

1 **Nephroprotective Potential of Biskhapra (*Trianthemafortulacastrum* Linn.) Leaves Extract in**  
2 **Cisplatin-Induced Acute Kidney Injury in Wistar Albino Rats: An Experimental Study**

3 **ABSTRACT**

4 The kidneys are essential organs responsible for excretion and maintaining homeostasis. Acute kidney  
5 injury (AKI) is a severe clinical condition characterized by sudden deterioration in renal function,  
6 often resulting from nephrotoxic agents such as cisplatin, which is widely used in chemotherapy.  
7 Cisplatin-induced nephrotoxicity is a significant clinical limitation, with oxidative stress,  
8 inflammation, and tubular damage being the primary mechanisms involved. The present study aimed  
9 to evaluate the nephroprotective potential of aqueous and hydro-alcoholic extracts of Biskhapra  
10 (*Trianthemafortulacastrum* Linn.) leaves in a cisplatin-induced AKI rat model.

11 The experimental study involved forty healthy Wistar albino rats (150–200 g) divided into five groups  
12 (n=8). Group I served as plain control, receiving distilled water; Group II was the negative control,  
13 receiving a single intraperitoneal injection of cisplatin (5 mg/kg b.w.); Group III was treated with  
14 cisplatin plus standard nephroprotective agent silymarin (50 mg/kg p.o.); Group IV and Group V  
15 received cisplatin plus aqueous and hydro-alcoholic extracts of Biskhapra leaves (450 mg/kg p.o.),  
16 respectively, for 14 days.

17 Renal function was assessed by serum creatinine, blood urea, blood urea nitrogen (BUN), serum uric  
18 acid, total serum protein, serum albumin, and serum globulin. Histopathological examination of kidney  
19 tissue was performed to assess structural changes. Physiochemical and phytochemical analyses of the  
20 extracts were conducted, including moisture content, pH, extractive values, ash values, and HPTLC  
21 profiling.

22 Results demonstrated a significant rise in serum creatinine, blood urea, and BUN in the cisplatin  
23 group, confirming renal injury. Treatment with Biskhapra extracts significantly reduced these levels  
24 ( $p < 0.001$ ), comparable to the standard control. Serum protein levels, albumin, and globulin, which  
25 were reduced in nephrotoxic rats, were restored upon treatment. Histopathology revealed marked

26 tubular regeneration and reduced necrosis in treated groups. Phytochemical screening confirmed the  
27 presence of flavonoids, alkaloids, tannins, saponins, steroids, and phenolics, which may contribute to  
28 the observed nephroprotective effect.

29 In conclusion, both aqueous and hydro-alcoholic extracts of Biskhapra leaves exhibit significant  
30 nephroprotective activity against cisplatin-induced AKI in rats, likely due to antioxidant, anti-  
31 inflammatory, and regenerative properties. This study validates the traditional Unani claim of  
32 Biskhapra as a MuqavviGurdawaMasana (renal tonic) and provides experimental evidence for its  
33 potential therapeutic application in AKI management.

34 **Keywords:**Biskhapra, Triantheportulacastrum, Cisplatin, Acute Kidney Injury

35

36

37

38

39

40

41

42

### 43 **Introduction**

44 The kidneys are vital organs that maintain internal stability by regulating fluid balance, electrolyte  
45 composition, acid-base homeostasis, and the elimination of metabolic waste products.<sup>1</sup>They play a

46 central role in the excretory system and are crucial for maintaining the biochemical equilibrium of the  
47 body. Disruption of kidney function has profound systemic effects, and acute kidney injury (AKI)  
48 represents a sudden, often reversible deterioration of renal function.<sup>2</sup> AKI, formerly referred to as acute  
49 renal failure, is defined clinically by a rapid rise in serum creatinine and/or decreased urine output.<sup>3</sup>

50 Globally, AKI is a significant health problem, with an incidence ranging from 2% to 7% of hospital  
51 admissions and affecting up to 67% of critically ill patients in intensive care units.<sup>3,4</sup> The condition is  
52 associated with increased morbidity, prolonged hospitalization, high healthcare costs, and elevated  
53 mortality rates.<sup>5</sup> AKI can be caused by prerenal, intrarenal, or postrenal factors. Acute tubular necrosis  
54 (ATN), the most common cause of intrinsic AKI, occurs due to ischemic or nephrotoxic injury.<sup>3</sup>  
55 Nephrotoxic drugs, including cisplatin, aminoglycosides, and nonsteroidal anti-inflammatory drugs  
56 (NSAIDs), directly damage renal tubular cells. Cisplatin, a platinum-based chemotherapeutic agent, is  
57 highly effective against solid tumors but is limited by its nephrotoxicity. Cisplatin-induced AKI is  
58 characterized by oxidative stress, mitochondrial dysfunction, DNA damage, inflammation, apoptosis,  
59 and reduced renal perfusion. Tubular epithelial cell death and inflammatory infiltration lead to  
60 impaired filtration and reabsorption, resulting in elevated serum creatinine, blood urea, and  
61 disturbances in protein metabolism.<sup>6</sup>

62 The management of AKI primarily involves addressing the underlying cause, supportive care, and  
63 renal replacement therapy in severe cases. Pharmacological interventions are limited to symptomatic  
64 relief and do not reverse the underlying injury. Dialysis remains the mainstay in advanced AKI but has  
65 limitations including invasiveness, cost, and complications. Furthermore, current treatments do not  
66 promote renal tissue regeneration, making the search for effective nephroprotective agents an urgent  
67 need.<sup>3</sup>

68 Alternative and complementary medicine systems, including Unani medicine, have long recognized  
69 the importance of natural remedies in preserving organ function. In the Unani system of medicine, the  
70 concept of *MuqavviAaza* (organ tonics) emphasizes the maintenance and restoration of organ function  
71 by normalizing the temperament (*mizaj*) of specific organs, thus enhancing their resistance to  
72 pathological influences. IbnSina defined a *Muqavvi* drug as one that strengthens an organ either  
73 through its intrinsic nature or by balancing its temperament—by cooling excessive heat or warming  
74 excessive cold. Galen also supported this view, using rose oil as an example of a drug that equilibrates  
75 temperament.<sup>7</sup> Within this framework, *MuqavviGurdawaMasana* (tonics for the kidneys and urinary  
76 bladder) are recognized for their nephroprotective properties. Many Unani and herbal drugs such as  
77 *Emblicaofficinalis* (Amla), *Punicagranatum* (Anar), *Cymbopogonjwarancusa* (Izkhar), *Artemisia*  
78 *absinthium* (Afsanteen), *Matricariachamomilla* (Baboona), and *Cinnamomumzeylanicum* (Darchini)  
79 are traditionally used to strengthen renal and urinary functions, along with indirect tonics like *Crocus*  
80 *sativus* (Zafran) and *Strychnosnux-vomica* (Kuchla).<sup>8</sup>

