

1 DIFFERENTIAL ANALYSIS OF GENE EXPRESSION IN SEVERELY SICK COVID-19 2 PATIENTS.

3 **Background:**

4 This study investigates differential gene expression in severely sick COVID-19 patients to
5 elucidate molecular mechanisms driving disease progression. Using transcriptomic data from
6 44 severe COVID-19 patients and 10 healthy individuals (NCBI GEO: GSE171110), we
7 analyzed whole blood samples via DESeq2, identifying 737 differentially expressed genes
8 (DEGs). Of these, 662 were upregulated (e.g., inflammatory and antiviral pathways) and 75
9 downregulated. Principal component analysis (PCA) and heatmaps revealed distinct
10 transcriptional signatures between groups, highlighting immune dysregulation. Key
11 upregulated pathways included Jak-STAT, MAPK, PI3K-Akt, Toll-like receptor, and TNF
12 signaling, associated with cytokine storms, neutrophil activation, and oxidative stress. MHC
13 class I genes (linked to CD8+ T-cell and NK-cell cytotoxicity) were elevated, while MHC class
14 II (involved in CD4+ T-cell cytokine production) was suppressed. Pro-inflammatory
15 interleukins (IL-1 β , IL-6, IL-12) and antiviral IFN- β were also upregulated, indicating
16 heightened inflammation and antiviral responses. Conversely, DNA repair pathways were
17 disrupted. The findings suggest severe COVID-19 involves hyperactivation of innate
18 immunity and cytotoxic T-cell responses, coupled with impaired adaptive immunity (reduced
19 CD4+ T-cell function). These mechanisms likely contribute to tissue damage and cytokine
20 release syndrome. The study underscores the role of transcriptional dysregulation in driving
21 severe outcomes, offering insights into potential therapeutic targets to modulate immune
22 responses.

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24 **Introduction:** Covid-19 caused by the novel coronavirus (SARS-CoV-2), Over 760 million
25 cases and 6.9 million deaths have been recorded worldwide since December 2019, but the
26 actual number is thought to be higher (1). Patients with this disease have very variable clinical
27 pictures, ranging from the absence of symptoms to respiratory deficiency, which progressively
28 leads to death. Studies have postulated that the immune response is fundamental in the
29 progression of this disease. When the immune system fails to control the virus efficiently in
30 the acute stage, a macrophage activation syndrome can develop that causes the flow of a
31 cascade of cytokines that lead the patient to a critical or severe state (2). However, it is still
32 not completely known how these mechanisms are carried out, nor the variables that trigger
33 the severe condition, which occurs in a small percentage of these patients (3). Therefore, for

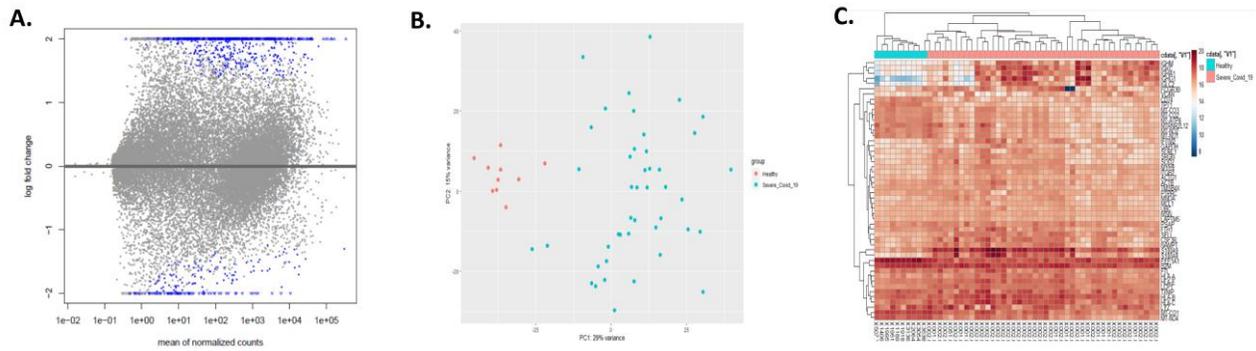
34 a greater understanding of the disease it is necessary to study the cellular pathways and the
35 transcriptional response to infection in the host cells. In order to clarify the signaling pathways
36 affected in the severe state of patients, the present work proposes the analysis of the
37 transcriptome of whole blood samples from patients with severe COVID-19 in comparison
38 with apparently healthy individuals.

