancreatitis

Ros : Acute pancreatitis treated with rosiglitazone (single dose 10mg/kg of bw by an orogastric catheter).

Tel : Acute pancreatitis treated with telmisartan (single dose 10mg/kg of bw by an orogastric catheter).

While Pretreatment with rosiglitazone plus telmisartan produced marked improvement MDA and CAT (P<0.05) compared with AP group. While, there was a significant increase (P<0.05) in serum MDA and CAT compared with control group (Table 3, Fig.5, 6).





At the same time, the histopathological findings also supported these biochemical observations and indicated the presence of a severe form of AP in the pancreas of L- arginine-injected rats Fig.1B); which typically characterized by a high score of necrosis, vacuolization and edema of pancreatic acini, and inflammatory cell infiltration into the necrotic areas as compared with normal control group (Tab.1 Fig.1A).

871

Parameters	Edema	Vacuolization	Infiltration	Necrosis	Total
Groups					
Control	0.00	0.00	0.00	0.00	0.00
AP	2.33±0.47*	2.75±0.60*	2.50±0.5*	0.42±0.64*	8.00±1.08*
Ros	1.58±0.04*#	1.62±0.62*#	1.92±0.00*#	00.00#	5.12±0.92*#
Tel	2.00±0.41*	2.25±0.72*	2.01±0.64*	0.17±0.37*	6.43±1.47*
Ros+Tel	1.54±0.50	1.59±0.42	1.85±0.00	00.00	4.98±0.41*#

Table (4):Histopathologic grading" of morphologic alterations of pancreatic gland in groups of rats .

* : Significant difference versus control group.

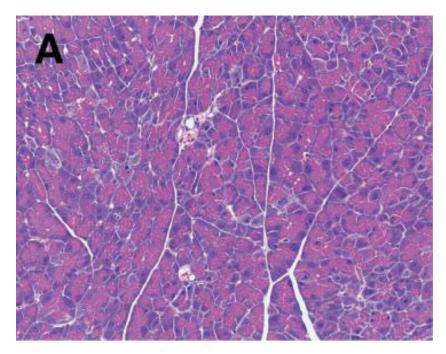
: Significant difference versus AP.

AP : Acute pancreatitis

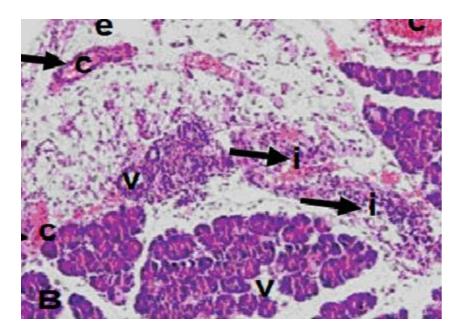
Ros : Acute pancreatitis treated with rosiglitazone (single dose 10mg/kg of bw by an orogastric catheter).

Tel : Acute pancreatitis treated with telmisartan (single dose 10mg/kg of bw by an orogastric catheter).

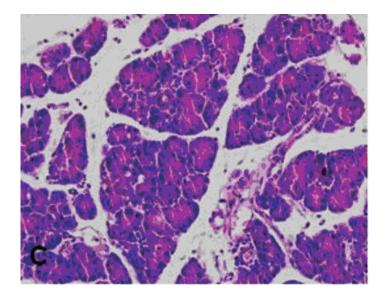
Fig.(7) Photomicrograph of pancreatic sections stained with hematoxylin- eosin



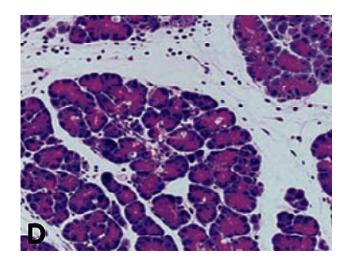
A section taken from the control rat shows normal pancreatic architecture, acini, blood vessels and normal islets of langerhans.



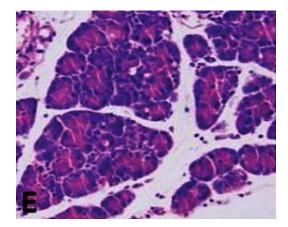
A section taken from the L-arginine induced acute pancreatitis shows sever interstitial edama "e",congested dilated vessels "c", vacuolization of acinar cytoplasm"v", and cellular inflammatory infiltration"i".



A section taken from rosiglitazone treated rats (10 mg/kg) shows a remarkable decrease in edema, disappearance of vacuolization and preservation of seminormal pancreatic acini.



A section taken from telmisartan treated rats (10 mg/kg) shows edema, less congestive blood vessels and vacuolization with cellular inflammatory infiltration. (magnificationX400).



A section taken from rosiglitazone+ telmisartan treated rats (10 mg/kg) shows a remarkable decrease in edema, disappearance of vacuolization and preservation of seminormal pancreatic acini and cellular inflammatory infiltration.

