

# Chemical Functionalization and cross-linking of biochar from cow dung for use as supper

## ABSTRACT

Water plays a vital role in maintaining the structure and posture of plants, keeps the cells turgid, aids in the transportation of nutrients from the soil, among other functions. It plays an important role in ensuring there's food security in communities, particularly in arid and semi-arid areas, when well managed. This is ensured through irrigation and mulching. Irrigation is a tedious exercise, while mulching is labor-intensive, and natural mulching materials do not retain water for long, hence the need for improvement. In this study, cow dung biochar was functionalized with ethan-1,2-diol, glycerol, and phosphoric acid and then cross-linked with phosphoric acid, citric acid, and urea to develop a super absorbent mulch substrate for agricultural purposes. To begin, cow dung was obtained from a cattle farm in Thika in Kiambu County. It was converted to biochar by burning under a limited supply of air, which was then functionalized and cross-linked to make a mulch for agricultural practices, a technique that has been put to test for the very first time. The modified biochar was tested for conservation of moisture and retention of nutrients based on watering frequency and the improved crop yields. A successful functionalization process was confirmed using Fourier Transform Infra-Red Spectroscopy (FTIR), which showed the additional functional groups in the chemical structure of the modified biochar. The degree of cross-linking was determined by a swelling experiment. The degree of water retention was determined by comparing the mulched and non-mulched maize and Calliandra calothyrsus trees in the field. Field experiments were conducted in Makuyu ward, Maragua constituency, Murang'a County, a semi-arid area where maize was grown using a randomized block design. Growth and yield parameters were monitored periodically. The FTIR peaks were observed for -OH at  $3350.73\text{ cm}^{-1}$ , P-OH at  $788.87\text{ cm}^{-1}$ , P=O at  $1279.5\text{ cm}^{-1}$ , C-H at  $2919.68\text{ cm}^{-1}$ , C-N at  $1045\text{ cm}^{-1}$ , N-H at  $1622.80\text{ cm}^{-1}$  and C=O at  $1602.5\text{ cm}^{-1}$ , confirming a successful modification process. Based on the results of this study, it is recommended that the modified mulch be used in arid and semi-arid areas.

## 1.1 Background Information

Mulching is the act of covering the top soil with a layer of materials such as grass clippings, wood chips, and straw, chopped leaves, twigs, saw dust, and even papers. Their utilization as mulching materials has been used in agriculture (Bach *et al.*, 2016). In nurseries, grass clippings have been the most widely used mulch because they are readily available, prevent excessive evaporation and also moderate soil temperatures (Wells *et al.*, 2016). These mulching materials have been used to improve soil moisture, beautify the land, control weeds and improve the soil fertility but do not retain water for long hence need for a more reliable mulch.

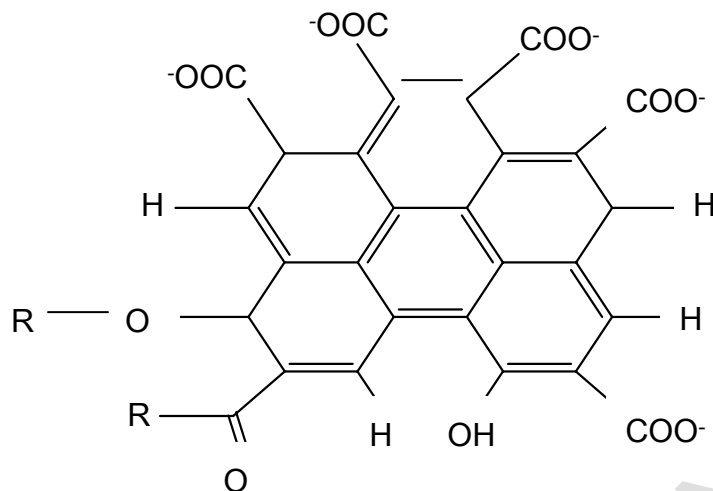
Biochar is the light weight black residue which remains after biomass has been burnt in the absence of oxygen. It has been found to be more porous (Chrisman, 2015) and can hold more water for long than natural mulches. It has been made in the past from biomass materials like rice husk, olive shell, crop residues, pig manure, cattle manure and wood (Ma *et al.*, 2019). Biochar in the soil has many benefits some of which include increasing water infiltration, promote better root development, reduce fertilizer run-off especially nitrogen and phosphorous, binds sandy soil together and improves water holding capacity (Lonappan *et al.*, 2019). It adsorbs and immobilizes heavy metals and reduces their uptake by plants and subsequent ingestion by humans and animals (Liet *et al.*, 2018). It has carbon bonds that readily accept functional groups to form hydrogel (Huff *et al.*, 2016). Most of the chemically improved biochar have been used in adsorption of heavy metals but there is limited information on use of chemically improved biochar in agricultural practices. In Kenya, intensive livestock farming system is a major source of environmental pollution especially in urban areas where people live in small plots of land. Even though cow dung has been used for biogas production (Mwangi *et al.*, 2019), the biogas slurry poses a problem in its disposal.

There are many adverse impacts of climate change especially in agricultural sector which is heavily affected by drastic weather and climate change resulting to food insecurity. There is therefore need to search for an amicable solution to mitigate the impact of climate change on agriculture which is the backbone of our economy. This can be done through development of super absorbents such as functionalized and cross-linked cow dung biochar with ability to retain water, nutrients and keep the crops and trees growing even when the weather conditions are not

favorable.

Cow dung is the undigested residue of plant matter which passes through the digestive system of the bovine animal species. It is rich in nutrients like nitrogen, phosphorus, potassium and high levels of ammonia (Randhawa *et al.*, 2011). It also consists of lignin, cellulose and hemicelluloses (Singh *et al.*, 2014). It has several benefits some of which include burning as fuel, mosquito repellent, plaster for buildings, bio-pesticides and as a cleansing agent (Mwangi *et al.*, 2019). Cow dung manure increases humus and this results in better water seepage and water binding capacity together with an improved cation storage capacity (Bio-fertilizers, 2014). It is also used for making biogas which is used for cooking thus lowering dependency on charcoal, firewood and fossil energy (Guo *et al.*, 2013). Cow dung poses a problem in its disposal since it is produced in large volumes especially in town set-up where people live in small plots of land (Duplaet *et al.*, 2024).