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40 **Objective:** To analyze the transcriptional profile in whole blood samples from healthy
41 individuals and patients with severe Covid-19 to compare and analyze the pathways involved
42 in patients with this disease.

43 **Methodology:** Transcriptomic data were obtained in the form of pre-processed count
44 matrices (aligned and filtered readings, not normalized) from the NCBI Gene Expression
45 Omnibus (GEO) platform (<https://www.ncbi.nlm.nih.gov/geo/>. Accession number: GSE171110)
46 (4), which were represented by 54 samples, of which 44 were from patients with severe
47 Covid-19 and 10 healthy individuals. The counting matrix was imported by the
48 DESeqDataSetFromMatrix function of the DESeq2 package (v1.32.0), it contained 30,185
49 transcribed genes. All normalized transcripts with a maximum mean of all rows less than 10
50 were excluded, resulting in 24,099 transcripts present. And again with DESeq2, the set of
51 differentially expressed genes was calculated (severe COVID-19 versus Healthy Individuals),
52 the p-value used was 0.05, and a FC of 1. The variance stabilization transformation (VST)
53 and Log2 were used for the calculation of normalized counts of each transcript. With all the
54 transcripts present, a principal component analysis was performed. The most variable
55 transcripts were visualized on a heat map. And an analysis of differentially expressed
56 pathways was performed using the KEGG package.

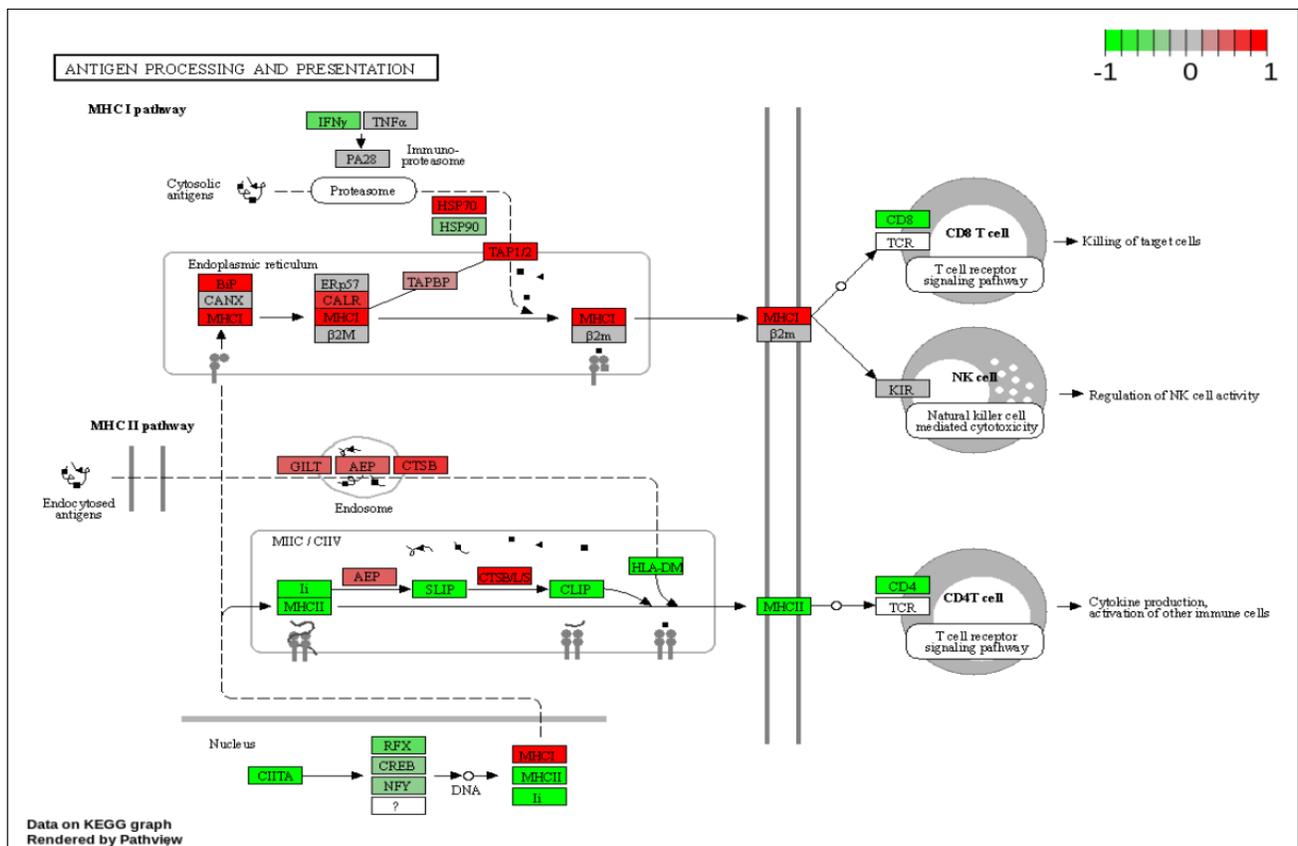
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58 **Results and Discussion:** The differential expression analysis for the 24,092, with p-value < 0.05 and
59 FC = 1, yielded a total of 737 differentially expressed genes (Figure 1A), of which 662 (2.7%) were
60 positively regulated (LFC > 1.00), while 75 (0.31%) were negatively regulated (LFC < -1.00). Both in
61 the analysis of main components (Figure 1B) and in the heat map (Figure 1C) it can be observed that
62 the samples of patients with "Severe COVID-19" present a differentially expressed gene signature
63 different from that of the samples provided by the "Healthy Individuals".