81 A wide variety of Unani and herbal plants have demonstrated nephroprotective, diuretic, and anti-  
82 inflammatory effects in both traditional practice and modern research. Seeds of  
83 *Peucedanumgraveolens* (TukhmeShibbat), *Daucuscarota* (Gazar), and *Trachyspermumammi* (Ajwain)  
84 are known for their diuretic and lithotriptic actions.<sup>7,9</sup> Plants like *Aerva lanata*,<sup>10</sup> *Cordyceps sinensis*,  
85 and *Trianthemaportulacastrum* (Biskhapra) have been shown to mitigate drug-induced nephrotoxicity  
86 and enhance renal recovery.<sup>11,12</sup> Similarly, *Tribulusterrestris* (Gokhru), *Icacinatricantha*, and  
87 *Raphanussativus* (Turb) exhibit protective effects on renal histology and function in experimental  
88 models.<sup>13,14</sup> Herbs such as *Taraxacumofficinale*, *Asparagus racemosus* (Satavar), *Ocimum sanctum*  
89 (Reehan), *Moringaoleifera* (Sahajna), and *Tinosporacordifolia* (Giloy) further contribute to renal  
90 protection through antioxidant and anti-inflammatory pathways.<sup>14,15</sup>

91 Additionally, compound and polyherbal Unani formulations have shown promising nephroprotective  
92 outcomes. Formulations like “Banadequl Buzoor,”<sup>16,17</sup> “NR-AG I,” “NR-AG II,” and  
93 “Jawarish Zarooni Sada” have been effective in preventing drug-induced renal injury and normalizing  
94 serum urea and creatinine levels.<sup>18,19</sup> Natural antioxidants such as ginsenoside Rb-I, quercetin, and  
95 herbal extracts like *Withania somnifera* (Asgandh) and *Echinacea pallida* have demonstrated potent  
96 protection against oxidative renal damage.<sup>20,21</sup> These findings collectively validate the traditional  
97 Unani principle of *Muqavvi Gurdawa Masana* and highlight the therapeutic potential of herbal  
98 nephroprotective agents in both preventive and restorative renal health.

99 *Biskhapra* (*Trianthem portulacastrum* Linn.), a member of the Aizoaceae family, is recognized by  
100 various names across languages—*Hand Qooqin* Arabic, *Dewasaptin* Persian, and *Horse Purslane* in  
101 English. It is an annual prostrate herbaceous plant, typically extending 4 to 6 feet in length, and is  
102 commonly found in moist environments such as riverbanks or near ponds. This species flourishes as a  
103 weed in both cultivated fields and wastelands across tropical and subtropical regions, with widespread  
104 occurrence throughout India.<sup>22</sup> It is recognized for its diuretic, resolvent (mohallil), astringent (qabiz),  
105 and calorific (musakkhin) properties. The fresh juice of the plant is traditionally administered for  
106 treating night blindness, ocular ulcers, and urinary dribbling.<sup>9,22,23</sup> In Unani literature, it is considered  
107 effective in treating conditions involving fluid retention, urinary disorders, and kidney dysfunction.  
108 The temperament of leaves and seeds of *Biskhapra* are Hot in second and Dry in first degree. The dose of  
109 *Biskhapra* leaf is 1 tola.<sup>7,9</sup>

110 Modern pharmacological studies have confirmed its antioxidant, anti-inflammatory, and diuretic  
111 actions, supporting its traditional use.<sup>22,23</sup> Phytochemical analysis reveals the presence of alkaloids,  
112 flavonoids, tannins, saponins, steroids, and phenolics, compounds known to exert protective effects  
113 against oxidative stress and inflammation.<sup>24,25</sup> Given the high incidence of AKI and the limitations of  
114 conventional therapies, there is a pressing need to explore alternative nephroprotective agents.

115 Biskhapra, with its rich history in traditional medicine and documented pharmacological activity, is a  
116 promising candidate. Prior studies have demonstrated its protective effects in adriamycin-induced  
117 nephrotic syndrome, but there is limited research on its role in cisplatin-induced AKI. This study aims  
118 to fill that gap by systematically evaluating the nephroprotective potential of aqueous and hydro-  
119 alcoholic extracts of Biskhapra leaves in a cisplatin-induced rat model. The primary objective of the  
120 study is to assess the efficacy of Biskhapra extracts in mitigating renal injury induced by cisplatin.  
121 Specific objectives include evaluating renal function parameters including serum creatinine, blood  
122 urea, BUN, serum uric acid, serum albumin, serum globulin, and total protein; conducting  
123 histopathological examination of kidney tissue to observe morphological changes; and performing  
124 physiochemical and phytochemical standardization of Biskhapra leaf extracts. This research provides  
125 experimental evidence validating the traditional Unani claim of Biskhapra as a nephroprotective agent.  
126 It could pave the way for developing cost-effective, plant-based therapies for AKI, reducing reliance  
127 on dialysis and improving patient outcomes. Furthermore, standardization of the drug through  
128 physiochemical and HPTLC analysis ensures reproducibility and quality control, laying a foundation  
129 for future pharmacological and clinical studies.

## 130 **Materials and methods**

### 131 **Study Design**

132 This experimental study was designed to evaluate the nephroprotective potential of aqueous and  
133 hydro-alcoholic extracts of *Trianthemaportulacastrum* Linn. (Biskhapra) leaves against cisplatin-  
134 induced acute kidney injury (AKI) in Wistar albino rats. The study was conducted in accordance with  
135 the institutional ethical guidelines, and approval was obtained from the Institutional Animal Ethics  
136 Committee (IAEC) of Bihar Animal Sciences University, Patna.

