



Journal Homepage: - www.journalijar.com
**INTERNATIONAL JOURNAL OF
 ADVANCED RESEARCH (IJAR)**

Article DOI: 10.21474/IJAR01/5765
 DOI URL: <http://dx.doi.org/10.21474/IJAR01/5765>



RESEARCH ARTICLE

EFFECT OF SEASONAL TEMPERATURE VARIATION ON THE DURATION OF LIFE CYCLE STAGES OF THE FLY OF FORENSIC IMPORTANCE, *PARASARCOPHAGA DUX* (THOMSON) (DIPTERA: SARCOPHAGIDAE).

Shabnamnaz Siddiki and S. P. Zambare.

Department of Zoology, Dr. Bababsheb Ambedkar Marathwada University, Aurangabad, (M.S.) India.

Manuscript Info

Manuscript History

Received: 04 September 2017
 Final Accepted: 06 October 2017
 Published: November 2017

Key words:-

Forensic entomology, *Parasarcophaga dux* (Thomson), fluctuate temperature, life cycle duration, seasons.

Abstract

Sarcophagid flies belonging to carrion insect communities provide the information about the corps. *Parasarcophaga dux* (Thomson) (Diptera: Sarcophagidae) collected from Latur district of Maharashtra (India) has been studied for the duration of different life cycle stages in three different seasons i.e. rainy, winter and summer seasons under the room temperature. The female released 1st instar larvae which after two larval mounds undergo prepupation and then pupation in the soil. The adult emerged out from pupae.

During rainy season the life cycle duration from larvaeposite to emergence of adult was 315.15 ± 3.28 hrs, during winter season in 359.56 ± 2.44 hrs, while in summer season it took 265.51 ± 6.28 hrs, under the fluctuating diurnal temperature and humidity of the laboratory.

The time durations of life cycle stages of *P. dux* in the different seasons can be used to determine the PMI using the meteorological data and *P. dux* life cycle stages collected from the cadaver.

Copy Right, IJAR, 2017.. All rights reserved.

Introduction:-

The more than 2600 species [1] of flesh flies (Family – Sarcophagidae) reported belongs to 1000 genera all around the world. In India the family Sarcophagidae represents 117 species and 38 genera of three subfamilies. The genus *Sarcophaga* includes more than 22 recorded species belonging to 6 subgenera. Most flesh flies are parasites or predators with relatively small numbers of species with a true preference for vertebrate carrion [2-3]. Flesh flies are usually considered as a taxon of high forensic importance [4-5]. However their participation in carrion insect community was tasted almost exclusively in experiments with small sized carrion [6-11].

Sarcophagidae are considered to be unimpeded by rain and to fly despite the weather [12]. The females deposit the larvae in the safe nutritious location [13]. As a result flesh flies may be the initial colonizers of the body outdoors, if there is long period of rainy weather. Despite this, many flesh flies prefer sunlight rather than shaded condition [4] although *Sarcophaga* (Subgenus – *Robineauella*) *caerulescens* is considered a shade lover [14].

The time taken by *Sarcophaga* larvae to emerge as an adult is considered to be between 8 – 15 days [15]. Kamal (1958) provide an indications of life cycle duration of various stages of American Sarcophagid species. Development time to adult emergence varied by as much as 252 hours [16] depending on the temperature.

Corresponding Author:- Shabnamnaz Siddiki.

Address:- Department of Zoology, Dr. Bababsheb Ambedkar Marathwada University, Aurangabad, (M.S.) India.

Therefore, this provides a source of variation and limitations in the use of this species to calculate the time since death.

The present research work was carried in the laboratory in Latur district, Maharashtra, India to study the time duration of different stages of the life cycle of *Parasarcophaga dux* (Thomson) in different seasons.

Material and method:-

The maggots were collected from dead dog from Latur district (MS), India and were reared in laboratory in rearing chamber on fresh chopped buffalo liver and sugar syrup on cotton. These chambers were covered with a fine muslin cloth in order to facilitate fresh air and to prevent the entry of parasites. These chambers were maintained under natural temperature and humidity condition. Soil was also supplied for pupation. The maggots were observed throughout their developmental stages at different time interval on hourly basis. After adult emergence the flesh flies were identified and were to be *P. dux* and some Calliphorids. Adult male and female *P. dux* were maintained in insect cages with fresh liver every day. The Ist instar larvae released on liver were monitored on hourly basis to find the duration with reporting of temperature. This experiment was repeated in three times in three different seasons i.e. rainy season, winter season and summer season.