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Figure 1. A. log₂ shift volcano diagram, differentially expressed genes are shown in blue color. B. Main component graph C. Heat map of differential gene expression between COVID-19 patients and healthy donors (each column represents a different sample, the rows represent different genes; the color represents the level of gene expression: red indicates a positive gene expression in the sample and blue indicates a negative gene expression in the sample.



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Figure 2. Antigen Processing and Presentation Pathways.

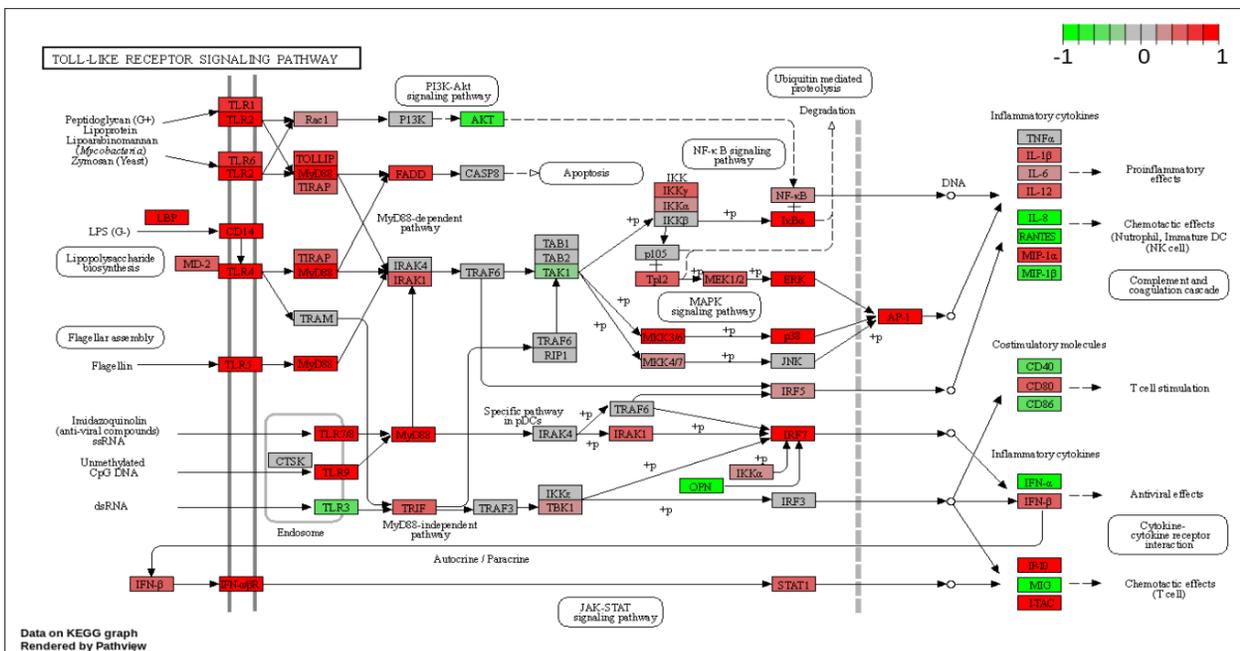
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77 Through the analysis of differentially expressed pathways, it was observed the positively
78 regulated expression of the Jak-STAT, MAPK, PI3K-Akt, and PLC γ signaling pathways,
79 activated by the chemokine/cytokine receptor, so that cellular functions such as proliferation,
80 cell differentiation, apoptosis, cell migration, and ubiquitination-mediated proteolysis are
81 altered. There is production of reactive oxygen species and which in turn produces
82 dysregulation of the DNA mismatch repair (MMR) system (5).

