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### RESEARCH ARTICLE

#### COMPARISON OF LABORATORY METHODS FOR THE DIAGNOSTIC OF TRICHOMONASIS.

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#### Abstract

In Mongolia, there is a variable number of *Trichomonas vaginalis* (*T.vaginalis*) infections reported and therefore we need a detection tool which is highly sensitive and specific. At the present study, we compared wet mount, gram stain, culture and PCR for detection of *T.vaginalis* from 109 samples collected in "UlaanTuuz" hospital in NCCID. As a result, 21.1%(23/109) were positive by wet mount, 18.3%(20/109) were positive by gram stain, 28.4%(31/109) were positive by culture and 36.6%(40/109) were positive by PCR respectively as compared to other methods. Further *T.vaginalis* trophozoite isolate was used to determine PCR sensitivity. Trophozoite isolate was counted by using haemocytometer and  $10.1 \times 10^4$  was counted in one mL diluted swab sample. From that sample, trophozoite isolate was diluted 3156, 1578, 100, 50, 12, 3, 1.5 (1-2) per mL sample. PCR was performed serially diluted samples and as a result, all the samples had 102bp *T.vaginalis* specific DNA band and confirmed PCR method was sensitive. Out of 109 samples 17 (15.5%) were positive by all detection methods. Since PCR had highest number of positivity, we have used it as a "golden standart" to calculate sensitivity and specificity of these methods. PCR had 100% specificity as compared to 98% wet mount, 98.5% gram stain and 100% culture. On the other hand culture had the highest sensitivity 77.5% as compared to PCR 100%, wet mount and gram stain 47.5-77.5%.

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#### Introduction:-

*Trichomonas vaginalis* commonly causes vaginitis and perhaps cervicitis in women, as well as urethritis in both sexes, 10-50% of those cases are clinically asymptomatic. Worldwide, more than 180 million people get the infection with this parasite annually. Statistics from NCCID reports every 4-6 (48-63.6% in 10000 population) people are infected with sexually transmitted diseases and *T.vaginalis* accounts 16.7-39.5% of those cases. Various methods have been used for the diagnosis of trichomoniasis such as wet mount, culture, Papanicolaou smear, and serologic test. Wet mount examination is an easy, simple, and rapid method but more than  $10^3$ /ml and live protozoa are required for detection. Culture demands a specialized medium and takes 2-5 days for the diagnosis. *T.vaginalis* is by indirect immunofluorescence test is the lack of sensitivity and specificity. In recent years, molecular biological

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techniques have a sexually transmitted infection disease. Therefore highly sensitive and specific clinical diagnostic methods needed for detection of *T.vaginalis*.

### Materials and methods:-

#### Patients and diagnostic test:-

Total 109 female patients swab and vaginal secretions were employed in this study. All the patients had informed consent at "Ulaan tuuz" voluntary clinic in NCCID hospital. All samples were tested the presence of *T.vaginalis* by wet smear, gram staining, culture by Feinberg-Whittington medium and PCR using TV-650 primer. Further *T.vaginalis* trophozoite isolate was used to determine PCR sensitivity. Trophozoite isolate was counted by using haemocytometer and  $10.1 \times 10^4$  was counted in one mL diluted swab sample. From that sample, trophozoite isolate was diluted 3156, 1578, 100, 50, 12, 3, 1.5 (1-2) per mL sample. PCR was performed serially diluted samples and as a result, all the samples had 102bp *T.vaginalis* specific DNA band and confirmed PCR method was sensitive. Sensitivity and specificity of clinical diagnostic method were determined by chi square test (SPSS-17).

#### Result:-

*T.vaginalis* was detected in culture, direct wet smear, Gram staining, and PCR techniques as follows, 28.4% (31/109), 21.1% (23/109), 18.3% (20/109) and 36.6% (40/109), respectively. Total 15.5% (17/109) samples were examined positive with *T.vaginalis*. PCR amplification method had highest sensitivity and specificity and therefore set as the "gold standard". In PCR TV-650 primer set used which produces single band with 330bp length. The specificity of the direct wet smear was 79%, gram staining was 76% and culture was 88% as compared to PCR amplification method. The sensitivity of the direct wet smear was 95%, gram stain was 95% and culture was 100% and as compared to PCR amplification method. *T.vaginalis* isolate was prepared from culture positive samples where sub-cultured in Trypticase-yeast extract-maltose, Meingassner and Heyworth medium. *T.vaginalis* trophozoite in culture was counted and diluted to make 3156, 1578, 100, 50, 12, 3, 1.5 cells per milliliter and DNA was extracted and subject to PCR amplification by TV-A5, TV-A6 primer sets (102 bp band). The highest dilution, 1.5 cells per milliliter Heyworth medium was positive by PCR. There was no correlation observed between clinical outcome and laboratory detection ( $r=0.001$ ).

#### Conclusions:-

1. We diagnosed *T.vaginalis* new cases by different clinical laboratory methods wet smear (21.1%), and gram stain (18.3%) and culture (28.4%) and PCR, respectively.
2. Sensitivity of the PCR assay is 100% for detection of *T.vaginalis* and culture is 100% specific, 77.5% sensitive and kappa coefficient is much more significant. Sensitivity of Gram stain and wet mount are 47.5-55.0% and specificity is 98-98.5% and kappa coefficient is also significant.

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