DISSCUTION

Experimental models of acute pancreatitis resembling the human situation are an integral tool for increasing the understanding of complex mechanisms as well as for developing therapeutic strategies for this disease

The present study aimed to explore the anti-inflammatory effects of the angiotensin type 1 receptor blocker telmisartan and rosiglitazone and their effect on peroxisome proliferator-activated receptor- γ (PPAR γ)-activating properties by analysis of serum interleukin 6 levels and TNF- α in addition to histopathological manifestation in L-arginine induced acute pancreatitis in rats.

In this work, intraperitoneal injection of L-arginine was used to induce acute pancreatitis as this model produces non-surgical, selective and dose-dependent acinar cell necrosis ^[24]. Furthermore, it was proven that oxygen-derived free radicals play a role in pathogenesis of L-arginine induced pancreatitis ^[34].

L-arginine was chosen in this experiment and animals were sacrificed 24hours after the second L-arginine injection as by the time acute pancreatitis would be established as proven by previous experimental works ^[35,36,37]. This experimentally induced AP is manifested both biochemical and histological changes which are very similar to the human disease.

In the present study, induction of pancreatitis significantly increased serum amylase, lipase levels at 24h. Serum amylase and lipase levels have been traditionally utilized and the important diagnostic markers for acute pancreatitis. These observations are in consistence with previous results which revealed that serum amylase and lipase usually rise within 4-8 hours of the initial attack, peaks at 24 hours and returns to control level over the27hrs ^[38,39,40]. The utility of serum amylase marker is complicated by significant limitations, including low sensitivity and specificity ^[41]. Because the pancreas is the only source of lipase, plasma lipase estimations are specific for pancreatic injury ^[42]. Serum lipase is recommended to replace or supplement amylase levels for the diagnosis of acute pancreatitis, an elevated lipase is virtually diagnostic for pancreatitis, approaching 100% specificity^[43].

In the present work, beside elevation of enzymes, plasma levels of pro-inflammatory cytokines (IL-6 and TNF- α) were significantly increased. Furthermore, serum level of MDA was remarkable increase with significantly decreased in serum catalase enzyme after induction of pancreatitis.

It is well known that the extent of pancreatic tissue damage in acute pancreatitis correlates with the levels of inflammatory mediators (IL, TNF and CRP) and free radical ^[35,38,44]. Similarly, the current study confirms and extends previous findings ^[37] that demonstrated the L-arginine induced pancreatitis is accompanied by remarkable increased plasma levels of IL-6 and TNF- α , together with amylase after induction of AP. ^{[[45,46]} found that the increased serum TNF- α , IL-6 andIL-1 levels were demonstrated at 24hs after the induction of pancreatitis with L-arginine in rat. Moreover ^[47,48] reported that increase serum level of TNF- α and IL-6 were detected 12hrs after administration of L-arginine and remained elevated at 48hrs versus normal control.

In consistent with our reports, ^[49] demonstrated that the pancreatic MDA level was significantly elevated at 24hrs and peaked at 48hrs after adminstration of L-arginine. In addition, the endogenous scavengers, superoxide dismutase and catalase activities decreased significantly throughout the entire study versus control. In accordance with this work, the results of ^[50] indicate that Oxygen free radicals (OFR) play an important mediator function in early and later courses of AP. Their findings suggest that OFR species are important mediators but not necessarily triggers of tissue damage in AP. Also, ^[37,38]) stated that induction of pancreatitis significantly increased the pancreatic total protein, MDA, nitrite, catalase and SOD and decreased the GSH levels. Lipid peroxidation is a process mediated by free radicals, which results in impairment of the membrane functional and structural integrity.

In addition, acute pancreatitis was also confirmed in our study by typical inflammatory features observed microscopically during the same duration in the present study, histopathological assessments revealed that induction of pancreatitis resulted in pancreatic damage characterized by acinar cell necrosis, cell infiltration, edema and haemorrhage this in agreement with previous reports by ^[34,38].

The present study focuses on the effect of rosiglitazone and telmisartan on experimental acute pancreatitis and possible anti-inflammatory and antioxidant effects. Rosiglitazone alone and in combination with telmisartan efficiently reduced serum amylase and lipase levels. The anti-inflammatory response in the pancreas reflected in reduced serum levels of TNF- α and IL-6. In this work, we have also evaluated the anti-oxidant effect of both drugs in experimental acute pancreatitis, our data indicate that they exhibit an antioxidant effect as evidenced by significant decrease in serum MDA with increase in catalase enzyme activity, this confirmed by histopathological manifestation improvement compared to acute pancreatitis

These results are in agreement with ^[51] who found statistically significantly decreased activity of amylase, lipase and pancreatic inflammation in male mice treated with rosiglitazone in course of edematous cerulean-induced acute pancreatitis. A study of ^[52] revealed that rosiglitazone acts directly through PPAR- γ in acini, duct and islets to suppress markers of the inflammatory response developed during cerulean-induced acute pancreatitis as the cell type specific knockout of PPAR- γ in epithelial cells of pancreas removes the inflammatory cytokine production, pancreatic edema and infiltration.