Biochar is a form of charcoal produced through the thermal chemical conversion of biomass under low oxygen conditions known as pyrolysis (Guo *et al.*, 2013). When air is excluded, oxygen for burning is removed from the biomass leading to its reduction to carbon single bonds of charcoal (Chen *et al.*, 2017). In ordinary burning, biomass would be oxidized into ash, carbon (IV) oxide, vapors and other gases (Ekebafe *et al.*, 2013). It has many benefits in the soil like increasing water penetration and water retention capacity, it improves the soil configuration thus improving root development which leads to better yield (Chrisman, 2015). Biochar amended soils improve nitrogen use efficiency, adsorption of ammonium, phosphate and calcium ions (Richardson *et al.*, 2011). It enhances fertility, nutrient retention, soil biological activity and diversity and reduces fertilizer run-off especially nitrogenous and phosphate (Zahra *et al.*, 2021). Biochar from animal waste is generally high in nutrient content particularly nitrogen, phosphorous, calcium and magnesium compared to plant residue. This has been stated by Utomo and Nugroho (2011) whose research looked at the production of a simple biochar from coconut and animal waste and its application in Lombok and the other eastern parts of Indonesia. They also found out that biochar made from animal waste was less alkaline (pH 8.9) compared to that from coconut shell which had a pH of 9.9 (Utomo *et al.*, 2011). The chemical structure of biochar components produced by the fast pyrolysis process at 500-600°C is shown in Figure 2.1



**Figure 2.1. Structure of biochar (Chen *et al.*, 2017)**

## 2.1 Chemical functionalization of biochar materials

Use of organic acids and alkalis solutions have been reported in the pretreatment of biochar materials to improve the internal surface area (Usman *et al.*, 2015). Chemical modification alters the outside chemistry of biochar, thereby leading to improved water holding capacity in comparison with un-functionalized biochar. The activated and functionalized biochar have several applications in diverse field which include soil recreation, carbon dioxide sequestration, and removal of heavy metals and degradation of organic pollutants (Yakout *et al.*, 2015). The commonly used activating chemicals include  $H_2SO_4$ ,  $H_3PO_2$ ,  $KOH$ ,  $ZnCl_2$  and  $K_2CO_2$ . Chemical functionalization can be done by introducing specific chemical functional groups like  $-SO_2H$  and  $-COOH$  into the biochar matrix (Ma *et al.*, 2019). In the present study, functionalization of cow dung biochar by incorporating functional groups inside the polymer chain and then cross-linking it enhanced water holding capacity, nutrient adsorption and increased swelling after water absorption.

Hydrogels are polymers which are loosely cross-linked and exhibit the capacity to swell and hold a large amount of water within its structure, but does not dissolve in it (Alkhasha *et al.*, 2018). The capability of hydrogel to take up water arises from the hydrophilic functional groups attached

to the polymeric backbone, while their resistance to dissolve arises from cross-links between its network chains (Wu *et al.*, 2021). They can be classified depending on their physical properties, method of preparation, nature of swelling, ionic charges, rate of their biodegradation and nature of cross-linking (Larrañeta, 2018).

Hydrogel are extensively implemented as input materials in various biomedical applications due to their spongy, stretchy, hydrophilic and quasi-solid nature (Ahmed, 2015). They are also used in bio-separation, tissue engineering, proteomic and chromatography where good water holding capacity is of prime importance (Ullah *et al.*, 2015). There are three polymerization techniques which can be applied to form gel, they include homo-polymeric which is derived from the same species of monomer (Ahmed *et al.*, 2019), the second one is co-polymeric hydrogel which comprise of two or more different monomer species. Multi-polymer interpenetrating polymeric hydrogel is the third one which is as a result of two free cross-linking synthetic or natural polymer components (Chrisman, 2015).

Superabsorbent hydrogel act as humus which join loose soil hence form loam soil which help better root penetration (Ahmed *et al.*, 2019), it increases the plant accessible water in the soil, encourage faster growth of plant even under best water conditions and lengthen the survival of plants under water shortage (Ekebafe *et al.*, 2013).

### **3.0 MATERIALS AND METHODS**

#### **3.1 Sampling of cow dung and pre-treatment**

Cow dung was collected by random sampling from Thika in Kiambu county from Freshian breed of cattle, sundried for 6-7 days until there was no change in weight. Stones and other foreign materials were removed and then the cow dung was burnt at a temperature of 300 °C in the absence of air to make biochar

#### **3.2 Preparation procedure of Biochar**

200 g of cow dung was placed in a tight rid tin where it was burnt for 30 minutes at a temperature of 300 °C. It was allowed to stay in that state for 10 minutes after which it was cooled at room temperature for one hour (Pattanik *et al.*, 2019)

#### **3.3 Experimentation Procedure for functionalization of biochar with phosphoric acid**

The reaction of biochar with phosphoric acid was performed by adding 100 cm<sup>3</sup> H<sub>3</sub>PO<sub>4</sub> (85%) to 50 g of biochar sample at 40 °C as described by Gyarmati *et al* (2015). The system temperature was raised to 100 °C and maintained for 40 minutes while stirring at 10 minutes intervals. The reaction mixture was then cooled in an ice-bath at a temperature of 15 °C. The mixture was decanted from the resulting suspension and washed four times with water. 10 g of urea was added to phosphorated biochar and the temperature raised to 150 °C for cross-linking to take place in the biochar chains. The cross-linked biochar hydrogel was then analyzed using Fourier transform Infrared spectroscopy (FTIR).

##### **3.3.2 Functionalization of biochar with Ethan-1,2-diol**

The functionalization procedure of biochar with ethan-1,2-diol was performed as per (Yakout *etal.*, (2015) method with slight improvement where 10 g of biochar was mixed with 100 ml of ethan-1, 2-diol in the presence of sulphuric acid and a temperature of 120° to increase surface area and porosity of biochar. The resulting ester was then cross-linked with phosphoric acid where heating

under reduced pressure in the presence of an inorganic base at a temperature of 150°C was done. The crude phosphate ester was purified by first washing with a chelating agent followed by water after which they were dried.