### 137 **Plant Material Collection and Authentication**

138 Fresh leaves of *Trianthemaportulacastrum* Linn. were collected from authenticated sources in the local  
139 area. The plant was identified and authenticated by a botanist Dr Ajay Kumar (Chief Scientist)  
140 Cognosmed Laboratories Pvt. Ltd. and a voucher specimen was deposited for reference. The leaves  
141 were washed thoroughly under running water to remove dust and impurities and shade-dried for 10–12  
142 days to preserve the phytochemical constituents. The dried leaves were then powdered using a  
143 mechanical grinder and stored in airtight containers at room temperature until further use.

144 **Preparation of extract:** Biskhapra was manually cleaned from any inorganic matters and other weeds  
145 and coarsely ground into powder by using an electric grinder in pharmacy of Govt Tibbi College and  
146 Hospital Patna. Two different extracts of aqueous and hydro-alcoholic solvents were extracted. The  
147 extracts were prepared by two different methods using maceration method and reflux method to check  
148 for the higher yield percentage.



149

150 **Figure: 01 Aqueous and Hydro-alcoholic extract of Biskhapra**

151 **Aqueous extract by maceration method:**

152 Crude powder drug (25 gram) was taken along with 400 ml of distilled water in a beaker and left for  
153 48 hours untouched in cool and dark place. The drug was then subjected to sonicator for sonication  
154 process for half an hour. After which it was filtered and kept for evaporation in a China petridish in a

155 water bath at 70°C and dried until extract was obtained. After cooling down, the extract was weighed  
156 to calculate the extractive yield and it stored in an air tight container for phytochemical screening and  
157 further study.<sup>26</sup>

#### 158 **Hydro-alcoholic extract by maceration method:**

159 Test drug powder (25 gm) was taken along with 200 ml distilled water and 200 ml methanol in a beaker  
160 and left for 48 hours untouched in cool and dark place. The drug was then subjected to sonicator for  
161 sonication process for half an hour. After which it was filtered and kept for evaporation in a Chinese  
162 dish in a water bath at 70°C and dried until the extract was finally obtained. After cooling down, the extract  
163 was weighed to calculate the extractive yield and stored in an air tight container for phytochemical  
164 screening.<sup>26</sup>

165 Physicochemical parameters such as extractive values, ash values, moisture content, and pH were  
166 determined according to standard pharmacognostic procedures.  
167 Preliminary phytochemical screening was performed to detect the presence of alkaloids, flavonoids,  
168 saponins, steroids, tannins, phenolics, and carbohydrates.  
169 Additionally, High-Performance Thin-Layer Chromatography (HPTLC) profiling was conducted for  
170 quality standardization and to identify chemical fingerprints of the extracts.

#### 171 **Experimental Animals**

172 Forty healthy Wistar albino rats (150–200 g) of either sex were procured from the Central Animal  
173 Research Facility, BASU, Patna. The animals were acclimatized for five days under standard  
174 laboratory conditions: temperature  $23 \pm 2^\circ\text{C}$ , humidity almost  $55 \pm 15\%$ , and a 12-hour light/dark  
175 cycle. They were provided free access to a standard pellet diet and water ad libitum.



176 Before drug administration, all animals were fasted overnight with free access to water to ensure  
177 uniform metabolic conditions.

### 178 **Experimental Grouping**

179 The animals were randomly divided into five groups, each consisting of eight rats (n=8):

180 ❖ Group I – Plain Control: Received distilled water (3.0 ml, orally) once daily for 12 days.

181 ❖ Group II – Negative Control: Received a single intraperitoneal injection of cisplatin (5 mg/kg body  
182 weight) on day 1.

183 ❖ Group III – Standard Control: Received cisplatin (5 mg/kg, i.p.) plus silymarin (50 mg/kg, orally) once  
184 daily for 14 days.

185 ❖ Group IV – Test Group A: Received cisplatin (5 mg/kg, i.p.) plus aqueous extract of *Biskhapra* leaves  
186 (450 mg/kg, orally) once daily for 14 days.

187 ❖ Group V – Test Group B: Received cisplatin (5 mg/kg, i.p.) plus hydro-alcoholic extract of *Biskhapra*  
188 leaves (450 mg/kg, orally) once daily for 14 days.

### 189 **Induction of Acute Kidney Injury (AKI)**

190 Acute kidney injury was induced by a single intraperitoneal injection of cisplatin (5 mg/kg body  
191 weight) in Groups II, III, IV, and V on day 1 of the experiment. This dose and route of administration  
192 were selected based on previous literature reports demonstrating consistent and reproducible induction  
193 of nephrotoxicity in experimental rat models.

### 194 **Biochemical Analysis**

195 At the end of the treatment period, all animals were anesthetized, and blood samples were collected via  
196 retro-orbital plexus puncture using sterile capillary tubes. The collected blood samples were allowed to  
197 clot at room temperature and then centrifuged at 3000 rpm for 15 minutes to separate the serum. The  
198 serum samples were analyzed for renal function markers, including:

199❖ Serum creatinine

200❖ Blood urea

201❖ Blood urea nitrogen (BUN)

202❖ Serum uric acid

203❖ Serum albumin

204❖ Serum globulin

205❖ Total serum protein

206 All biochemical estimations were performed using standard diagnostic kits (Manufacturer: XXX) and  
207 analyzed using an automated biochemical analyzer following the manufacturer's instructions.

### 208 **Histopathological Examination**

209 After completion of the treatment protocol, animals were sacrificed on the 14th day by cervical  
210 dislocation under anesthesia. Both kidneys were carefully excised, washed with ice-cold normal saline,  
211 and immediately fixed in 10% formalin solution.

212 Tissue specimens were processed through dehydration, clearing, and paraffin embedding. Thin  
213 sections of 5  $\mu\text{m}$  thickness were prepared and stained with hematoxylin and eosin (H&E) for  
214 microscopic examination.

215 Histopathological evaluation was performed to assess tubular necrosis, degeneration, glomerular  
216 changes, interstitial inflammation, and overall renal architecture, and scoring was done according to  
217 standard histological grading methods.

## 218 **Statistical Analysis**

219 All data were expressed as mean  $\pm$  standard deviation (SD). Statistical analysis was performed using  
220 one-way analysis of variance (ANOVA) followed by Tukey's post hoc test for multiple comparisons  
221 among groups. A p-value  $< 0.05$  was considered statistically significant. Data visualization and  
222 tabulation were done using GraphPad Prism software.

## 223 **Results& Observation**

224 The physicochemical and phytochemical analysis of *Trianthemapotulacastrum* Linn. (Biskhapra)  
225 leaves revealed valuable information regarding its composition and quality parameters.

