1 "Antimicrobial and Antioxidant Activity of Triphala: An Ayurvedic

2 Formulation"

Abstract

3

18

19

23

- 4 **Triphala**, an ancient Ayurvedic formulation composed of the dried fruits of *Emblica officinalis*
- 5 (Amla), Terminalia chebula (Haritaki), and Terminalia bellerica (Bahera), is widely recognized
- 6 for its broad-spectrum therapeutic applications. This study aimed to scientifically evaluate its
- 7 phytochemical composition, antimicrobial efficacy, and antioxidant potential using different
- 8 solvent extracts. Preliminary phytochemical screening revealed that the **ethanolic extract** was
- 9 the most bioactive, containing alkaloids, tannins, glycosides, saponins, terpenoids, and
- 10 phlobatannins. Antimicrobial testing against Escherichia coli and Staphylococcus aureus using
- the disc diffusion method demonstrated significant activity in ethanolic and aqueous extracts,
- with the ethanolic extract showing the highest inhibition zones. Furthermore, **DPPH radical**
- scavenging assays confirmed the antioxidant capacity of both ethanolic and aqueous extracts in
- a dose-dependent manner, although their efficacy was lower than that of standard ascorbic acid.
- These results substantiate the traditional use of Triphala and highlight its potential as a natural
- source of antimicrobial and antioxidant agents. Further phytochemical characterization and in
- vivo studies are recommended for clinical validation.

Keywords

- 20 Triphala, Antioxidant Activity, Antimicrobial Activity, Phytochemical Screening, Ayurvedic
- 21 Medicine, Ethanolic Extract, DPPH Assay, Herbal Formulation, Natural Therapeutics,
- 22 Escherichia coli, Staphylococcus aureus.

1. INTRODUCTION

- 24 Triphala, a traditional Ayurvedic formulation, has been extensively used for centuries in Indian
- 25 medicine owing to its broad spectrum of therapeutic properties. Composed of the dried fruits of
- 26 three medicinal plants—Emblica officinalis (Amla), Terminalia chebula (Haritaki), and
- 27 Terminalia bellerica (Bahera)—Triphala is revered for its ability to promote digestive health,
- 28 enhance immunity, and delay the aging process. Its multifaceted medicinal applications are
- 29 increasingly being validated by modern scientific investigations, particularly for its antimicrobial
- and antioxidant potential.
- 31 The biological activities of Triphala are largely attributed to the presence of diverse
- 32 phytochemicals. Phytochemicals are naturally occurring, biologically active compounds found in
- plants, excluding essential nutrients such as vitamins and minerals. These compounds act as the
- 34 plant's defense mechanisms against pathogens and environmental stress, and they offer similar
- 35 protective benefits when consumed by humans. Rich in flavonoids, tannins, phenolic acids, and
- other bioactive constituents, phytochemicals exhibit a variety of therapeutic properties, including
- antioxidant, anti-inflammatory, antibacterial, antiviral, and immune-boosting effects.

- 38 Each component of Triphala contributes uniquely to its overall pharmacological efficacy.
- 39 Emblica officinalis (Amla) is a potent source of ascorbic acid (vitamin C), along with tannins,
- 40 flavonoids, and polyphenols such as gallic acid and ellagic acid, all known for their antioxidant
- 41 activity. Terminalia chebula (Haritaki) is rich in tannic acid, chebulinic acid, and betulinic acid,
- 42 which possess significant antimicrobial and free radical scavenging properties. Terminalia
- 43 bellerica (Bahera), on the other hand, contains ellagitannins, gallotannins, and a spectrum of
- 44 polyphenolic compounds that further enhance the formulation's antioxidant and antimicrobial
- 45 activities.
- 46 Given the rising global concern regarding antibiotic resistance and oxidative stress-related
- disorders, there is a pressing need to explore plant-based alternatives with minimal side effects.
- 48 Triphala, with its rich phytochemical profile, presents a promising candidate for such therapeutic
- 49 exploration. This study aims to evaluate the antimicrobial and antioxidant activities of Triphala,
- 50 thereby providing scientific validation for its traditional usage and exploring its potential as a
- 51 natural alternative for managing microbial infections and oxidative stress.

52 **2. MATERIAL & METHOD**

- 53 **2.Materials**
- Triphala powder (100 g) was procured from Vindhya Herbals, Sanjeevani Ayurveda, Bhopal,
- and stored for further use. Extraction solvents included **chloroform**, **petroleum ether**, and
- ethanol, all of analytical grade. Microbial activity was assessed using Nutrient Agar Media
- 57 (NAM) and Nutrient Broth, both with standard compositions and pH adjusted to 7.0.
- 58 **2.1 Instruments**
- 59 Key instruments used included a **Soxhlet extractor** for phytochemical extraction, **rotary**
- vacuum evaporator for solvent recovery, digital pH meter, vertical autoclave, laminar
- 61 **airflow chamber, incubator,** and **antibiotic zone scale** for zone measurement.
- **2.2 Extraction Procedure**
- 63 **Soxhlet extraction** was carried out using 25 g of Triphala powder and 250 mL of solvent at 50–
- 64 60°C until clear extract was obtained. Solvent was then recovered via **rotary evaporation under**
- vacuum, and the residue was mixed with chloroform water and refrigerated at 4°C for further
- 66 analysis.