Several studies have demonstrated that the use of PPAR- γ ligands inhibits the intensity of the inflammatory response in different processes including colitis^[53], adjuant-induced arthritis^[54] and cerulien induced pancreatitis^[51]. There are more and more reports suggesting that PPAR plays a crucial role in the development of immune reaction; particularly in inflammation^[55,56]. PPARs- γ present in the alimentary canal (the liver, pancreas, large intestine) are characterized by the widest spectrum of the effects mentioned^[57,58]. The receptors might be indirectly activated by synthetic ligands such as thiazolidinediones (troglitazone, ciglitazone, pioglitazone, rosiglitazone^[59]. Many studies describe beneficial effects of PPAR- γ stimulation on inhibition of gastro-intestinal inflammation. The literature available, including studies in animal models, demonstrates decreased expression of inflammatory

cytokines, such as TNF-a, IL-1b, IL-6 and myeloperoxi- nki ^[60.61]. According to some other experimental studies, PPAR- γ ligands, through selective blockage of NF-kB signaling pathway in vitro, lead to a significant decrease in inflammatory IL-8 and MCP1 expression ^{[62,63].}

A cross-talk between pro-inflammatory cytokines and oxidative stress occurs in the development of the inflammatory response in AP, particularly TNF-alpha amplifies the inflammatory cascade through different mechanisms, such as the activation of mitogen activated protein kinases and nuclear factor-kappa B (NF-kB) and/or the inactivation of protein phosphatases^[64,65]. The current study confirms and extends previous reports, for example PPAR- γ stimulation by rosiglitazone reserved the oxidative changes induced by cold restraint stress (CRS). Rosiglitazone protective treatment significantly reduced gastric mucosal MDA and SOD activity and increase CAT activity, restoring their normal balance. ^[66,67]. The ROS lowering effect of the PPAR- γ agonist, rosiglitazone could be attributed to its inhibitory effect on TNF- α , an inflammatory mediator that increases ROS production during CRS. Furthermore, it has been reported that rosiglitazone has antioxidant activity and improves enzymatic antioxidant parameters like superoxide dismutase, catalase and glutathione peroxidase ^[67,68] which was confirmed in the present study.

Our results demonstrated that telmisartan was unable to alter the increased serum amylase and lipase levels to significant level. Also, inflammatory response showed insignificant reduced serum levels of TNF- α and IL-6 in telmisartan treated than experimental acute pancreatitis. In addition, we have evaluated the anti-oxidant effect of telmisartan which demonstrated significant improvement versus experimental acute pancreatitis. Combination of rosiglitazone and telmisartan treated rats was insignificantly improved parameters of AP compared to rosiglitazone alone but significant compared to AP.In accordance with previous reports, $^{[69]}$ reported that lesser serum TNF- α concentration in rosiglitazone and combination treatment as compared to disease control rats while rats treated with telmisartan showed higher serum concentration as compared to rats treated with rosiglitazone or its combination with telmisartan. These data suggest that telmisartan alone is ineffective in controlling TNF- α release. Effect observed in combination group is only due to the rosiglitazone which inhibits the TNF- α and attenuates the disease. Also, ^[70] reported that, in low-risk patient population, telmisartan (80 and 160 mg) treatment did not significantly affect serum interleukin 6 levels. These data are in contrast to a previously published study by ^[71]. In this study, the ARB olmesartan exhibited potent anti-inflammatory actions by reducing C-reactive protein, tumor necrosis factor-a, IL-6, and monocyte chemoattractant protein 1 serum levels .Also, ^[72,73] reported that telmisartan manifests powerful anti-inflammatory effects beyond class effects of angiotensin II Type 1 blocker by inhibiting TNF-a, induced IL-6 expressions through Peroxisome Proliferator Activated Receptor γ Activation .

^[74] Observed that Telmisartan directly ameliorates IL-1β-induced neuronal inflammatory response by inhibition of oxidative stress and the JNK/c-Jun pathway. This observation supported our result that the insignificant decrease in pancreatic inflammation of telmisartan may be due to antioxidant effect.

The histopathological manifestations in this work confirmed other results. These results supported by ^[69] who used rosiglitazone and telmisartan alone or in combination as anti-inflammatory in rheumatoid arthritis, through their observation that histopathology of synovial joint showed decreased vascular proliferation, decreased cartilage destruction, lesser decrease in synovial joint space and decreased chondrocytes migration in rats treated with rosiglitazone as compared to disease controlled rats. Rats treated with telmisartan showed higher cartilage destruction, higher vascular proliferation, decreased synovial joint space and higher chondrocytes migration as compared to rats treated with rosiglitazone or its combination with telmisartan. These findings suggest that telmisartan alone is ineffective in controlling the progression of Rheumatoid arthritis as compared to rosiglitazone. In contradicting to our results, ^[75] demonstrated that blockade of AT1 receptor by an AT1 receptor antagonist may act by normalizing pancreatic blood flow, resulting in reduced pancreatic inflammation.