## 226 **PhysicoChemicalStudies**

227 The extractive values were recorded as 28.00% w/w for the hydro-alcoholic extract and 27.24% w/w  
228 for the aqueous extract. The moisture content was found to be 13.6% w/w, while total ash, acid-  
229 insoluble ash, and water-soluble ash were 18.91% w/w, 1.78% w/w, and 13.20% w/w, respectively.  
230 The pH of a 5% aqueous solution of Biskhapra leaves was 6.36, indicating a slightly acidic nature  
231 suitable for biological compatibility. Preliminary phytochemical screening confirmed the presence of  
232 alkaloids, flavonoids, tannins, saponins, steroids, phenolics, proteins, and reducing sugars, which are  
233 known to contribute to antioxidant and nephroprotective effects. HPTLC profiling showed  
234 characteristic peaks at R<sub>f</sub> values 0.04 and 0.05 under UV 254 nm and 366 nm, serving as a  
235 chromatographic fingerprint for standardization and quality control.

## 236 **Blood Urea**

237 In the plain control group (Group I), blood urea was  $40.85 \pm 0.576$  mg/dl. In the negative control group  
238 (Group II) treated with cisplatin (5 mg/kg b.w.), the value rose markedly to  $91.86 \pm 0.822$  mg/dl ( $p <$   
239  $0.001$ ), indicating severe nephrotoxicity. The standard control group (Group III) treated with silymarin  
240 showed  $45.25 \pm 0.485$  mg/dl, significantly lower than the negative control ( $p < 0.001$ ). The aqueous  
241 extract-treated group (Group IV) showed  $52.55 \pm 0.499$  mg/dl ( $p < 0.001$  vs. negative control), while  
242 the hydro-alcoholic extract-treated group (Group V) demonstrated  $45.4 \pm 0.541$  mg/dl ( $p < 0.01$ ),  
243 indicating a marked reduction in blood urea. [Table No-01]

#### 244 **Serum Creatinine**

245 Serum creatinine in the plain control group was  $0.75 \pm 0.032$  mg/dl, while in the negative control it  
246 increased significantly to  $3.08 \pm 0.056$  mg/dl ( $p < 0.001$ ). The standard control group recorded  $0.85 \pm$   
247  $0.012$  mg/dl ( $p < 0.001$  vs. negative control). Treatment with aqueous extract (450 mg/kg b.w.) reduced  
248 serum creatinine to  $1.50 \pm 0.139$  mg/dl ( $p < 0.001$ ), and hydro-alcoholic extract treatment showed  $0.98$   
249  $\pm 0.009$  mg/dl ( $p < 0.01$ ), confirming significant nephroprotection. [Table No-01]

#### 250 **Blood Urea Nitrogen (BUN)**

251 BUN in the plain control was  $19.04 \pm 0.259$  mg/dl, which increased to  $42.85 \pm 0.379$  mg/dl in the  
252 negative control group ( $p < 0.001$ ). The standard control group showed  **$21.06 \pm 0.222$  mg/dl** ( $p <$   
253  $0.001$ ). Treatment with aqueous extract resulted in  $24.49 \pm 0.23$  mg/dl ( $p < 0.001$ ), while the hydro-  
254 alcoholic extract group showed  $21.14 \pm 0.25$  mg/dl ( $p < 0.01$ ), both showing significant improvement  
255 compared with the negative control. [Table No-01]

#### 256 **Serum Uric Acid**

257 The plain control group exhibited  $2.10 \pm 0.047$  mg/dl, while the negative control recorded  $2.41 \pm 0.093$   
258 mg/dl ( $p < 0.001$ ). Standard control (silymarin) showed  $2.14 \pm 0.157$  mg/dl ( $p < 0.001$  vs. negative

259 control). The aqueous extract group showed  $2.07 \pm 0.116$  mg/dl ( $p < 0.001$ ), and the hydro-alcoholic  
 260 extract group  $2.38 \pm 0.179$  mg/dl ( $p < 0.01$ ). [Table No-01]

261 **Serum Protein**

262 Serum protein in the plain control group was  $7.21 \pm 0.097$  mg/dl, which decreased significantly to  $4.97$   
 263  $\pm 0.090$  mg/dl in the negative control group ( $p < 0.001$ ). The standard control group recorded  $6.55 \pm$   
 264  $0.106$  mg/dl ( $p < 0.001$  vs. negative control). The aqueous extract group showed  $5.33 \pm 0.092$  mg/dl ( $p$   
 265  $< 0.001$ ), and the hydro-alcoholic extract group  $6.05 \pm 0.086$  mg/dl ( $p < 0.01$ ), indicating significant  
 266 recovery of serum protein levels. [Table No-01]

267 **Serum Albumin**

268 Serum albumin in the plain control was  $3.40 \pm 0.064$  mg/dl, while in the negative control it decreased  
 269 to  $2.49 \pm 0.063$  mg/dl ( $p < 0.001$ ). The standard control group recorded  $3.53 \pm 0.025$  mg/dl ( $p < 0.001$   
 270 vs. negative control). Aqueous extract treatment showed  $2.59 \pm 0.071$  mg/dl ( $p < 0.001$ ), while the  
 271 hydro-alcoholic extract group recorded  $3.13 \pm 0.067$  mg/dl ( $p < 0.001$ ). [Table No-01]

272 **Serum Globulin**

273 Serum globulin in the plain control group was  $3.81 \pm 0.064$  mg/dl, which reduced to  $2.49 \pm 0.063$   
 274 mg/dl in the negative control ( $p < 0.001$ ). The standard control group showed  $3.53 \pm 0.025$  mg/dl ( $p <$   
 275  $0.001$ ). Aqueous extract treatment resulted in  $2.59 \pm 0.071$  mg/dl ( $p < 0.05$ ), while the hydro-alcoholic  
 276 extract showed  $3.13 \pm 0.067$  mg/dl ( $p < 0.01$ ). [Table No-01]

277 Table01:Effectofaqueousextractandhydro-  
 278 alcoholicextractofBiskhapra(*Trianthemaportulacastrum*Linn.)leavesextractoncisplatin induced acute  
 279 kidney injury.