### **3.3.3 Functionalization procedure of biochar with glycerol**

In the presence of sulphuric acid, 10 g of biochar was reacted with glycerol at 120 °C for 2 hours to form an ester. The cross-linking method was adopted from Gyarmati, (2015). The ester was heated at 165 °C for 10 minutes, 100 g of the ester was placed in a beaker with 0.1 M NaOH solution with pH of 12.00 necessary for cross-linking for stability to be attained due to the alkali environment. The liquid was stirred with a stirrer at a speed of 250 rpm for one and a half hours and the liquid discarded. The residue was washed four times using 400 ml of water for 1 hour and then dried on a porcelain tray in the open for 4 hours.

The treated sample was added in to 600 ml of 0.6 M citric acid solution which was then stirred (400 rpm) for 40 minutes. The acid treated hydrogel was washed with water to remove any excess of citric acid until the effluent turned non-turbid with lead (II) nitrate in buffer solution at pH 4.6. Drying was then done in an inert environment.

### **3.3.4 Procedure for functionalization of biochar with phosphoric acid and cross-link it with urea and citric acid**

The hydrogel in procedure 3.2.3 was cross-linked again with citric acid where 200 g of cross-linked biochar with urea was placed in 4 l of 0.1 M NaOH with a pH of 12.00 necessary for cross-linking so as to attain an alkali environment. The mixture was stirred at a speed (250 rpm) for 1 hour and the liquid discarded. The residue was washed 4 times with 400 ml of water for one hour and then dried on a porcelain tray in the open for 4 hours.

The treated biochar sample was added to 120 ml of 0.6 M citric acid solution which was then stirred (400 rpm) for one hour. The acid treated hydrogel was washed with water to remove any excess citric acid until the effluence turned non-turbid with lead (II) nitrate in buffer solution at pH 4.6, it was then dried in an inert environment.

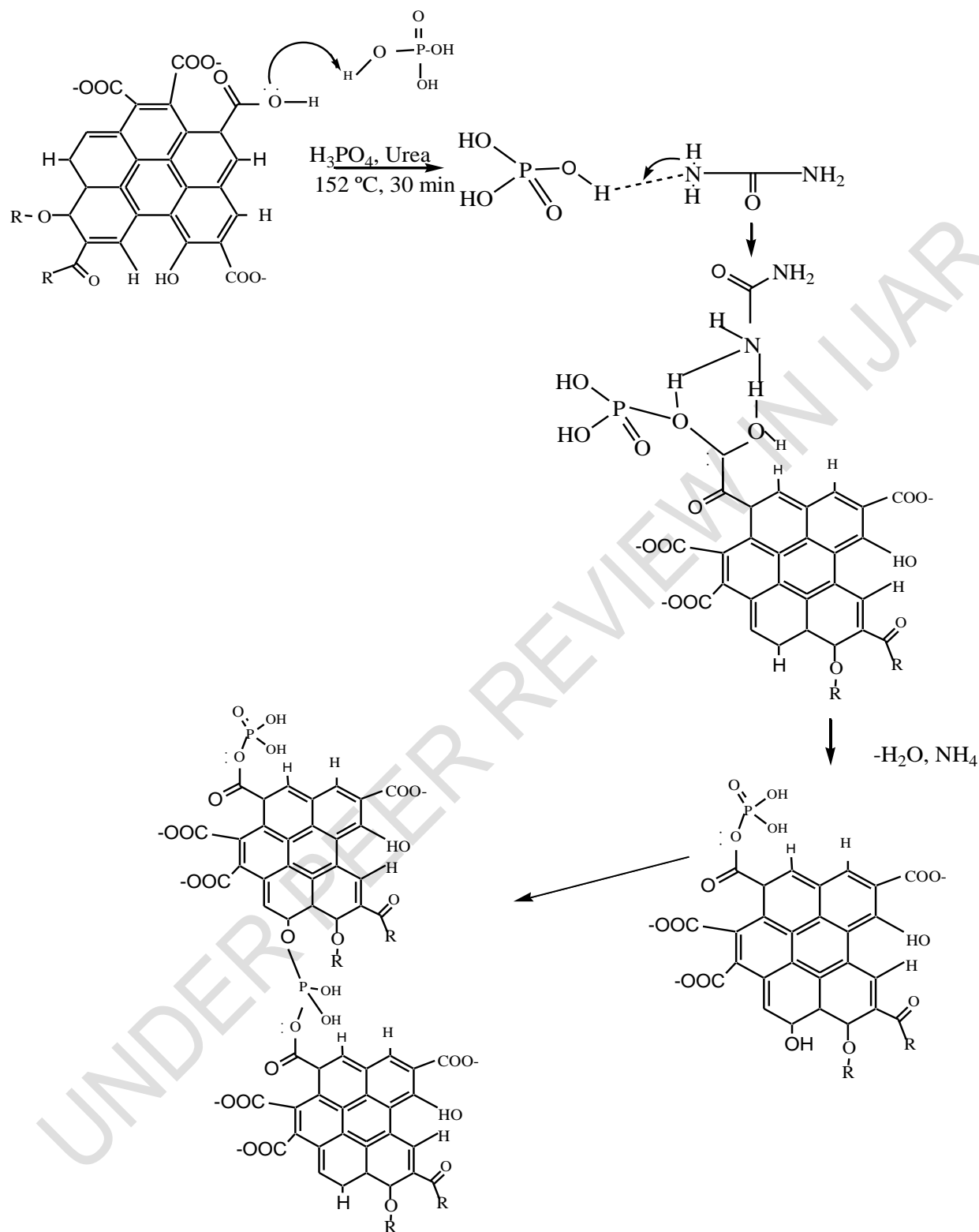
## RESULTS AND DISCUSSION

### 4.1 Functionalization of biochar and cross-linking

#### 4.1.1 Functionalization of biochar with phosphoric acid followed by cross-linking with urea

Phosphoric acid was linked to biochar chain under the form of phosphoric acid group by ester linkage through reactions from biochar hydroxyl to form phosphorylated biochar. The phosphorylated biochar was then cross-linked by the use of urea; the temperature was raised to 150 °C because the melting point of urea is 144 °C (Martinez-Villadiego *et al.*, 2024).