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84 At the level of immunity, positively regulated expression of the MHC I was observed, which
85 triggers the CD8 T lymphocyte response and regulates the activity of NK cells, which function
86 as inducers of cell death. On the other hand, the MHC II showed negatively regulated
87 expression, as well as the activity of CD4 T lymphocytes, which act in the production of
88 cytokines (Figure 2). In addition, there is a strong regulation of positive expression in genes
89 associated with the inflammatory response, which includes the Toll-Like receptor signaling
90 pathway, the TNF signaling pathway, pro-inflammatory interleukins: IL-1b, IL-6, IL-12, which
91 also plays an important role in the recruitment, function and survival of neutrophils (6). On the
92 other hand, positive expression of IFN- β , which functions as a mediator of the antiviral
93 response, was observed (Figure 3).

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97 **Figure 3.** Signaling pathway of the Toll-Like receptor.

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99 Gene expression studies conducted on blood samples from COVID-19 patients indicated that
100 genes associated with inflammatory and hypercoagulability pathways (7) and the imbalance
101 between innate and adaptive immune responses are the main factors responsible for severe
102 disease course (8). Thair et al (9) found a differential transcriptomic pattern in blood that
103 differentiated COVID-19 patients from other viral infections, suggesting that their top
104 differentially expressed genes (DEGs) may be linked to new mechanisms of pathogen
105 evasion from host immune response. Most recently, Aschenbrenner et al. (10) revealed
106 neutrophil activation signatures in severe cases, coupled with an elevated expression of
107 genes related to coagulation and platelet function, and absence of T-cell activation.

108 Considering the ability of coronavirus infections, particularly SARS-CoV-2 infections, to induce
109 inflammation and lung injury and also the similarity of this novel virus to MERS-CoV and
110 SARS-CoV in its ability to infect lung epithelial cells, it is expected that this emerging life-
111 threatening coronavirus utilizes some of the same cellular signaling pathways as MERS-CoV
112 and SARS-CoV. As a therapeutic approach, the use of anti-inflammatory agents during
113 COVID-19 to affect inflammatory signaling pathways might be beneficial for reducing the
114 severity of the disease (11). The NF- κ B, cytokine regulation, ERK, and TNF- α signaling
115 pathways have been shown to be likely causes of inflammation in MERS-CoV and SARS-
116 CoV infections, with neutrophilia and basophilia exacerbating the disease in SARS patients
117 [210]. The increased levels of inflammatory cytokines in serum of COVID-19 patients have
118 shed light on the involvement of such signaling pathways in the pathogenesis infection of
119 SARS-CoV-2. Further studies are needed to clarify the exact roles of cellular signaling
120 pathways once the SARS-CoV-2 initiates infection in its host cell. Also, researchers in
121 molecular medicine should consider the roles of the most strongly up- and downregulated
122 components of cellular signaling pathways during COVID-19 to identify and design better
123 molecular drugs that will decrease the fatality rate of this novel pandemic coronavirus.
124 The findings suggest severe COVID-19 involves hyperactivation of innate immunity and
125 cytotoxic T-cell responses, coupled with impaired adaptive immunity (reduced CD4+ T-cell
126 function). These mechanisms likely contribute to tissue damage and cytokine release
127 syndrome. The study underscores the role of transcriptional dysregulation in driving severe
128 outcomes, offering insights into potential therapeutic targets to modulate immune responses.

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130 **Conclusion:** In the severe state of SARS-CoV-2 viral infection, differential gene expression is
131 represented, genes mainly involved in signaling pathways with CD8 T lymphocyte recruitment,
132 pro-inflammatory, cell cycle alteration, and antiviral response functions.

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134 **Competing Interests:-** The authors declare that they have no financial or nonfinancial
135 competing interests.

136 137 **References**

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