Groups	Agents	S.Creatinine( mg/dl)	BloodUr ea	BUN(gm/ dl)	SerumU ric Acid	S.Protei n	S.Album in	S.Globu lin
--------	--------	-------------------------	---------------	----------------	--------------------	---------------	---------------	----------------

			(mg/dl)		(gm/dl)	(gm/dl)	(gm/dl)	(mg/dl)
<b>Plain control</b>	Distilled water 3.0ml	0.75±0.032	40.85±0.576	19.037±0.259	2.098±0.047	7.21±0.097	4.025±0.064	3.18±0.081
<b>Negative control</b>	Cisplatin 5mg/kg	3.08±0.056 a***	91.86±0.822 a***	42.85±0.379 a***	2.412±0.093 a#	4.97±0.090 a***	2.487±0.063 a***	2.48±0.058 a***
<b>Standard Control</b>	Cisplatin 5mg/kg Silymarin 50mg/kg	0.85±0.012 b***	45.25±0.485 b***	21.062±0.222 b***	2.14±0.157 b#	6.55±0.106 b***	3.525±0.025 b***	3.01±0.093 b***
<b>Test group A</b>	Cisplatin 5mg/kg Plus Aqueous extract of leaves of Biskhapra 450mg/kg	1.50±0.139 b***	52.55±0.499 b***	24.487±0.23 b***	2.07±0.116 b#	5.33±0.092 b***	2.587±0.071 b***	2.75±0.046 b*
<b>Test group B</b>	Cisplatin 5mg/kg Plus Hydro- alcoholic extract of leaves of Biskhapra 450mg/kg	0.98±0.009 b**	45.4±0.541 b**	21.137±0.25 b***	2.38±0.179 b#	6.05±0.086 b**	3.125±0.067 b***	2.92±0.070 b**

280 Values expressed in MEAN±SEM, #-

281 nonsignificant, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, n=10 in each group, test used ANOVA one way followed by

282 Tukey Kramer comparison test. a- Vs Plain control and b-Negative control A.

### 283 Histopathological Examination of rat kidney

284 Histopathological observations strongly supported biochemical findings. The negative control group

285 revealed extensive tubular necrosis, epithelial degeneration, and mononuclear infiltration, confirming

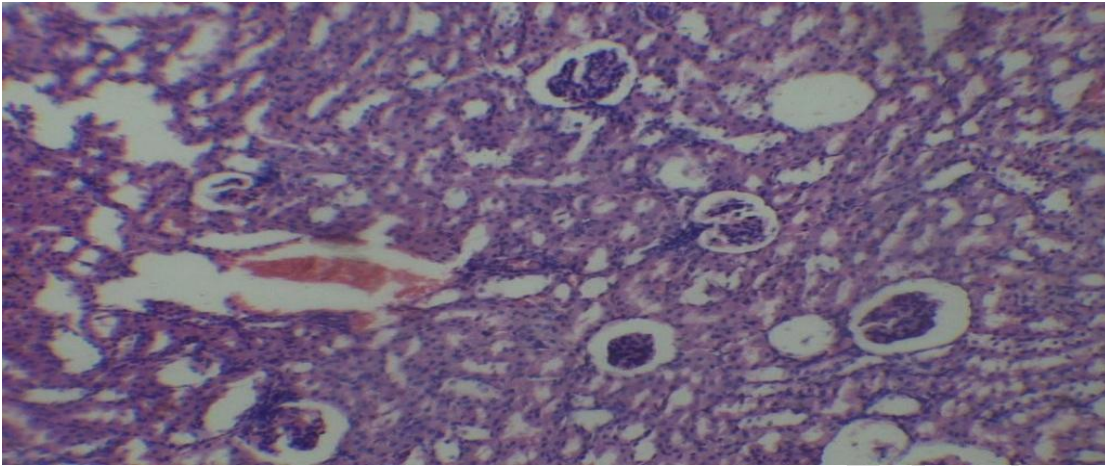
286 severe renal damage. In contrast, the plain control group exhibited normal renal histology with intact

287 glomeruli and tubular epithelium. The silymarin-treated group showed regenerative changes in tubular

288 epithelial cells and reduction in inflammation. Notably, the Biskhapra-treated groups displayed

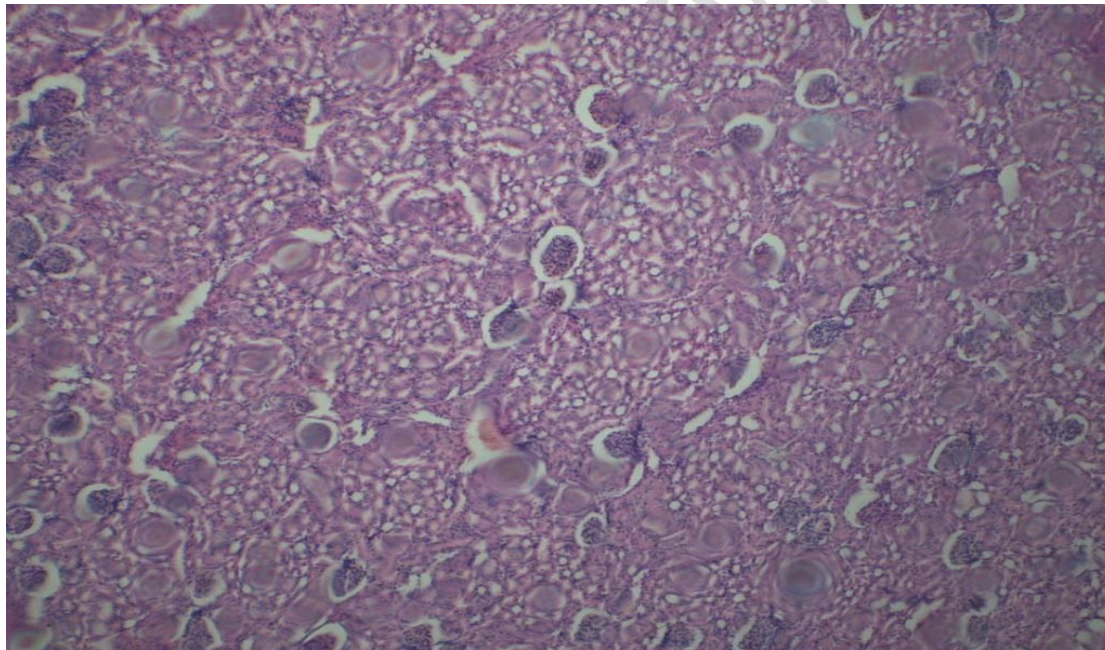
289 significant renal protection: both extracts reduced tubular necrosis, restored epithelial integrity, and