67 2.3 Phytochemical Screening

- 68 Preliminary phytochemical screening of Triphala extracts was conducted following standard
- 69 protocols (Brain & Turner, 1975; Evans, 1996) to identify key bioactive constituents. Various
- 70 qualitative tests were performed using specific reagents to detect the presence of the following
- 71 phytochemicals:

72

73

- Alkaloids: Detected using Mayer's reagent, indicated by a cream-colored precipitate.
- Carbohydrates & Glycosides: Identified by a red precipitate upon reaction with Fehling's solution.

- **Phenolic Compounds & Tannins:** Confirmed by a bluish-black coloration with ferric chloride.
 - Proteins & Amino Acids: Indicated by a purplish-pink color upon addition of ninhydrin.
 - **Terpenoids:** Detected by the formation of a red-brown interface with chloroform and concentrated H₂SO₄.
 - **Phlobatannins:** Identified through the deposition of red precipitate after boiling with 1% HCl.
 - **Saponins:** Confirmed by persistent frothing upon boiling with distilled water.
 - **Flavonoids:** Detected by a yellow color with NaOH, which disappears after adding dilute HCl

2.4 Antimicrobial Activity (Disc Diffusion Method)

- 86 Escherichia coli and Staphylococcus aureus were used to evaluate antimicrobial activity via the
- disc diffusion method. Inocula were prepared in sterile nutrient broth and incubated at 37°C.
- Nutrient Agar Media (NAM) was prepared and sterilized. Using the pour plate method, 1 mL of
- inoculum was added to Petri dishes, followed by 15 mL of NAM. Sterile filter paper discs loaded
- 90 with 20 μL of Triphala extract were placed on the solidified media and incubated at 37°C for 24
- 91 hours. Zones of inhibition were measured in millimeters to assess antimicrobial efficacy.

92 **2.5** Antioxidant Activity (DPPH Method)

- Antioxidant activity was determined using the DPPH free radical scavenging assay. A 0.004%
- 94 DPPH methanolic solution and ascorbic acid (standard) were prepared in varying concentrations
- 95 (20–100 µg/mL). Triphala extract was similarly diluted. Each sample (2 mL) was mixed with 0.5
- 96 mL DPPH solution and incubated in the dark for 10 minutes. Absorbance was measured at 517
- 97 nm.

98

99

77 78

79

80 81

82

83 84

85

Calculation of Antioxidant Activity

The percentage of DPPH radical scavenging activity was calculated using the following formula:

100 % Inhibition=
$$\underbrace{(A_0 - A_t)}_{A_0} \times 100$$
 101

- A_0 = absorbance of the control
- A_t = absorbance in the presence of sample or standard
- All tests were performed in triplicate, and the results were expressed as mean \pm standard
- 105 deviation.

106 **3. RESULT**

107 3.1 Phytochemical Screening

Preliminary phytochemical screening of Triphala extracts using various solvents revealed the presence of several bioactive compounds (Table 1). The ethanolic extract exhibited the presence of alkaloids, carbohydrates and glycosides, tannins, saponins, terpenoids, and phlobatannins. Chloroform extract showed positive results for alkaloids and terpenoids, while the aqueous extract confirmed the presence of tannins, saponins, and terpenoids. No phytochemicals were detected in the petroleum ether extract.

These findings affirm the presence of secondary metabolites in Triphala, which are known for their therapeutic potential. The ethanolic extract, being the richest in phytochemicals, was subjected to further antimicrobial and antioxidant testing.

	Triphala			
Phytochemical tests	Chloroform	Ethanol	Petroleum ether	Aqueous
Alkaloids (Mayers Reagent)	+	+	-	-
Carbohydrates & glycosides (Fehling Solution)	-	+	-	-
Phenolic compounds & tannins (Ferric Chloride solution)	0.0	+	-	+
Proteins/Aminoacids (Ninhydrin test)		-	-	-
Saponins	-	+	-	+
Terpenoid	+	+	-	+
Phlobatannins	-	+	-	-
Flavanoides	-	-	-	-

3.2 Antimicrobial Activity

The antimicrobial efficacy of Triphala extracts was evaluated using the disc diffusion method against *Escherichia coli* and *Staphylococcus aureus*. The ethanolic extract exhibited the highest

antimicrobial activity, with zones of inhibition measuring 10 mm against *E. coli* and 9 mm against *S. aureus*. The aqueous extract showed moderate activity with inhibition zones of 9 mm and 7 mm, respectively. The chloroform and petroleum ether extracts displayed negligible or no activity.