Observation and result:-

In the present study the life cycle stages of *Parasarcophaga dux* (Thomson) are larvae (i.e. Ist instar, IInd instar and IIIrd instar), pre-pupa, pupa and adult. The female releases the Ist instar larvae directly on the liver. The duration of life cycle stages changed as per the temperature variations in different seasons.

1. Rainy season:

In rainy season it has been observed that the time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) was completed in 315.15 ± 3.28 hours from larviposite up to emergence of adult fly at the average temperature ranging from $23 - 26.7$ °C and average humidity ranging from $28.5 - 58.6\%$. It was observed that the time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) was 25.10 ± 1.13 hours, 27.45 ± 1.22 hours, 43.28 ± 5.25 hours, 23.47 ± 3.10 hours and 195.05 ± 6.39 hours for Ist instar, IInd instar, IIIrd instar, pre-pupa and pupa respectively (Table 1).

Table 1:- Time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) during rainy season

life stages	PMI (H:MM)	Average temperature (°C)	Average humidity (%)
Larva	I st instar	$25:10 \pm 1:13$	25.5 ± 1.1
	II nd instar	$27:45 \pm 1:22$	26 ± 1.04
	III rd instar	$43:28 \pm 5:25$	26.7 ± 0.7
Pre-pupa	$23:47 \pm 3:10$	23 ± 0.9	51.5 ± 1.03
Pupa	$195:05 \pm 6:39$	23.4 ± 1.2	58.6 ± 1.4
Total duration	$315:15 \pm 3:28$		

(± indicates standard deviation of three values)

2. Winter season:

In winter season the time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) was completed in 359.56 ± 2.44 hours from larviposite to emergence of adult fly at the average temperature ranging from $18 - 22.7$ °C and average humidity ranging from $44 - 68.8\%$. The time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) was 26.50 ± 2.04 hours, 28 ± 4.12 hours, 45.55 ± 3.21 hours, 24.05 ± 2.19 hours and 235.06 ± 3.02 hours of Ist instar, IInd instar, IIIrd instar, pre-pupa and pupa respectively (Table 2).

Table 2:- Time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) during winter season

life stages	PMI (H:MM)	Average temperature (°C)	Average humidity (%)
Larva	I st instar	$26:50 \pm 2:05$	18.5 ± 1.7
	II nd instar	$28:00 \pm 4:12$	18 ± 1.4
	III rd instar	$45:55 \pm 3:21$	22.7 ± 2.2
Pre-pupa	$24:05 \pm 2:19$	20.5 ± 1.5	66.5 ± 0.2
Pupa	$235:06 \pm 3:02$	20.3 ± 1.8	44 ± 1.2
Total duration	$359:56 \pm 2:44$		

(± indicates standard deviation of three values)

3. Summer season:

In summer season, the time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) was completed 260.51 ± 6.28 hours from the larviposite to emergence of adult fly at the average temperature ranging from 31.5 to 33.7°C and average humidity 13.6 – 20.5 %. The time duration of different lifecycle stages of *Parasarcophaga dux* was 22.51 ± 1.52 hours, 24.30 ± 1.71 hours, 26.45 ± 4.43 hours, 18.39 ± 2.52 hours and 168.06 ± 2.17 hours of Ist instar, IInd instar, IIIrd instar, pre-pupa and pupa respectively (Table 3)

Table 3:- Time duration of different life cycle stages of *Parasarcophaga dux* (Thomson) during summer season

life stages	PMI (H:MM)	Average temperature (°C)	Average humidity (%)
Larva	I st instar	22:51 ± 1:52	31.5±0.8
	II nd instar	24:30 ± 1:17	31.7±1.1
	III rd instar	26:45 ± 4:48	31.6±1.1
Pre-pupa	18:39 ± 2:52	33±0.5	20.5±0.5
Pupa	168:06 ± 2:17	33.7±0.7	17.7±0.7
Total duration	260:51 ± 6:28		

(± indicates standard deviation of three values)