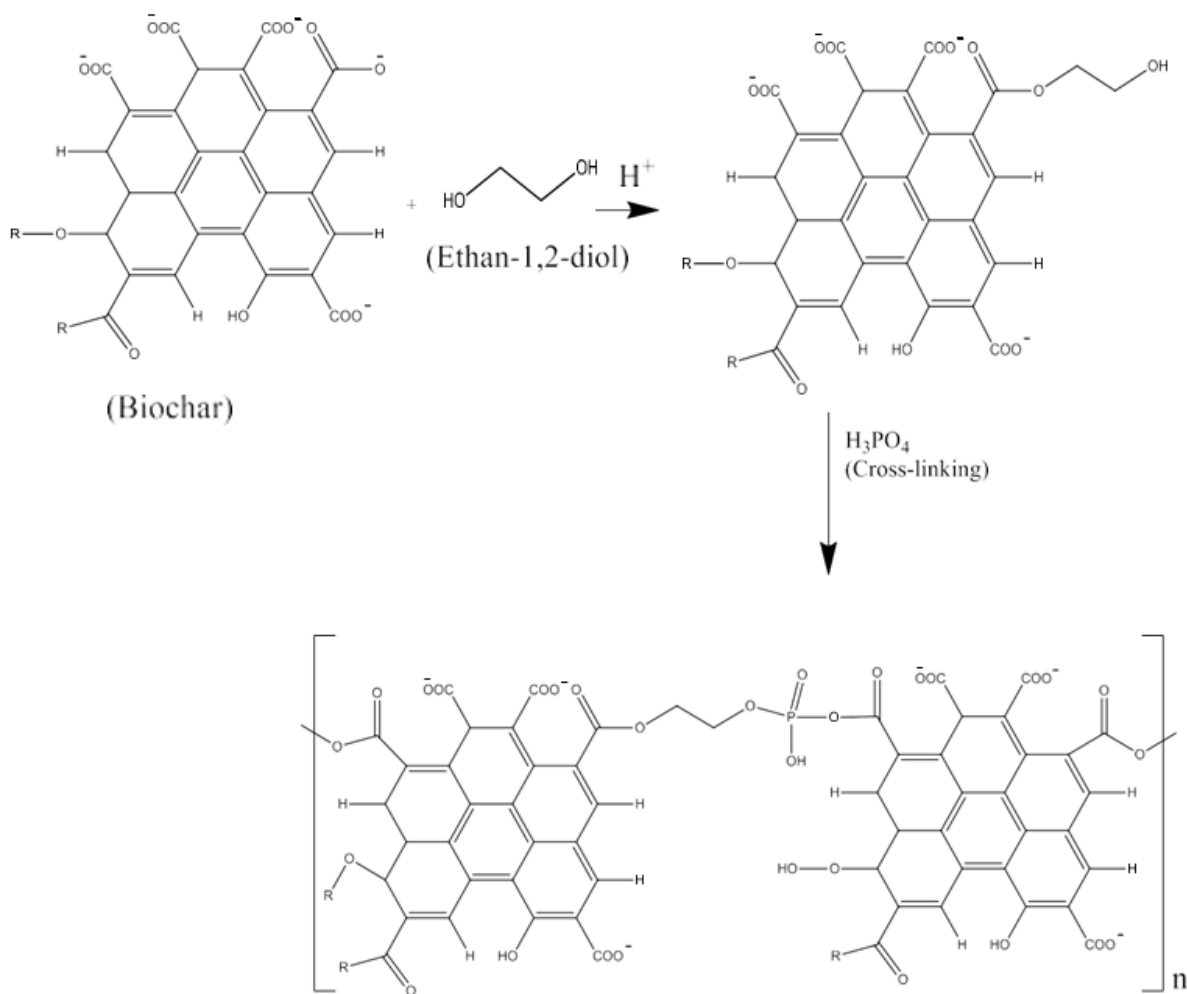




**Figure 1. Reaction Scheme for the functionalization and cross-linking of biochar with phosphoric acid followed by urea**

#### 4.1.2 Functionalization of biochar with Ethan-1,2-diol followed by cross-linking with phosphoric acid

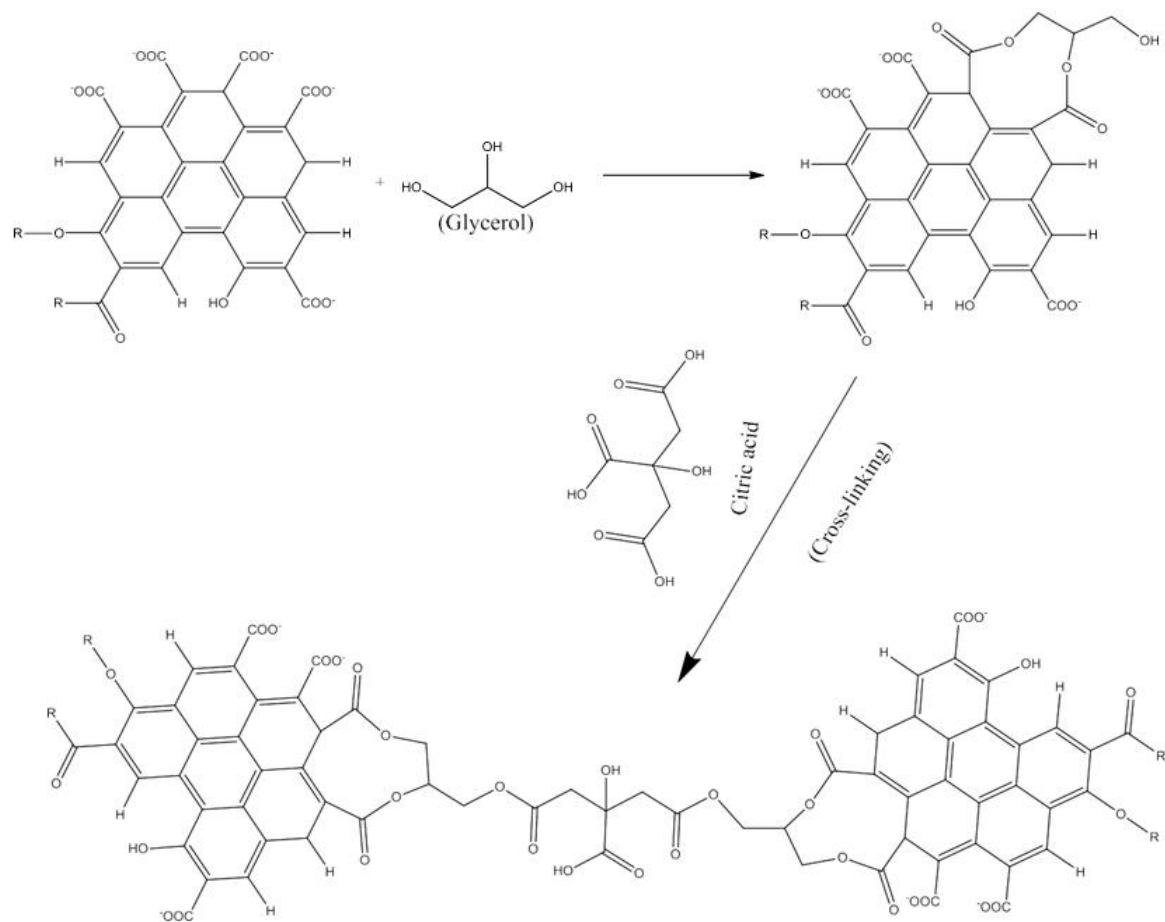
It involved an esterification reaction of ethan-1,2-diol with biochar in the presence of sulphuric acid which was then cross-linked with phosphoric acid to give a phosphoric Ester hydrogel