290 preserved glomerular architecture. Among the two, the aqueous extract (Group IV) showed slightly  
291 better histological recovery than the hydro-alcoholic extract (Group V). [Figures 2-6]



292

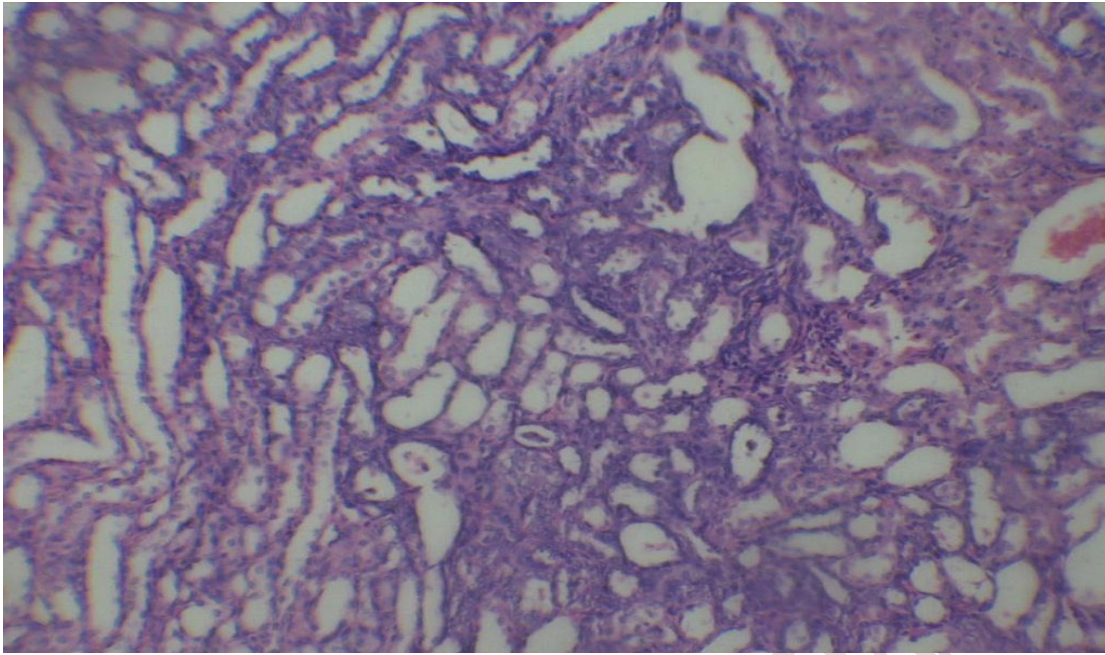
293 Figure: 2 Plain control group showing normal structure of the rat kidney.



294

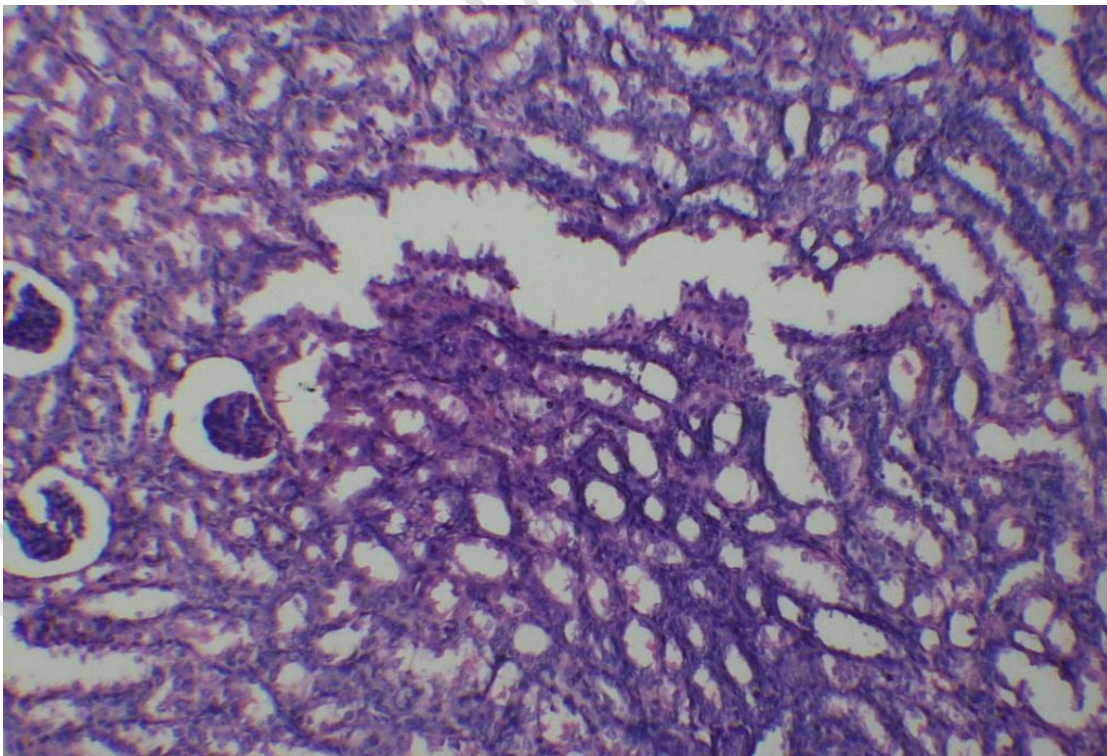
295 Figure: 3 Negative group showing degeneration and tubular necrosis with mononuclear cell infiltration.