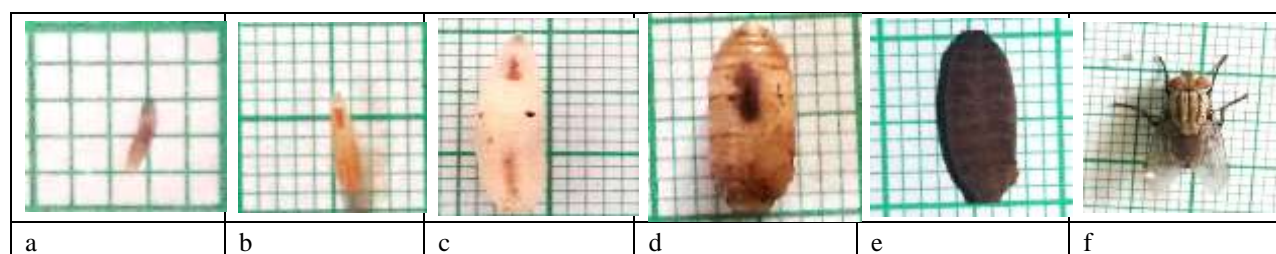


Fig.1:- Life cycle stages of *Parasarcophaga dux* (Thomson). a. Ist instar larvae; b. IInd instar larvae; c. IIIrd instar larvae; d. Pre-pupa; e. Pupa; f. Adult fly.

Discussion:-

Species of flesh flies larvae are very similar in appearance and traditionally have been difficult to identify [4]. Therefore accurate identification of either larvae or adults has had to be made by taxonomic specialists [17]. Taxonomically *P. dux* has been confused with *Parasarcophaga misera* by many previous authors [4], but more recent research has a clarified classification of these two species as distinct species and separated them based on adult males [18-20].

It was observed that at higher temperature and humidity the maggots emerged faster than lower temperature and higher humidity and increase in temperature will increase the rate of metabolism [21]. An earlier study done on *P. terraenovae* it was found that fluctuating temperature actually lade to significant faster rate of development [22]. In present study it was observed that IIIrd instar larvae took higher time period than Ist and IInd instar larvae. It was also recorded that IInd and IIIrd instars took prolonged time for development as compare to first instar larvae [23-24].

The time taken by *Sarcophaga* (Subgenus Robineauilla) *caerulescens* larvae to emerge as an adult is considered to be between 8-12 days [15]. An indication was also provided on the lifecycle duration of various stages of the American *Sarcophagid* species [25]. However different observations have been reported for the developmental rates of the *S. dux*. Life cycle of *Sarcophaga dux* from the first instar to the adult was 312.0 ± 3.0 h under uncontrolled indoor temperature in Malaysia [26]. Report from study in north Thailand in the year 2002-2003 under natural temperature indicates seasonal variation in development of fly with rapid larval development in summer at around 72 hrs, while it took 72-96 hrs.in rainy season and 96 hrs.in winter [27-28]. The life cycle of *Sarcophaga cultellata* at 25°C and 50% humidity showed a total development time from larvae position to first adult emergence was 330 ± 12 hr [29]. The only report from India on the development of larvae of *Sarcophaga(L) tibialis* raised on chicken liver indicated maximum development between 15-30°C [30-35].

The present results are similar with the developmental duration of *Chrysomya megacephala* (Diptera: Calliphoridae), in rainy season and low constant temperature of 10°C and humidity 19%, the total life cycle duration in rainy season was completed in 265 h ± 2 h (11.04 days ± 0.08 days) when the temperature ranged between 26°C and 29°C and humidity ranged between 35% and 50%, while in the low constant temperature 10°C ± 0.5°C the life cycle was completed in 609h ± 4 h (25.38 days ± 0.16 days) indicating a delay in the life cycle by 14.37 days ± 0.13 days [36]. Similarly our current results are in agreement with another study on the effect of different temperature and humidity on the lifecycle duration and morphological parameters of *Chrysomya rufifacies* (Diptera: Calliphoridae) in different seasons. It was reported that the life cycle of *Chrysomya rufifacies* in summer was completed in 241 ± 2.17 h (10.04 days ± 0.12 days) when the temperature ranged between 30.1°C and 37.2°C, but in the rainy season it was completed in 275 h ± 2.27 h (11.46 days ± 0.45 days), when the temperature ranged between 26.2°C and 30.1°C, while in winter the life cycle was completed in 318h ± 2.45 h (13.25 days ± 0.25 days) when the temperatures ranged between 26.4°C and 18.2°C respectively [37].

In previous study in laboratory, at the fluctuating temperature during rainy season, winter season and summer season, *Chrysomya megacephala* took 237 hours 47 minutes, 263 hours 51 minutes and 211 hours 13 minutes respectively. Whereas the *Chrysomya rufifacies* took 239 hours 14 minutes, 286 hours 02 minutes and 216 hours 26 minutes during rainy season, winter season and summer season respectively [38].