From the previous observation we suggest that rosiglitazone is a potent anti-inflammatory in AP. On the other hand telmisartan was unable to reduce inflammation in AP in spite of telmisartan's bimodal mechanism of action (AT₁ receptor blockade + PPAR γ modulation). These observations can explain by other investigators, who revealed that TZDs agents were shown to act by stimulating a member of the nuclear hormone receptor family, Peroxisome proliferator–activated receptor (PPAR). TZDs are agonists for PPAR- γ , and, when bound to PPAR- γ receptor, lead to modulation of expression of specific genes, thus inhibiting production of a number of inflammation mediators when used at higher doses and hence it is having more adverse effect like weight gain and fluid retention^[76]. There is also a newer class of this family called Selective PPAR- γ Receptor Modulators (SPPARMs). These are the agents that partially activate the receptor by binding at the different site than the full agonist binds hence also called as partial agonist. Telmisartan, an antagonist of Angiotensin 2 type 1 is a SPPARM. In addition to its AT₁ receptor– blocking properties^[77,78,79]Compared with glitazones, as full PPAR γ agonist, telmisartan binds to the receptor in a different manner, resulting in distinct pharmacological actions. The lesser efficacy of telmisartan (binds to the receptor in a different manner) compared to rosiglitazone could be due to its partial agonistic activity on PPAR- γ ^[69]. The discrepancy between our results and other data may relate to the doses used.

These results are supported by the finding of several previous studies that indicated a dose dependency of telmisartan-mediated PPAR γ activation and show that further induction of monocytic PPAR γ can be achieved with higher doses ^{[70].}

Several studies have shown that the RAS is present intrinsically in the pancreas and that its level is enhanced during acute pancreatitis and chronic pancreatic hypoxia in experimental animals ^[80,81,82], suggesting the role of the RAS in pancreatic diseases. However, there are several controversial reports regarding the effects of the inhibition of the RAS on acute pancreatitis. For example, ^[83] reported that losartan, an AT1 receptor antagonist, ameliorates cerulean induced acute pancreatitis in rats. Moreover, ^[84] reported that ramipril, an angiotensin-converting enzyme (ACE) inhibitor, enhances acute pancreatitis in the same model.

In addition ^[85] demonstrated that the ANG II-AT1 receptor pathway is not essential for the local pancreatic injury in acute pancreatitis but plays an important role in the development of pancreatic fibrosis through pancreatic stellate cell (PSC) activation and proliferation.

^[86,87] reached far more that, they reported that clinicians should be aware to some ARB as irbesartan or losartan may cause acute pancreatitis. If abdominal pain develops, the medication should be discontinued and the patient investigated for acute pancreatitis. Angiotensin II receptors are thought to be important in regulation of pancreatic secretion and microcirculation, but the mechanism of pancreatitis induced by angiotensin II receptor antagonists remains unclear.

CONCLUSION:

Hence on the basis of the above discussion, our study adds some support to previous experimental findings, use of rosiglitazone associated with a reduced risk of AP by improving inflammatory status and oxidative stress reflecting the important role of PPAR- γ in AP. Furthermore, telmisartan does not ameliorate acute pancreatic inflammation. Both agents have affinity for PPAR- γ , telmisartan binds to the receptor in a different manner, resulting in distinct pharmacological actions. Further studies are needed to establish the full potential effect of high-dose telmisartan therapy.

ACKNOWLEDGEMENTS

The authors would like to thank Faculty of Medicine, Benha University especially Dr Asmaa M. Elhady lecturer of histopathologylogy for her sincere help and her effort in this work.

REFERENCES

- ^[1]Al Mofleh IA (2008): Severe acute pancreatitis: pathogenetic aspects and prognostic factors. World J. Gastroenterol., 14: 675-68.
- ^[2]Wu BU and Conwell DL (2010): Update in acute pancreatitis. Curr. Gastroenterol. Rep., 12: 83-90
- ^[3]Chan YCand Leung PS (2007): Acute pancreatitis: animal models and recent advances in basic research. Pancreas, 34: 1-14.
- ^[4]Pandol SJ, Saluja AK, Imrie CW, Banks PA (2007): Acute pancreatitis: bench to the bedside. Gastroenterology, 133: 1056. 1-1056.
- ^[5]Mangelsdorf D J, Thhummel C, Beato M L (1995):The nuclear receptor superfamily: the second decade. Cell,83:835-839.
- ^[6]Chinetti G, Fruchart J C, Steals B (2003): peroxisome proliferators activated receptors and inflammation: from basic science to clinical applications. Int J Obse. Relat. Metab. Disord.,27 (suppl3): S41-45
- ^[7]Landreth G, Jiang Q, Mandrekar S (2008): PPAR gamma agonists as therapeutics for the treatment of Alzheimer's disease. Neurotherapeutics 5: 481–489.
- ^[8]Folch-Puy E, Granell S, Iovanna J L Barthet M, Closa D (2006): peroxisome proliferators activated receptor-gamma agonist reduced the severity of post-ERCP pancreatitis in rat. World J Gastroentrol., 12(40): 6458-6463.