**Figure 2. Reaction Scheme for the functionalization of biochar with Ethan-1,2-diol and cross-linked with phosphoric acid**

#### 4.1.3 Functionalization of biochar with glycerol and cross-linking with citric acid

It involved reacting 10g of biochar with glycerol at 120 °C for two hours in the presence of sulphuric acid to form an ester. The cross-linking method was adopted from Gyamati, 2015 where the ester was heated at 90° C for 20 minutes after which 100g of the ester was placed in a beaker with 0.1M NaOH solution at a pH of 12.00 necessary for cross-linking to attain stability due to the alkali environment. The liquid was discarded and a carboxylation reaction followed as shown by the reaction scheme Figure 3.



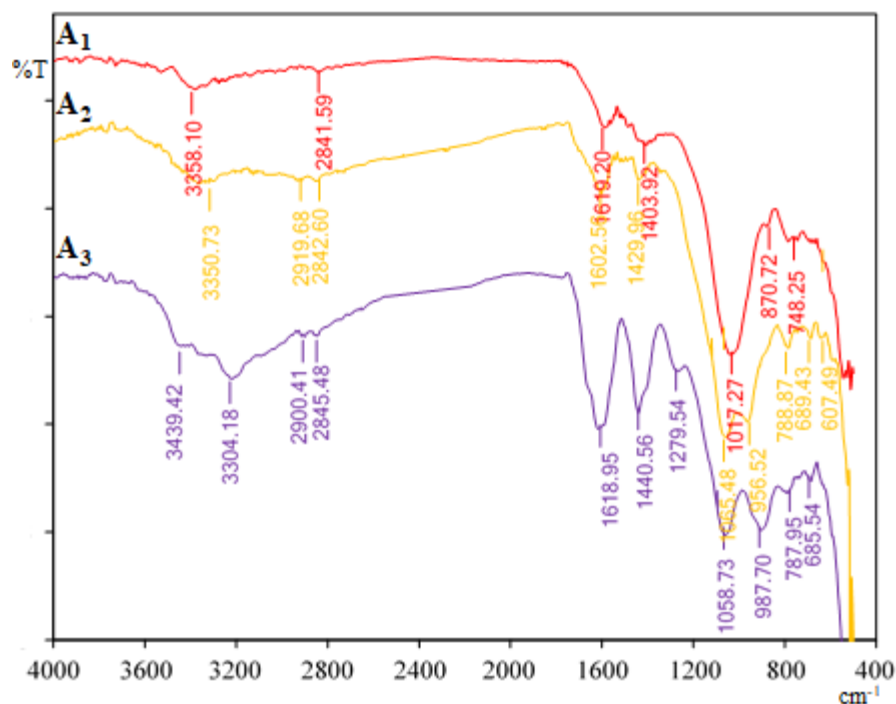
**Figure 3. Reaction scheme of functionalization of biochar with glycerol then cross-linked with citric acid**

#### **4.1.4 Functionalization of biochar with ethan-1,2-diol**

Biochar was reacted with ethan-1,2-diol by ester linkage and then cross-linked twice by the use of phosphoric acid to get a phosphoric Esther followed by a second cross-link with urea to give urea phosphate. The mulch did not hold a lot of water as one would expect as compared to the mulches with single cross-link since its swelling was 7.76 times its weight as compared to phosphate functionalized with urea cross-link which had a swelling capacity of 12.09 times its weight. This result was in line with a study which was done by An *et al.*, (2021) on the relationship between cross-linking and swelling capacity. He found out that if too much cross-linking agent is used, then the high cross-linking degree causes a low swelling degree of hydrogel hence little water is held. However, if the amount of cross-linking agent is too small, then the physical interaction between polymer bonds breaks easily causing the hydrogel to become water soluble.

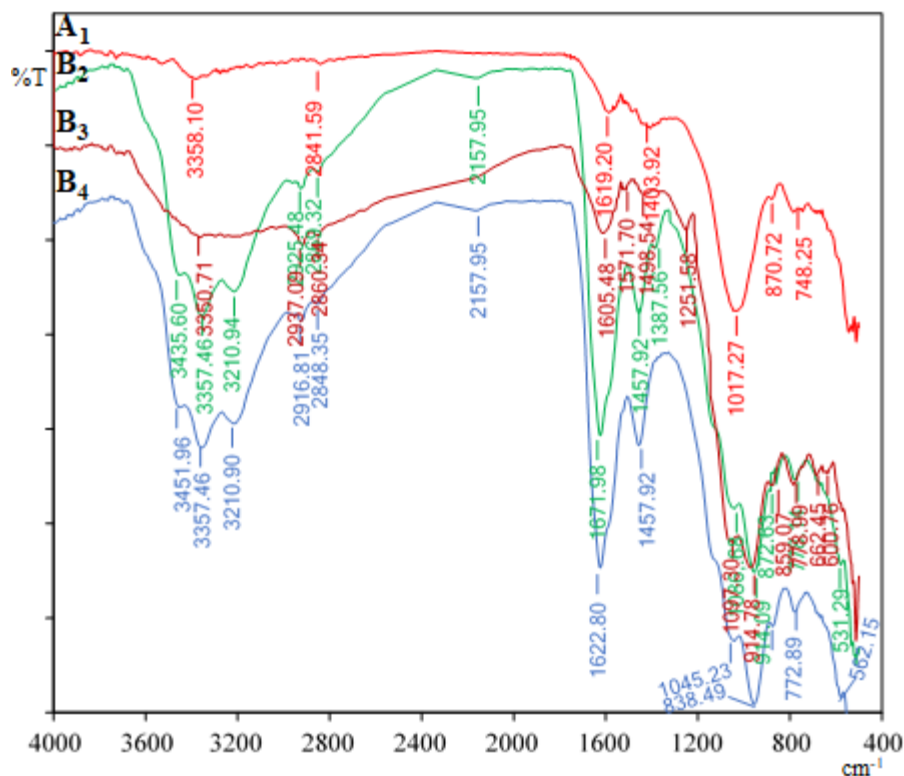
#### **4.2 FTIR Spectra characterization of modified and unmodified biochar**