Conclusion:-

It is observed that developmental time of different stages of life cycle of *Parasarcophaga dux* from larviposite to emergence of adult differ in various seasons. On the basis of result it was clearly seen that a changes in temperature and humidity bring about a significant changes in developmental rate of the larval stages. In the condition with the higher temperature, larvae developed quickly and matured into pupa as compared to that grown in cooler temperature.

Acknowledgement:-

Authors are thankful to the Department of Zoology, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (MS), India for providing the laboratory facilities to carry out this research work.

References:-

1. Pape T (1996): Catalogue of the Sarcophagidae of the world (Insecta: Diptera). Mem Entomol Int 8:1–558
2. Povolny, D. and Verves, Y. (1997): The flesh-flies of Central Europe. Spixiana, Supplement 24, Munchen
3. Richet, R., Blackith R.M., Pape, T. (2011): Sarcophaga of France (Diptera:Sarcophagidae). Pensoft Series Faunistica, Sofia
4. Smith KGV (1986): A manual of forensic entomology. The Trustees of the British Museum, London.
5. Byrd, J.H. and Castner, J.L. (2009): Forensic Entomology: the utility of arthropods in legal investigations. CRC Press, Boca Raton.
6. Denno, R.F. and Cothran, W.R. (1976): Competitive interactions and ecological strategies of Sarcophagid and Calliphorid flies inhabiting rabbit carrion. Ann Entomol Soc Am 69(1):109–113
7. Hanski, I. (1976): Breeding experiments with carrion flies (Diptera) in natural conditions. Ann Entomol Fenn 42:113–121
8. Hanski, I. (1987): Carrion fly community dynamics: patchiness, seasonality and coexistence. Ecol Entomol 12:257–266. doi:10.1111/j.1365-2311.1987.tb01004.x
9. Hanski I, Kuusela S (1980): The structure of carrion fly communities: differences in breeding seasons. Ann Zool Fenn 17:185–190
10. Kuusela, S. and Hanski, I. (1982): The structures of carrion fly communities: the size and the type of carrion. Ecography 5(4):337–348. doi:10.1111/j.1600-0587.1982.tb01048.x
11. Blackith, R.E. and Blackith, R.M. (1990): Insect infestations of small corpses. J Nat Hist 24(3):699–709. doi:10.1080/00222939000770481
12. Erzinclioglu, Z. (2000): Maggots, murder and men: memories and reflections of forensic entomologists. Harley books, 256 pp.
13. Archer, M. S. and Elgar, M. A. (2003): Yearly activity pattern in southern Victoria (Australia) of seasonally active carrion insects. Forensic Science International 132 (3): 173-176.
14. Singh, D. and Bharti, M. (2008): Some notes on the nocturnal larviposition by two species of Sarcophaga (Diptera: Calliphoridae). Forensic Science International 177: 19-126.