- ^[9]Welters H J, Mc Bain S C, Tadayon M (2004): Expression and functional activity of PPAR gamma in pancreatic beta cells.Br.J.Pharmacol.,142:1162-70.
- ^[10]Semple R K, Chatterjee V K, ORahilly S (2006): PPARgamma and human metabolic disease. J clin. Inves.,116 : 581-589.
- ^[11]Lehrke M and Lazar MA(2005):Themany faces of PPARgamma.Cell,123(6)993-999.
- ^[12]Owens D R (2002) Thiazolidinediones: a pharmacological overview. Clin. Drug Invest., 22: 485-505.
- ^[13]Li NC, Lee A, Whitmer RA (2010): Use of angiotensin receptor blockers and risk of dementia in a predominantly male population: prospective cohort analysis. 340: b5465. 10.
- ^[14]Nicolakakis N, Aboulkassim T, Ongali B (2008): Complete rescue of cerebrovascular function in aged Alzheimer's disease transgenic mice by antioxidants and pioglitazone, a peroxisome proliferator-activated receptor gamma agonist. J Neurosci 28: 9287–9296.
- ^[15]Halvorsen B, Heggen E, Ueland T (2010): Treatment with the PPAR gamma agonist telmisartan downregulates interleukin-1 receptor antagonist in individuals with metabolic syndrome. Eur J Endocrinol 162: 267–273.
- ^[16]Zhang Q, Hu W, Meng B (2010): PPARgamma agonist telmisartan is neuroprotective after traumatic spinal cord injury via anti-inflammatory in adult rats. Neurol Res11,2:13-18.
- ^[17]Zhang X and Young H A (2002): PPAR and immune system- what do we know? Int.Immunopharmacol., 2:1029-44
- ^[18]Dharmendr S P, Jigna SS and Dhrabo J S (2011): Modulatory effect of telmisartan on anti-inflammatory of rosiglitazone in adjuvant arthritis model. Int j of Research in Pharmaceutical and Biomedical Sciences. 2(2),554-566.
- ^[19]Delerive P, Fruchart J, Staels B (2001): peroxisome proliferator-activated receptors in inflammation control. J Endocrinol.,169: 453-9.
- ^[20]Mohanty P, Aljada A, Ghanim H (2004): Evidencefor a potent anti-inflammatory inflammation control. J Endocrinol.,169: 453-9.
- ^[21]Goebel M, Clemenz M, Unger T (2006): Effective treatment of hypertension by AT(1) receptor antagonism: the past and future of telmisartan. Expert Rev Cardiovasc Ther, 4: 615–629.
- ^[22]Marketou M E, Kontaraki, J E, Tsakountakis NA (2011) :Differential effect of telmisartan and amlodipine on monocyte chemoattractant protein-1 and peroxisome proliferators activated receptor-gamma gene expression in peripheral monocytes in patients with essential hypertension. American Journal of Cardiology, 107, 1, 59–63.
- ^[23]Sukumaran V, Watanabe V K, Veeraveedu P T (2011) :Telmisartan ameliorates experimental autoimmune myocarditis associated with inhibition of inflammation and oxidative stress, European Journal of Pharmacology, vol. 652,. no. 1–3, pp. 126–135.
- ^[24]Moreira M, Matias JE, Souza CJ, Nicoluzzi JE, Caron PE, Repka JC(2011): Action of tacrolimus in arginine induced experimental acute pancreatitis. Rev Col Bras Cir.38: 260-265.
- ^[25]Sanchez-Hidalgo M, Martin AR, Villegas I, Alarcon de la Lastra C. (2007) Rosiglitazone, a PPARg ligand, modulates signal transduction pathways during the development of acute TNBS induced colitis in rats. Eur J Pharmacol 562: 247-258.
- ^[26]Winn-Deen ES, David H, Sigler G, Chavez R (1988): Development of a direct assay for alpha amylase. Clin.Chem.,(10):2005-2008
- ^[27]El-Shemi A G, Basalamah M A, Kensara O A and Ashshi A M (2011): Interleukin-22 therapy attenuates the development of acute pancreatitis in rats.Journal of Clinical Medicine and Research Vol. 3(6), pp. 82-88, 15..
- ^[28]Corti A,Fassino J,Marcucci F (1992): Oligometric tumor necrosis factor- α slowly converts into the reactive forms at bioactive levels. Biochem. J, 284:905-910.
- ^[29]Klinik M,Muller AM, Rose-John S (1998): Interlukin-6 and soluble interlukine -6 receptor: direct stimulation of gp130 and hemopoiesis. Blood,15(29):3495-3504.
- ^[30]Draper H, and Hadley M (1991): Malondialdehyde determination as index of lipid

peroxidation. In: Methods in Enzymology, Glazer PL (ed.), NewYork: Academic Press, P421-443.