Functionalized and non-functionalized mulch were characterized using FTIR. The spectra of biochar, phosphate functionalized and urea cross-linked biochar are given in Figure 4.1



**Figure 4.1: The FTIR spectra of non-functionalized biochar (A<sub>1</sub>), phosphate functionalized Biochar (A<sub>2</sub>) and urea cross-linked mulch (A<sub>3</sub>)**

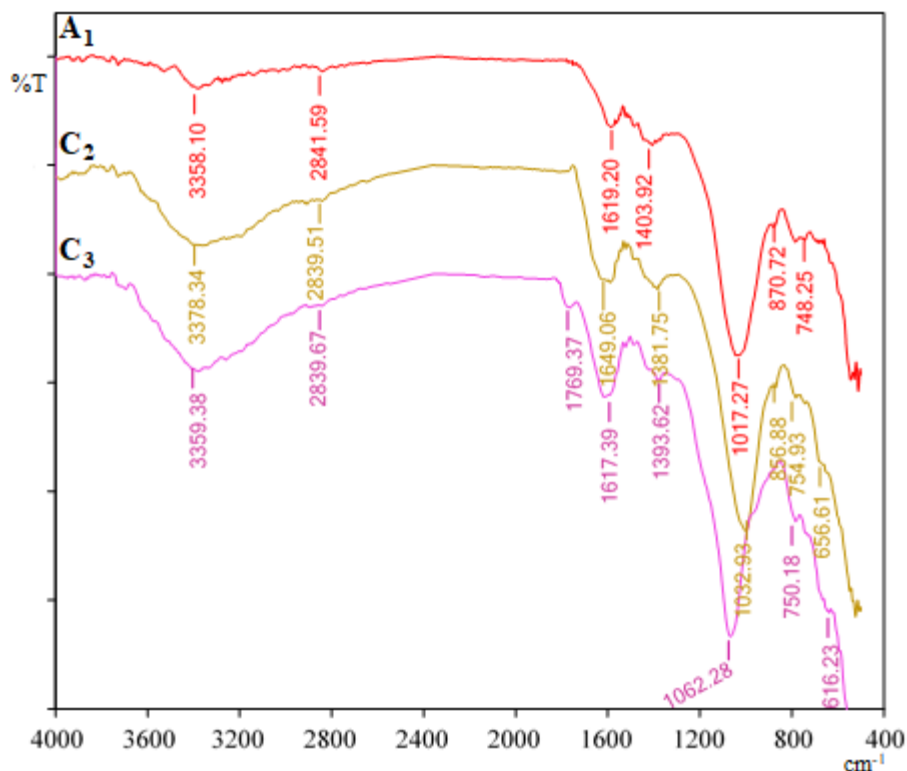
All the spectra bands about 3350.73-3439.42  $\text{cm}^{-1}$  are attributed to either -NH or -OH groups (Petit *et al.*, 2018). The bands at 2919.68 $\text{cm}^{-1}$  and 2900.41 $\text{cm}^{-1}$  were attributed to C-H stretches while 1618.95  $\text{cm}^{-1}$ , 1610.20  $\text{cm}^{-1}$  and 1602.5 $\text{cm}^{-1}$  were attributed to stretching C=O, N-H or -OH (Nandiyanto *et al.*, 2019). The peaks at 788.87 and 956.52  $\text{cm}^{-1}$  correspond to P-OH stretching vibration mode of phosphoric group (Zhang *et al.*, 2021) which was used to functionalize biochar. A band at 1279.5  $\text{cm}^{-1}$  corresponds to the P=O stretching mode (Kubicki *et al.*, 2012) and a peak at 1058.73  $\text{cm}^{-1}$  was attributed to C-N stretching which indicate a successful cross-linking with urea.



**Figure 4.2** FTIR spectra of non-functionalized biochar (A<sub>1</sub>), Ethan-1,2-diol functionalized (B<sub>2</sub>), Phosphate cross-linked (B<sub>3</sub>) and second cross-linking with Urea (B<sub>4</sub>)

There are broad and strong bands at about  $3451.96\text{ cm}^{-1}$ ,  $3435.60\text{ cm}^{-1}$  and  $3358.10\text{ cm}^{-1}$  which were attributed to -OH or -NH groups (Enders *et al.*, 2021) from biochar. A peak at  $1387.56\text{ cm}^{-1}$  corresponds to C-H stretching from Ethan-1,2-diol while a band at approximately  $1251.58\text{ cm}^{-1}$  correspond to the P=O stretching mode (Kubicki *et al.*, 2012) and a peak at  $859.07\text{ cm}^{-1}$  is attributable to P-OH stretching vibration mode (Frost *et al.*, 2013).

A band at approximately  $3357.46\text{ cm}^{-1}$  corresponding to N-H stretching vibration mode (Branca *et al.*, 2016), a peak at  $1622.80\text{ cm}^{-1}$  can be attributed to N-H bending vibration of amines according to Asyana, (2016) and a band at approximately  $1045.23\text{ cm}^{-1}$  correspond to C-N stretching vibration mode (Krichen *et al.*, 2019)



**Figure 4.3: FTIR spectra of non-functionalized biochar (A<sub>1</sub>), glycerol functionalized (C<sub>2</sub>) and citric acid cross-linked (C<sub>3</sub>).**

The observed peaks at 3359.38, 3378.34 and 3558.10  $\text{cm}^{-1}$  in all the three spectra's is due to -OH Stretching vibrations (Fontues *et al.*, 2021), the peak around 1617.39  $\text{cm}^{-1}$  stretching was attributed to a carbonyl (Cai *et al.*, 2019) which indicates that the C=O from citric acid cross-link was successively anchored.