296

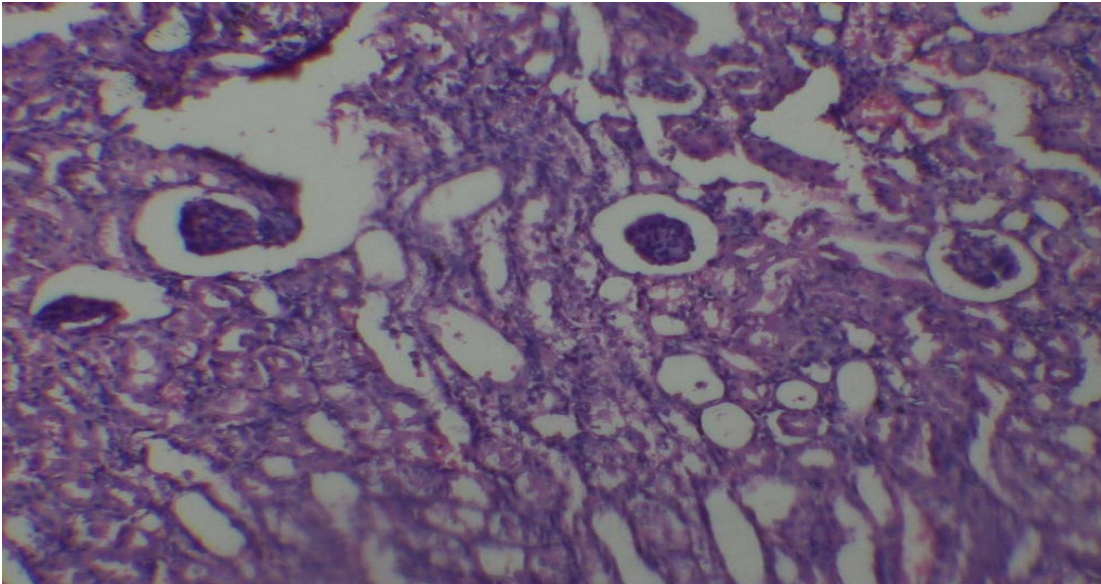
297 **Figure: 4 Standard control group showing regenerative tubular epithelium mild to moderate**  
298 **glomeruli and tubular architecture.**



299

300 **Figure: 5 group A showing regeneration of epithelium lining.**





301

302 **Figure: 6 Testgroup B showing mild regeneration of tubular cells and surrounding cytoplasm.**

303 In comparative efficacy, both aqueous and hydro-alcoholic extracts of Biskhapra demonstrated marked  
304 nephroprotective effects against cisplatin-induced renal injury. However, the aqueous extract exhibited  
305 superior activity across biochemical and histopathological parameters, suggesting greater stability and  
306 bioavailability of its active constituents. These findings collectively validate the traditional Unani use  
307 of Biskhapra as *Muqavvi Gurdawa Masana* (kidney and bladder tonic) and affirm its potential as a  
308 natural nephroprotective agent.

### 309 **Discussion**

310 Acute Kidney Injury (AKI) is a serious clinical disorder marked by a sudden decline in renal function,  
311 often leading to high morbidity and mortality.<sup>2</sup>In this study, cisplatin was used to induce AKI in Wistar  
312 albino rats, providing a reliable experimental model for evaluating nephroprotective agents. Cisplatin-  
313 induced AKI closely resembles the pathological features seen in humans, such as tubular necrosis,  
314 elevated serum creatinine and urea, and altered protein levels.<sup>28</sup>The present study demonstrated that  
315 both aqueous and hydro-alcoholic extracts of *Trianthem portulacastrum* Linn. (Biskhapra)

316 significantly attenuated the biochemical and histopathological alterations caused by cisplatin, thus  
317 scientifically validating its traditional Unani use as a *MuqavviGurdawaMasana* (kidney and bladder  
318 tonic).

319 The pathogenesis of cisplatin-induced AKI involves oxidative stress, mitochondrial dysfunction,  
320 inflammation, and apoptosis.<sup>28</sup> Cisplatin accumulates in proximal tubular cells, generating reactive  
321 oxygen species (ROS) that activate MAPK pathways, resulting in DNA damage and tubular epithelial  
322 cell death. These effects impair filtration and excretion functions, leading to tubular necrosis and  
323 glomerular degeneration, which were clearly observed in the negative control group.<sup>29</sup> In contrast,  
324 Biskhapra-treated groups showed marked improvement in both biochemical and histological  
325 parameters. The aqueous extract was slightly more effective than the hydro-alcoholic extract, possibly  
326 due to better preservation and bioavailability of active constituents in water-based preparations.  
327 Restoration of serum creatinine, urea, and total protein levels in treated rats reflected improved  
328 glomerular filtration and renal recovery. These results align with the findings of Karimet. al  
329 substantiating the use of Biskhapra extracts in ameliorating blood urea levels under nephrotoxic  
330 conditions.<sup>32</sup>

331 Phytochemical analysis confirmed the presence of alkaloids, flavonoids, tannins, saponins, steroids,  
332 and phenolic compounds in Biskhapra leaves. These constituents are known for potent antioxidant and  
333 anti-inflammatory activities. Flavonoids and phenolics neutralize free radicals, reducing lipid  
334 peroxidation and oxidative damage, while alkaloids such as punarnavine protect renal tissues from  
335 oxidative stress. Tannins and saponins stabilize cell membranes, decrease inflammation, and contribute  
336 to structural recovery. Thus, the synergistic action of these phytochemicals accounts for the observed  
337 nephroprotective effects.

338 Pharmacologically, the nephroprotective potential of Biskhapra arises from its antioxidant, anti-  
339 inflammatory, and diuretic properties.<sup>9,30,31</sup> The extracts likely reduce ROS generation, inhibit  
340 cytokine-mediated injury, and promote diuresis, which aids in flushing out toxic metabolites. From a  
341 Unani perspective, kidney diseases are associated with “Sue MizajBaridRatabMaddi” (cold and moist  
342 derangement of temperament), causing morbid humour accumulation and obstruction in renal  
343 pathways. Biskhapra, with its “Hot and Dry” temperament,<sup>7</sup> restores normal mizaj, improves renal  
344 perfusion, and enhances urine flow. Its *Mohallil* (resolvent) property dissolves morbid matter, while  
345 *Qabiz* (astringent) action strengthens renal structures, reducing protein loss and edema.<sup>30</sup>

346 Histopathological observations supported the biochemical findings, showing regeneration of tubular  
347 epithelium, restoration of glomerular structure, and reduced inflammatory infiltration in treated groups  
348 compared to the negative control. Between the two extracts, the aqueous extract demonstrated superior  
349 nephroprotective activity, likely due to optimal extraction of water-soluble antioxidants and other  
350 active compounds. These findings emphasize the importance of extraction methods in preserving  
351 phytochemical integrity and therapeutic potency.