When compared with the standard antibiotic streptomycin, which showed inhibition zones of 18 mm (*E. coli*) and 13 mm (*S. aureus*), the ethanolic extract demonstrated moderate but significant antimicrobial potential. This activity may be attributed to the presence of phenolic compounds and other bioactive constituents in the extract.

Extract Type	Microorganism	Zone of Inhibition (mm)
Ethanolic	E. coli	10
	S. aureus	9
Aqueous	E. coli	9
	S. aureus	7
Streptomycin	E. coli	18
	S. aureus	13

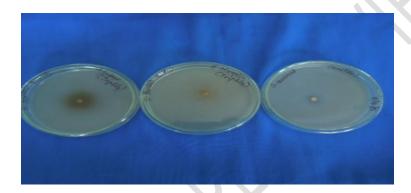


Fig: Zone of inhibition of triphla for S. aureus

3.3 Antioxidant Activity

120

121

122123

124

125

126

127

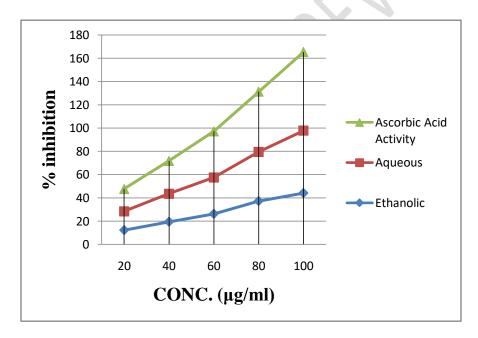
128

- The antioxidant potential of Triphala was evaluated using the DPPH free radical scavenging assay. Both ethanolic and aqueous extracts demonstrated dose-dependent radical scavenging
- activity, although lower than the standard antioxidant, ascorbic acid.
- 134 At the highest tested concentration (100 μg/mL), the aqueous extract showed a scavenging
- activity of 53.6%, followed by the ethanolic extract with 44.2%, compared to 67.6% for ascorbic
- acid. This suggests a significant antioxidant potential, likely due to the presence of phenolic
- compounds, flavonoids, and other phytochemicals.



139 Fig: Antioxidant activity of extract of triphala

Concentration	Ethanolic Extract	Aqueous Extract	Ascorbic Acid
(µg/mL)	(%)	(%)	(%)
20	12.3	16.2	19.2
40	19.5	24.15	28.15
60	26.3	31.3	39.6
80	37.3	42.3	51.7
100	44.2	53.6	67.6



These findings highlight Triphala as a promising source of natural antioxidants with potential applications in food and pharmaceutical industries. The presence of flavonoids, tannins, alkaloids, glycosides, and phenolics may contribute to its free radical scavenging activity. Further isolation and characterization of the active compounds are warranted to understand their mechanisms and enhance the formulation of polyherbal therapeutics.

4. DISCUSSION

146

155

- 147 This study validated the traditional medicinal use of *Triphala* by confirming its phytochemical
- richness, antimicrobial, and antioxidant properties through various solvent extractions. The
- ethanolic extract exhibited the highest concentration of bioactive compounds—including
- alkaloids, tannins, glycosides, and phenolics—correlating with its superior antimicrobial and
- antioxidant effects. Antimicrobial activity was strongest against E. coli and S. aureus with the
- ethanolic extract, while antioxidant potential was observed in both ethanolic and aqueous
- extracts, though less than that of ascorbic acid. These findings highlight *Triphala*'s therapeutic
- potential and suggest its suitability as a natural source for antimicrobial and antioxidant agents.

5. CONCLUSION

- The present study affirms the pharmacological potential of **Triphala**, validating its traditional
- 157 Ayurvedic applications with modern scientific evidence. Among the various solvent extracts
- analyzed, the **ethanolic extract** demonstrated the most potent antimicrobial and antioxidant
- activities, likely due to its rich phytochemical content, especially phenolic compounds, tannins,
- and glycosides. The findings support the role of Triphala as a promising **natural alternative to**
- synthetic antimicrobial and antioxidant agents, with possible applications in the
- pharmaceutical and nutraceutical industries. However, further chemical characterization and
- in vivo investigations are essential to isolate active constituents and determine their mechanisms
- of action and clinical safety profiles.