## Conclusions

The functionalization with Ethan-1,2-diol, glycerol, phosphate and cross-linking with urea, phosphate and citric acid were confirmed with the FTIR spectrum which showed peaks for -OH at 3350.73  $\text{cm}^{-1}$ , P-OH at 788.87  $\text{cm}^{-1}$ , P=O at 1279.5  $\text{cm}^{-1}$ , C-N at 1045.23  $\text{cm}^{-1}$ , N-H at 1622.80  $\text{cm}^{-1}$ , C-H at 2919.68  $\text{cm}^{-1}$  and C=O at 1602.5  $\text{cm}^{-1}$ . This confirmed that the diol, phosphate and amino

used for functionalization and cross-linking had been successively anchored.

## REFERENCES

Ahmed, E.M. (2015). Hydrogel: Preparation, characterization, and applications: A review.

Ahmed, S., Ahmad, M., Swami, B. L., Ikram, S., Ahmed, S., Ahmad, M., Swami, B. L. (2019). Green synthesis of silver nanoparticles using *Azadirachta indica* aqueous leaf extract  
*Journal of Radiation Research and Applied Sciences*, 9(1), 1–7. <https://doi.org/10.1016/j.jrras.2019.06.006>

Alkhasha, A., Al-omran, A., & Aly, A. (2018). *Effects of biochar and synthetic polymer on the Hydro-physical properties of sandy soils*. (C), 1–18. <https://doi.org/10.3390/su10124642>

An, Y., Xiong, S., Liu, R., You, J., Yin, T., & Hu, Y. (2021). The effect of cross-linking degree on physicochemical properties of surimi gel as affected by MTGase. *Journal of the Science of Food and Agriculture*, 101(15), 6228–6238.

Asyana, V., Haryanto, F., Fitri, L. A., Ridwan, T., Anwary, F., & Soekersi, H. (2016, March). Analysis of urinary stone based on a spectrum absorption FTIR-ATR. In *Journal of Physics: Conference Series* (Vol. 694, No. 1, p. 012051). IOP Publishing.

Bach, M.A., & Pelacho, A.M. (2016). *Biodegradable Mulching in an Organic Tomato Production System*. (February). <https://doi.org/10.17660/ActaHortic.2008.767.28>

Bio-fertilizers, K. (2014). *Cow Dung for Ecofriendly and Sustainable Productive Farming* Abhishek Raj Manoj Kumar Jhariya Pratap Toppo *Environmental Science*. (2277), 10–11. <https://doi.org/10.1098/rstb.2007.2189>.

Branca, C., D'Angelo, G., Crupi, C., Khouzami, K., Rifici, S., Ruello, G., & Wanderlingh, U. (2016). Role of the OH and NH vibrational groups in polysaccharide-nanocomposite interactions: A FTIR-ATR study on chitosan and chitosan/clay films. *Polymer*, 99, 614–622.

Cai, Z., Ji, B., Yan, K., & Zhu, Q. (2019). Investigation on reaction sequence and group site of citric acid with cellulose characterized by FTIR in combination with two-dimensional correlation spectroscopy. *Polymers*, 11(12), 2071.

Chen, Y., Zhang, X., Chen, W., Yang, H., & Chen, H. (2017). The structure evolution of biochar from biomass pyrolysis and its correlation with gas pollutant adsorption performance. *Bioresource technology*, 246, 101–109.



- Chrisman, R. (2015). *Economics of Biochar Production, Utilisation and GHG Offsets* Bruce A. McCarl Department of Agricultural Economics See coauthors below Presented at Newcastle upon Tyne, UK. (June).
- Dupla, X., Bonvin, E., Deluz, C., Lugassy, L., Verrecchia, E., Baveye, P. C., ... & Boivin, P. (2024). Are soil carbon credits empty promises? Shortcomings of current soil carbon quantification methodologies and improvement avenues. *Soil Use and Management*, 40(3), e13092.
- Ekebafé, M. O., Ekebafé, L. O., & Maliki, M. (2013). *Utilisation of biochar and superabsorbent polymers for soil amendment*. 96, 85–94.  
<https://doi.org/10.3184/003685013X13587941096281>
- Enders, A. A., North, N. M., Fensore, C. M., Velez-Alvarez, J., & Allen, H. C. (2021). Functional group identification for FTIR spectra using image-based machine learning models. *Analytical Chemistry*, 93(28), 9711–9718.
- Fontes, E. H., Ramos, C. E. D., Ottoni, C. A., de Souza, R. F. B., Antolini, E., & Neto, A. O. (2021). Glycerol dehydrogenation steps on Au/C surface in alkaline medium: An in-situ ATR-FTIR approach. *Renewable Energy*, 167, 954–959.
- Frost, R., Siegelman, N., Narkiss, A., & Afek, L. (2013). What predicts successful literacy acquisition in a second language?. *Psychological science*, 24(7), 1243–1252.
- Guo, Y., Tang, H., Li, G., & Xie, D. (2013). *Effects of Cow Dung Biochar Amendment on Adsorption and Leaching of Nutrient from an Acid Yellow Soil Irrigated with Biogas Slurry* Effects of Cow Dung Biochar Amendment on Adsorption and Leaching of Nutrient from an Acid Yellow Soil Irrigated with Biogas Slurry. (November).  
<https://doi.org/10.1007/s11270-013-1820-x>
- Gyarmati, B., Mészár, E. Z., Kiss, L., Deli, M. A., László, K., & Szilágyi, A. (2015). *Acta Biomaterialia Super macroporous chemically cross-linked poly(aspartic acid) hydrogels*. <https://doi.org/10.1016/j.actbio.2015.04.033>
- Huff, M. D., & Lee, J. W. (2016). Biochar-surface oxygenation with hydrogen peroxide. *Journal of Environmental Management*, 165, 17–21.  
<https://doi.org/10.1016/j.jenvman.2015.08.046>
- Krichen, F., Walha, S., Lhoste, J., Bulou, A., Hemon-Ribaud, A., Goutenoire, F., & Kabadou, A. (2019). Supramolecular and heterometallic architectures based on [Fe(CN)<sub>6</sub>]<sup>3-</sup> metallotectons and diverse organic cations: Crystal structure, Hirshfeld surface analysis,

spectroscopic and thermal properties. *Inorganica Chimica Acta*, 486, 36-47.