- ^[31]Johansson LH and Borg LAH (1988):A spectrophotometric method for determination of catalase activity in small tissue sample. Anal. Biochem. , 174:331-336.
- ^[32]Ethridge RT, Chung DH, Slogof FM, Ehler RA, Hellmich MR, Rajaraman S, Saito H, Uchida T, Evers BM (2002): Cyclooxygenase-2 gene disruption attenuates the severity of acute pancreatitis and pancreatitis-associated lung injury. Gastroenterology, 123: 1311-1322.
- ^[33]Bhatia M, Wong FL, Cao Y, Lau HY, Huang J, Puneet P, Chevali (2005): Pathophysiology of acute pancreatitis. Pancreatology, 5: 132-144.
- ^[34]Hegyi P, Rakonczay JR, Sari R(2004): L-arginine induced experimental pancreatitis. World J.Gastroentrol., 10(4):2003-2009.
- ^[35]Takacs T ,Czako L, Morschel E, Laszlo F, , Tiszlavicz L,Rakonczay Z Lonovics J.(2002): the role of nitric oxide in edema formation in L-arginine induced acute pancreatitis. Pancreas 25 (3)277-82.
- ^[36]Hardman J, Shields C, Schofield D, Mc-Mahon R, Redmond HP and Siriwardena AK.(2005) : Intravenous antioxidant of end organ damage in Larginine induced experimental acute pancreatitis. Pancreatology 5 (4-5):380-6.
- ^[37] Biradar S and Veeresh B (2012): Screening of Natural Antioxidants by using L-Arginine induced acute Pancreatitis Model. International J Drug Development & Research 4(4)284-279.
- ^[38]Szabolcs A, Reiter RJ, letoha T, Hegyi P, Papai G, Varga I, Jarmay K, Kaszaki J,Sari R, Rakonczey Z Jr, Lonovics J, Takacs T.(2006): Effect of melatonin on the severity of in L-arginine induced experimental acute pancreatitis in rats. World J Gastroentrol 14;12 (2) 251-8.
- ^[39]Melo CM; CarvalhoKMMB; Neves JCDS; MoraisTC;RaoVS; SantosFA; BritoGADC and Chaves MH (2010):α,β amyrin a natural triterpenoid ameriolates L-arginine induced experimental acute pancreatitis in rats. World J.Gastroentrol.,16 (34) 4272
- ^[40]Sidhu S, pandhi P, MalhortaS, Vaiphei K, Khanduji KL(2010): Melatonin treatment is beneficial in pancreatic repair process after experimental acute pancreatitis. Eue J pharmacol.628,282
- ^[41]Young M. (1989): Acute diseases of the pancreas and biliary tract. Emerg Med Clin North Am 7: 555–573
- ^[42]Ventrucci M, Pezzilli R, Gullo L, Plate L, Sprovieri G, Barbara L(1989) Role of serum pancreatic enzyme assays in diagnosis of pancreatic disease. Dig Dis Sci; 34: 39-45.
- ^[43]Kazmierczak S, Catrou P, VanLente F(1993): Diagnostic accuracy of pancreatic enzymes evaluated by use of multivariate data analysis. Clin Chem 39: 1960–1965.
- ^[44]Esrefoglu M, Gul M, Ates B (2006): Antioxidative effect of melatonin, ascorbic acid and N-acetylcysteine on caerulein-induced. pancreatitis and associated liver. injury in rats.World J.Gastroenterol 12:259-264.
- ^[45]Rakonczay Z,Jarmay K,Kazzaki J (2003): NF-kappaβ activation is determined in arginine-induced acute pancreatitis. Free Radic Biol Med.,34:696-709
- ^[46]Pezzilli R, Ceciliato R, Barakat B, Corinaldesi R(2004): Immune-manipulation of the inflammatory response in acute pancreatitis. What can be expected? JOP 5:115-121.
- ^[47]Czako L, Takacs T, Varga RS, Hai DQ, Tiszlavicz L, Hegyi P, Tiszlavicz L, Matkovecs B, Lonovics J. (2000):Oxidative stress in distant organs and the effects of allopurinol during experimental acute pancreatitis. Int J Pancreatol 27(3)209-16
- ^[48]Schafer C, Tietz AB, Goke B. (2005): Pathophysiology of acute experimental pancreatitis: lessons from genetically engineered animal models and new molecular approaches. Digestion 71: 162-72.

^[49]Czako, L, Takacs T, Varga I, Tiszlavicz L, Hai D Q ,P Hegyi P, Matkovics B, Lonovics J (1998):. Involvement of oxygen derived free radicals in L-arginine induced acute pancreatitis. Dig. Dis. Sci., 43: 1770-1777.