15. Pohjoismäki JL, Karhunen PJ, Goebeler S, Saukko P, Sääksjärvi IE (2010): Indoors forensic entomology: colonization of human remains in closed environments by specific species of sarcosaprophagous flies. *Forensic Sci Int* 199(1):38–42. doi:10.1016/j.forsciint.2010.02.033
16. Byrd, J.H. and Bulter, J.E. (1998): Effect of temperature on *Sarcophaga haemorrhoidalis* (Diptera: Sarcophagidae) development. *Journal of Medical Entomology* 35: 694-698.
17. Zumpt, F., (1965): Myiasis in man and animal in the old world, Butterworths, London.
18. Sugiyama E, Shinonaga S, Kano R. (1988): Sarcophagine flies from Nepal with the description of a new species (Diptera: Sarcophagidae). *Jpn J Sanit Zool.*; 39:355–362.
19. Kano, R. and Shinonaga, S. (1994): Studies on the Sarcophagid flies from Nepal (Diptera: Sarcophagidae). *Jap. J. Sanit. Zool.*, 45: 253-275.
20. Greenberg, B. and Kunich, J.C. (2002): *Entomology and the law. Flies as forensic indicators.* Cambridge University Press: Cambridge; pp 249-283.
21. Gomes, L., Gomes, G., Van ZCJ (2009): The influence of temperature on the behavior of burrowing in larvae of the blowflies, *Chrysomya albiceps* and *Lucilia cuprina*, under controlled conditions. *J Insects Sci.* 9: 14.
22. Davies, L. and Ratcliffe, G. (1994): Developmental rate of some pre-adult stages in blowfly with reference to low temperatures. *Med. Vet. Entomol.* 8:245-254.
23. Nabity, P.D., Higley, L.G., and Heng-Moss, T.M. (2006): Effect of temperature on development of *Phormia regina* (Diptera: Calliphoridae) and use of developmental data in determining time intervals in forensic entomology. *Journal of Medical Entomology* 43: 1276-1286.
24. Bharti, M., Singh, D. and Sharma, Y. P. (2007): Effect of temperature on the development of forensically important blowfly *Chrysomya megacephala* (Fab.) (Diptera: Calliphoridae) *Entomon*, 32(2): 149-151.
25. Kamal, A.S. (1958): Comparative study of thirteen species of Sarcosaprophagous Calliphoridae and Sarcophagidae (Diptera). *Binomics. Annals of the Entomological Society of America.* 51(3): 261-271. doi: 10.1093/aesa/51.3.261.
26. Jeffrey, D., Wells, and Joshua, L. and Smith (2013): First Report of *Blaesoxiphaplin thopyga* (Diptera: Sarcophagidae) from a Human Corpse in the U.S.A. and a New State Geographic Record Based on Specimen Genotype. *J Forensic Sci.*; 58(5): 1378-80.
27. Sukontason K, Sukontason KL, Piangjai S, Chaiwong T, Boonchu , Vogtsberger, R.C. (2003): Larval ultrastructure of *Parasarcophaga dux* (Thomson) (Diptera: Sarcophagidae). *Micron.* 34(8): 359-364.
28. BanzigerH, and Pape T. (2004): Flowers; Faeces and cadavers: natural feeding and laying habits of flesh flies in Thailand (Diptera: Sarcophagidae, Sarcophaga spp.). *Journal of Natural History*; 38:1677-1694.
29. Kumara TK, Hassan AA, Salmah MR, Bhupinder S. (2013): Larval growth of *Liosarcophaga dux* Thompson (Diptera: Sarcophagidae) under uncontrolled indoor temperatures in Malaysia. *Southeast Asian J Trop Med Public Health*; 44(2):182-7.
30. Sharma.R, and Mitra, B. (2014): Check-list of Indian flesh flies (Insecta: Diptera: Sarcophagidae). *Zsigov in/checklist/Indian_Sarcophagidae.pdf*.
31. Sukontason, K.L., Sanit, S., Klong-klaew, T., Tomberlin, J.K. and Sukontason, K.(2014): *Sarcophaga (Liosarcophaga) dux* (Diptera: Sarcophagidae): A flesh fly species of medical importance. *Biological Research*.47: (14) 1-9.
32. Nandi, B.C. (2002): *Fauna of India: Diptera: Sarcophagidae.* Published by Zoological Survey of India. 10: 270-274.
33. Heo, C.C., Latif, B.,Kurahashi, H., Nazni, W.A. and Omar,B. (2011): Oviposition of forensically important dipterans on a high-rise building, with a new host record of parasitoids in Malaysia (abstract), 9th Annual Conference of North American Forensic Entomology Association, College Station, Texas. Ref Type: Conference Proceeding.
34. Mercedes, D.D. and Hardy, D.E. (1977): A catalog of the Diptera of the Oriental region, Suborder Brachycera through Division Aschiza, Suborder Cyclorhapha: The University Press of Hawaii, Honolulu Vol-II: 557-583.
35. Senior-White, R., Aubertin, D. and Smart, J. (1940): The fauna of British India, including the remainder of the Oriental region, Diptera. Family Calliphoridae. Vol. VI. London: Taylor and Francis 6 208-281.
36. Abd-ALGalil FM, Zambare SP (2015): Effect of temperature on development of Calliphorid fly of forensic importance *Chrysomya megacephala* (Fabricius, 1794). *Indian J Appl Res* 5: 767-769.
37. Abd-ALGalil FM, Zambare SP (2015a): Effect of temperature on development of Calliphorid flies of forensic importance, *Chrysomya rufifacies* (Macquart, 1842). *IJAR*: 1099-1103.
38. Siddiki S, Zambare SP (2017). Studies on time duration of life stages of *Chrysomya megacephala* and *Chrysomya rufifacies* (Diptera: Calliphoridae) during different seasons. *J Forensic Res* 8:379. doi:10.4172/2157-7145.1000379.