Kuberi, J. R., Nannim, S., Daboro, P. C., & Kim, I. B. (2024). FRUIT YIELD AND QUALITY OF STRAWBERRY AS INFLUENCED BY VARIETY AND NUTRIENT SOURCE IN JOS, NIGERIA. *Nigerian Journal of Horticultural Science*, 28(3).

Larrañeta, E. (2018). *Hydrogels for Hydrophobic Drug Delivery. Classification, Synthesis and Applications*. <https://doi.org/10.3390/jfb9010013>

Li, S., & Chen, G. (2018). *Using hydrogel-biochar composites for enhanced cadmium removal from aqueous media*. (February 2019), 2–7. <https://doi.org/10.15406/mseij.2018.02.00073>

Lonappan, L., Liu, Y., Rouissi, T., Brar, S. K., & Surampalli, R. Y. (2019). Development of biochar-based green functional materials using. *Journal of Cleaner Production*. <https://doi.org/10.1016/j.jclepro.2019.118841>

Ma, J., Li, T., Liu, Y., Cai, T., Wei, Y., & Dong, W. (2019). Bioresource Technology Rice husk derived double network hydrogel as efficient adsorbent for Pb (II), Cu (II) and Cd (II) removal in individual and multicomponent systems. *Bioresource Technology*, 290(July), 121793. <https://doi.org/10.1016/j.biortech.2019.121793>

Martínez-Villadiego, K., Arias-Tapia, M. J., & Jaramillo, A. F. (2024). Improving thermal stability of starches native cross-linked with citric acid as a compatibilizer for thermoplastic starch/polylactic acid blends. *Polymer Bulletin*, 1-22.

Mwangi, I., Kiriho, G., Swaleh, S., Wanjau, R., Mbugua, P., & Catherine, J. (2019). Remediation of degraded soils with hydrogels from domestic animal wastes. *International Journal of Recycling of Organic Waste in Agriculture*, 8(2), 159–170. <https://doi.org/10.1007/s40093-019-0242-1>

Nandiyanto, A. B. D., Oktiani, R., & Ragadhita, R. (2019). How to read and interpret FTIR spectroscopy of organic material. *Indonesian Journal of Science and Technology*, 4(1), 97-118.

Pattanaik, B. P., & Misra, R. D. (2019). Effect of reaction pathway and operating parameters on the deoxygenation of vegetable oils to produce diesel range hydrocarbon fuels: A review. Effect of reaction pathway and operating parameters on the deoxygenation of vegetable oils to produce diesel range hydrocarbon fuels: A review. *Renewable and Sustainable Energy Reviews*, 73(February 2017), 545–557. <https://doi.org/10.1016/j.rser.2017.01.018>

Petit, A., Trinquand, A., Chevret, S., Ballerini, P., Cayuela, J. M., Grardel, N., ... & Asnafi, V. (2018). Oncogenetic mutations combined with MRD improve outcome prediction in pediatric T-cell acute lymphoblastic leukemia. *Blood, The Journal of the American Society of Hematology*, 131(3), 289-300.

- Bui, M., Adjiman, C. S., Bardow, A., Anthony, E. J., Boston, A., Brown, S., ... & Mac Dowell, N. (2018). Carbon capture and storage (CCS): the way forward. *Energy & Environmental Science*, 11(5), 1062–1176.
- Randhawa, G. K., & Kullar, J. S. (2011). *Bioremediation of Pharmaceuticals, Pesticides, and Petrochemicals with Gomeya/ Cow Dung*. 2011. <https://doi.org/10.5402/2011/362459>
- Richardson, D. M., Carruthers, J., Hui, C., Impson, F. A. C., Miller, J. T., Robertson, M. P., ... Wilson, J. R. U. (2011). Human-mediated introductions of Australian acacias. *A journal of conservation Biogeography*. 771–787. <https://doi.org/10.1111/j.1472-4642.2011.00824.x>
- Singh, A. A., Agrawal, S. B., & Shahi, J. P. (2014). *Assessment of growth and yield losses in two Zea mays L. cultivars (quality protein maize and nonquality protein maize) under projected levels of ozone*. 2628–2641. <https://doi.org/10.1007/s11356-013-2188-6>
- Ullah, F., Bisyrul, M., Javed, F., & Akil, H. (2015). *Classification, processing and application of hydrogels: A review*. 57, 414–433. <https://doi.org/10.1016/j.msec.2015.07.053>
- Usman, A. R. A., & Ahmad, M. (2015). Chemically modified biochar produced from conocarpus waste increases NO<sub>3</sub> removal from aqueous solutions. *Environmental Geochemistry and Health*, (June). <https://doi.org/10.1007/s10653-015-9736-6>
- Utomo, W. H., Nugroho, W. H., & Kusuma, Z. (2011). *Simple biochar production generated from cattle dung and coconut*. 1(10), 1680–1685.
- Wells, M. S., Brinton, C. M., & Reberg-Horton, S. C. (2016). Weed suppression and soybean yield in a no-till cover-crop mulched system as influenced by six rye cultivars. *Renewable Agriculture and Food Systems*, 31(5), 429–440. <https://doi.org/10.1017/S1742170515000344>
- Wu, Y., Brickler, C., Li, S., & Chen, G. (2021). Synthesis of microwave-mediated biochar-hydrogel composites for enhanced water absorbency and nitrogen release. *Polymer Testing*, 93(August 2020), 106996. <https://doi.org/10.1016/j.polymertesting.2020.106996>
- Yakout, S. M., El, A., Daifullah, H. M., & El-reefy, S. A. (2015). *Pore structure characterization of chemically modified biochar derived from rice straw*. 14(2), 473–480.
- Zahra, S. I., Abbas, F., Ishaq, W., Ibrahim, M., Hammad, H. M., Akram, B., & Salik, M. R. (2016). Carbon Sequestration potential of soils under maize production in Irrigated Agriculture of the Punjab province of Pakistan.
- Zhang, L., Liu, Y., Wang, Y., Li, X., & Wang, Y. (2021). Investigation of phosphate removal mechanisms by a lanthanum hydroxide adsorbent using p-XRD, FTIR and XPS. *Applied Surface Science*, 557, 149838.

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