- ^[50]Rau B, Poch B, Gansauge F, Bauer A, Nüssler AK, Nevalainen T, Schoenberg MH, Beger HG (2000): Pathophysiologic role of oxygen free radicals in acute pancreatitis: initiating event or mediator of tissue damage? Ann Surg; 231: 352-360.
- ^[51]Cuzzocrea S, Pisano B,Dugo L (2004): Rosiglitazone, a ligand of the PPAR-γ reduces acuta pancreatitis induced by cerulein. Care Med.,30: 951-956.
- ^[52]Ivashchenko CY, Duan SZ, Usher MG and MortensenRM (2007): PPAR-γ knockout in pancreatic epithelial cell abolishes the inhibitory effect of rosiglitazone on cerulein induced acute pancreatitis . Am J Physiol Gastrointest. Liver Physiol,239:319-326.
- ^[53]Sanchez-Hidalgo M, Martin AR, Villegas I, Alarcon de la Lastra C. (2005):Rosiglitazone, an agonist of peroxisome proliferator-activated receptor gamma, reduces chronic colonic inflammation in rats. Biochem Pharmacol; 69:1733-1744.
- ^[54]Shiojiri, Wadak, Nakajima, Katayama, Shibuya, Kudo, Kaowaki, Mayumi T, Yura Y, Kamisaki Y (2002): PPAR γ ligands inhibit nitrotyrosine formation and inflammatory mediator expressions in adjuvant-induced rheumatoid arthriti mice. Eur. J. Pharmacol. ;448:231–238.
- ^[55]Konturek PC, Dembinski A, Warzecha Z (2005): Pioglitazone, a specific ligand of peroxisome proliferator activated receptor-gamma, protects pancreas. against acute cerulein-induced pancreatitis. World J Gastroenterol;11: 6322-6329.
- ^[56]Ptak-Belowska A, Pawlik MW, Krzysiek-Maczka G, Brzozowski T, Pawlik W.W. (2007): Transcriptional up-regulation of gastrin in response to peroxisome Proliferator-activated receptor gamma agonist triggers cell survival pathways. J Physiol Pharmacol; 58(4): 793-80.
- ^[57]Chawla A, Schwarz EJ, Dimaculangan DD, Lazer MA. (1994): Peroxisome proliferator-activated receptor (PPAR) gamma: adipose-predominant expression and induction early in adipocyte differentiation. Endocrinology; 135: 798-800.
- ^[58]Rousseaux C, Lefebvre B, Dubuquoy L (2005): Intestinal anti inflammatory effect of 5-aminosalicylic acid is dependent on peroxisome proliferator-activated receptor gamma. J Exp Med; 201: 1205-1215.
- ^[59]Decker M, Hofflich H, Elias AN (2008): Thiazolidinediones and. Joosen AM, Bakker AH, Kersten S, Westerterp KR. The PPARgamma ligand rosiglitazone influences triacylglycerol metabolism in non-obese males, without increasing the transcriptional activity of PPARgamma in the subcutaneous adipose tissue. Br J Nutr; 99: 487-493.
- ^[60]Arita M, Yoshida M, Hong S (2005): Resolvin E1, an endogenous lipid mediator derived from omega-3eicosapentaenoic acid, protects against 2,4,6trinitro- benzene sulfonic acid-induced colitis. Proc Natl Acad Sci 102: 7671-7676.
- ^[61]Dworzanski T;Celinski K; Korolczuk A;Slomk M;Radej S;Czechowska G;Madro A;Cichoz-Lach H (2010): Influence of the PPARgamma agonist, rosiglitazone and antagonist, biphenol-A-diglicydyl ether (BADGE) on the course of inflammation in the experimental model of colitis in rat. J of physiology and pharmacology.16,6 683-693.
- ^[62]Nakai M, Sudo K, Yamada Y (2005): The role of the tumor necrosis factor receptor in 2,4,6-trinitrobenzene sulfonic acid (TNBS)-induced colitis in mice. Digest Dis Sci 50: 1669-1676.
- ^[63]Hollenbach E, Vieth M, Roessner A (2005): Inhibition of RICK/nuclear factor-kB and p38 signaling attenuates the inflammatory response in a murine model of Crohn disease. J Biol Chem; 280: 14981-14988.
- ^[64]Pereda J, Sabater L, Aparisi L, Escobar J, Sandoval J, Viña J, López-Rodas G, Sastre J. (2006):Interaction between cytokines and oxidative stress in acute pancreatitis. Curr Med Chem;13: 2775-2787.

^[65]Escobar J, Pereda J, Arduini A, Sandoval J, Sabater L, Aparisi L, López-Rodas G, Sastre J.(2009):Cross-talk between oxidative stress and pro-inflammatory cytokines in acute pancreatitis:a key role for protein phosphatases. Curr Pharm Des;15: 3027-3042.

^[66]Cuzzocrea S, Pisano B, Dugo L, Ianaro A, Patel NS, Di Paola R, (2003): Rosiglitazone and 15-deoxy-delta12,14-prostaglandin j2, ligands of the peroxisome proliferator-activated receptor-gamma (ppar-gamma), reduce ischaemia/reperfusion injury of the gut. Br J Pharmacol;140:366-376.

^[67]Villegas I, Martin AR, Toma W, de la Lastra CA(2004): Rosiglitazone, an agonist of peroxisome proliferator-activated receptor gamma, protects against gastric ischemia-reperfusion damage in rats: Role of oxygen free radicals generation. Eur J Pharmacol;505:195-203.

^[68]El-Moselhy M A and Nazmy W H(2011): Role of PPARgamma in gastric ulcerations induced by cold restrain stress in adult male albino rats. EL-minia Med Bull22,2,53-67.

^[69]Dharmendra S. Prajapati, Dr. Jigna S. Shah and Dr. Dhrubo Jyoti Sen (2011): Modulatory effect of telmisartan on anti-inflammatory .Effect of rosiglitazone in adjuvant arthritis model. International Journal of Research in Pharmaceutical and Biomedical Sciences Vol. 2 (2) Apr – Jun.