6. REFERENCES

- 167 [1] Bhowmik D, Tiwari P, Tripathi K, Kumar S. Traditional Indian memory enhancer herbs and
- their medicinal importance. Ann Biol Res. 2010;1(1):41–46.
- 169 [2] Biradar Y, Sharma S, Khendalwal KR. Preparation method of optimization and
- physicochemical evaluation of traditional formulation Triphala. Indian J Tradit Knowl.
- 171 2006;6(2):292–297.
- 172 [3] Chaitali H, Nikhil S, Bhamicee S, Bhamicee SB. Cytotoxicity screening of selected Indian
- medicinal plants using Brine-Shrimp lethality bioassay. Adv Nat Appl Sci. 2010;4(3):389–395.
- 174 [4] Dhanalakshmi S, Kumar R, Sundaramahalingam P, Naraganaperumal Jeya. Antioxidant
- property of Triphala on cold stress induced oxidative stress in experimental mice. J Health Sci.
- 176 2006;52(6):843–842.
- 177 [5] Girish C, Koner B, Jayanthi S, Rao KR, Rajesh B, Pradhan SC. Hepatoprotective activity of
- six polyherbal formulations in paracetamol induced liver toxicity in mice. Indian J Med Res.
- 179 2009;129:569–578.
- 180 [6] Gupta D, Mradu R. Therapeutic uses of the polyherbal drug Triphala in geriatric diseases. Int
- 181 J Pharm Bio Sci. 2010;1(2):145–158.
- 182 [7] Gupta M. Therapeutic uses of the polyherbal drug Triphala in geriatric diseases. Int J Pharm
- 183 Bio Sci. 2010;1(2):267–285.
- 184 [8] Hussin A. Adverse effects of herbs and drug-herbal interactions. Malays J Pharm.
- 185 2001;1(2):39–44.
- 186 [9] Kaur S, Jaggi RK. Antinociceptive activity of chronic administration of different extracts of

- 187 Terminalia bellirica Roxb and Terminalia chebula Retz. fruits. Indian J Exp Biol. 2010;48:925–
- 188 930.
- 189 [10] Khan KH. Terminalia chebula reduces the oxidative stress induced by Salmonella
- typhimurium in mice and may reduce the risk of getting typhoid. Adv Biol Res. 2009;3:1–2.
- 191 [11] Khan K, Hayat. The effect of regular intake of Terminalia chebula on oxidative stress in
- mice originated from Salmonella typhimurium. EurAsian J Biosci. 2009;3:113–121.
- 193 [12] Lagae L, Buyse G, Ceulemans B, Claeys P, Dedeurwaerdere S, De Meirleir L, Hauman R,
- Janssen A. Anti-epileptogenesis research: the clinical relevance. Acta Neurol Belg.
- 195 2003;103:78–82.
- 196 [13] Masola SN, Mosha RD, Wambura PN. Assessment of antimicrobial activity of crude
- 197 extracts of stem and root barks from Adansonia digitata (Bombacaceae) (African baobab). Afr J
- 198 Biotechnol. 2009;8:5076–5083.
- 199 [14] Mahajan A, Pai D, Nandini R. Development and validation of HPLC method for
- quantification of phytoconstituents in Haritaki Churna. Int J ChemTech Res. 2011;3:329–336.
- 201 [15] Mahesh R, Ramalingam B, Bhuvana S, Begum HV, Vava M. Effect of Terminalia chebula
- aqueous extract on oxidative stress and antioxidant status in the liver and kidney of young and
- 203 aged mice. Cell Biochem Funct. 2009;27:358–363.
- [16] Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TPA. Indian herbs and herbal
- drugs used for the treatment of diabetes. J Clin Biochem Nutr. 2007;40:163–173.
- 206 [17] Preethi R, Vimal V, Devanathana V, Loganathan M. Antimicrobial and antioxidant
- efficiency of some medicinal plants against food borne pathogens. Adv Biol Res. 2010;4(2):122–
- 208 125.

220221

222

223

224

- 209 [18] Rahman S, Akbar M, Howlader A, Jabber A. Antimicrobial and cytotoxic activity of the
- 210 alkaloids of Amlaki (Emblica officinalis). Pak J Biol Sci. 2009;12(16):1152–1155.
- 211 [19] Singh M, Sharma CS. Wound healing activity of Terminalia chebula in experimentally
- induced diabetic mice. Int J PharmTech Res. 2009;1(4):1267–1270.
- 213 [20] Sharma US, Sharma U, Kumar U, Singh A, Singh N, Puspak J. Screening of Terminalia
- bellirica fruits extracts for its analgesic and antipyretic activities. Int J PharmTech Res.
- 215 2010;3(3):121–124.
- 216 [21] Tambekar DH, Dahikar SB, Lahare MD. Antibacterial potentials of some herbal
- preparations available in India. Res J Med Med Sci. 2009;4(2):224–227.
- 218 [22] Tambekar DH, Dahikar SB. Exploring antibacterial potential of some Ayurvedic
- preparations to control bacterial enteric infections. J Chem Pharm Res. 2010;2(5):494–501.