- ^[70]Bähr IN, Tretter P, Krüger J, Stark RG, Schimkus J, Unger T, Kapper K, Scholze J, Parhofer KG, Kintscher U (2011): . High-Dose Treatment With Telmisartan Induces Monocytic Peroxisome Proliferator-Activated Receptor-γ Target Genes in Patients With the Metabolic Syndrome Hypertension. 2011 Oct;58(4):725-32
- ^[71]Fliser D, Buchholz K, Haller H(2004):Antiinflammatory effects of angiotensin II subtype 1 receptor blockade in hypertensive patients with microinflammation. Circulation.; 110: 1103–1107.
- ^[72]Ichiki T ; Tian Q; Imayama I; Sunagawa K (2008): Telmisartan Manifests Powerful Anti-Inflammatory Effects Beyond Class Effects of Angiotensin IIType1 Blocker by Inhibiting Tumor Necrosis Factor-Induced Interleukin 6 Expressions through Peroxisome Proliferator Activated Receptor γActivation.Circulation;118 513
- ^[73]Waleed K.G. Al-Hejjaj, Intesar T. Numan, Raghdan Z. Al-Sa'ad, Saad A. Hussain (2011): Anti-inflammatory activity of telmisartan in rat models of experimentally-induced chronic inflammation: Comparative study with dexamethasone .Saudi Pharmaceutical Volume 19, Issue 1, January Pages 29–34.
- ^[74]Pang T, WangJ, Benicky J, Sánchez-Lemus E and Saavedra J M (2012): Telmisartan directly ameliorates the neuronal inflammatory response to IL-1β partly through the JNK/c-Jun and NADPH oxidase pathways Journal of Neuro inflammation, 9:102.
- ^[75]Yamada T, Kuno A, Masuda K, Ogawa K, Sogawa M, Nakamura S, Ando T, Sano H, Nakazawa T, Ohara H, Nomura T, Joh T, Itoh M. (2003): Candesartan, an Angiotensin II Receptor Antagonist, Suppresses Pancreatic. Inflammation and Fibrosis in Rats. J Pharmacol Exp Ther. ;307(1):17-23.
- ^[76]Staels B, Dallongeville J, Auwerx J, Schoonjans K, Leitersdorf E, Fruchart JC.
 (1998): Mechanism of action of brates on lipid and lipoprotein metabolism. Circulation; 98: 2088–2093.
- ^[77]Benson SC, Pershadsingh HA, Ho CI, Chittiboyina A, Desai P, Pravenec M, Qi N, Wang J, Avery MA, Kurtz TW (2004):Identification of telmisartan as a unique angiotensin II receptor antagonist with selective PPARγ-modulating activity. Hypertension; 43: 993–1002.
- ^[78]Schupp M, Janke J, Clasen R, Unger T, Kintscher U (2004): Angiotensin type receptor blockers induce peroxisome proliferator-activated receptor-γ activity. Circulation; 109: 2054–2057.
- ^[79] Schupp M, Clemenz M, Gineste R, Witt H, Janke J, Helleboid S, Hennuyer N,

Ruiz P, Unger T, Staels B, Kintscher U(2005) :Molecular characterization of new selective peroxisome proliferator-activated receptor- γ modulators with angiotensin receptor blocking activity. Diabetes. 54: 3442–3452.

- ^[80]Chan WP, Fung ML, Nobiling R, and Leung P (2000): Activation of local renin-. angiotensin system by chronic hypoxia in rat pancreas. Mol Cell Endocrinol 160: 107–114.
- ^[81]Leung PS, Chan HC, Fu LX, and Wong PY. (1997): Localization of angiotensin II receptor subtypes AT1 and AT2 in the pancreas of rodents. J Endocrinol 153: 269–274.
- ^[82]Leung PS, Chan WP, Wong TP, and Sernia C. (1999):Expression and localization of the renin-angiotensin system in the rat pancreas. J Endocrinol160: 13–19.
- ^[83]Tsang SW, Ip SP, and Leung PS.(2004): Prophylactic and therapeutic treatments with AT1 and AT2 receptor antagonists and their effects on changes in the severity of pancreatitis. Int J Biochem Cell Biol 36: 330–339.
- ^[84]Tsang SW, Ip SP, Wong TP, Che CT, and Leung PS (2003):Differential effect of saralasin and ramiprilat, the inhibitors of renin-angiotensin system, on ceruleininduced acute pancreatitis. Regul Pept 28: 47–53.
- ^[85]Nagashio Y, Hiroshi Asaumi H, Watanabe S, Nomiyama Y,Taguchi M, Tashiro M, Sugaya T and Otsuk M . (2004): Angiotensin II type 1 receptor interaction is an important regulator for the development of pancreatic fibrosis in mice .Am J Physiol Gastrointest Liver Physiol 287: G170–G177.
- ^[86]Fisher AA and Basset ML(2002): acute pancreatitis associate with angiotensinII receptor antagonists. Ann Pharmacother.36:1883-6.
- ^[87]Balani AR and Grendell JH (2008):Drugs induced pancreatitis: incidence, management and prevention. Drug Saf. 31:823-37. and prevention. Drug Saf. 31:823-37.