352 The study’s outcomes reinforce traditional Unani wisdom regarding the use of organ tonics  
353 (*MuqavviAaza*) in maintaining renal health.<sup>8</sup> Biskhapra not only protects against nephrotoxic insult but  
354 also promotes functional recovery, bridging classical Unani principles with modern pharmacological  
355 validation. Moreover, the nephroprotective and curative properties of *Biskhapra* leaf extract have been  
356 established in an Adriamycin-induced nephrotic syndrome rat model, prompting further investigation  
357 into its efficacy in other models of renal injury.<sup>32</sup> However, further research is required to isolate and  
358 characterize the active compounds, determine molecular mechanisms, and conduct clinical trials to  
359 confirm efficacy and safety in humans. In conclusion, Biskhapra exhibits promising nephroprotective  
360 potential against cisplatin-induced AKI through antioxidant, anti-inflammatory, and restorative  
361 mechanisms, supporting its therapeutic relevance in both traditional and modern medicine.

## 362 **Conclusion**

363 The present study validates the nephroprotective potential of *Trianthemaportulacastrum* Linn.  
364 (Biskhapra) leaves in a cisplatin-induced acute kidney injury model. Both aqueous and hydro-alcoholic  
365 extracts significantly mitigated cisplatin-induced elevations in serum creatinine, urea, and BUN, while  
366 restoring serum protein levels and preserving renal architecture. Histopathological findings supported  
367 these biochemical results, showing reduced tubular necrosis, regeneration of renal tubules, and  
368 improved glomerular integrity in treated groups.

369 Among the extracts, the aqueous preparation demonstrated superior nephroprotective activity,  
370 suggesting better retention of active phytoconstituents. The phytochemical profile of Biskhapra  
371 revealed the presence of alkaloids, flavonoids, saponins, tannins, steroids, and phenolics, which  
372 collectively contribute to its antioxidant, anti-inflammatory, and diuretic actions. From the perspective  
373 of Unani medicine, Biskhapra acts as MuqavviGurdawaMasana, restoring renal temperament,  
374 enhancing perfusion, and promoting elimination of morbid humours.

375 These findings provide compelling evidence for the traditional use of Biskhapra in kidney disorders  
376 and encourage further clinical and mechanistic studies to establish its role as a nephroprotective  
377 therapeutic agent in modern integrative medicine.

## 378 **Acknowledgement**

379 I would like to sincerely thank to Prof. MahfoozurRahman sir, Professor in the Department of  
380 IlmuAdvia at the Govt. Tibbi College & Hospital, Patna, for his valuable guidance and continuous  
381 support during the writing of this article. His helpful suggestions, encouragement, and feedback greatly  
382 improved the quality of this work.

## 383 **Author's Contribution**

384 Not available

385 **Conflict of Interest**

386 Not available

387 **Financial Support**

388 Not available

389 **References**

- 390 1. Guyton AC. Textbook of Medical Physiology. 8th Edn. Singapore: Saunders company,  
391 Hartcourt publishers International company; 2000. 264–379 p.
- 392 2. Winther-Jensen M, Kjaergaard J, Lassen JF, Køber L, Torp-Pedersen C, Hansen SM, et al. Use  
393 of renal replacementtherapy afterout-of-hospital cardiac arrest in Denmark 2005–2013.  
394 ScandCardiovasc J. 2018 Sep 3;52(5):238–43.
- 395 3. Harrison’s Manual of Medicine: 16th Edition [Internet]. [cited 2025 Jun 14]. Available from:  
396 [https://www.mheducation.com/highered/  
397 edition.html](https://www.mheducation.com/highered/mhp/product/harrison-s-manual-medicine-16th-edition.html)
- 398 4. ParkS, LeeS, LeeA, PaekJH, ChinHJ, NaKY, et al. Awareness, incidenceand clinical significance of  
399 acute kidney injury after non-general anesthesia: A retrospective cohort study. Medicine  
400 (Baltimore). 2018;97(35):e12014.
- 401 5. Kirkley MJ, Boohaker L, Griffin R, Soranno DE, Gien J, Askenazi D, et al. Acute kidney  
402 injury in neonatal encephalopathy: an evaluation of the AWAKEN database. PediatrNephrol.  
403 2019 Jan;34(1):169–76.

- 404 6. Barley M Brenner, Floyd C Rector. Barley M Brenner; Floyd C Rector; 2000;” The Kidney  
405 “6th EdnVol 1st; WB Saunders Company Philadelphia; pp no 3-  
406 67.YahooIndiaSearchResults[Internet].2000[cited2025Jun16].
- 407 7. Sina I. Alqanoon Fit Tibb (Urdu Translation by Hakeem Ghulam Husain Kantoori). Vol. 4.  
408 New Delhi, India: Eijaz Publishing House; 2010. 1205, 1206p.
- 409 8. HkmWaseem Ahmed Azmi. KuliyaAdvia. Ajaz publishing house; 1997. 138p.
- 410 9. Ghani N. KhazainulAdvia. Ed. 1st. New Delhi: IdaraKitabusShifa; YNM. 231, 371, 410, 661,  
411 665, 667, 676, 1005, 1016, 1033, 1053 p.
- 412 10. Sugiyama S, Hayakawa M, Kato T, Hanaki Y, Shimizu K, Ozawa T. Adverse effects of anti-  
413 tumor drug, cisplatin,on rat kidney mitochondria: Disturbances in glutathione peroxidase  
414 activity. BiochemBiophys Res Commun. 1989 Mar 31;159(3):1121–7.
- 415 11. Singh RB, Jindal VK. Water soluble polysaccharides from Cassia auriculata seeds. J Econ Bot  
416 Phytochem. 1990;1:25–8.
- 417 12. Yu HI. Effect of tripterygiumwilfordii with radix salviaemiltiorrhizae in purpuric nephritis.  
418 Chin J IntegdTradlWestn Med. 1992;12(6):343–4.
- 419 13. Asuzu I.U, Abubaker I. The antineoplastic effects of an extract from Leacinatricantha” .Vol 3  
420 (4) pp no 9-20. - Yahoo India Search Results [Internet]. 1995 [cited 2025 Jun 16].
- 421 14. Mustea I, Postescu ID, Tamas M, Rasnita TD. Experimental evaluation of protective activity  
422 ofEchinaceapallida against cisplatin toxicity. Phytother Res. 1997 May;11(3):263–5.
- 423 15. Sunanda Panda SP, Pratima Gupta PG, AnandKar AK. Protective role of ashwagandha in  
424 cadmium-induced hepatotoxicity and nephrotoxicity in male mouse. 1997 [cited 2025 Jun 16];  
425 Available from: <https://www.cabidigitallibrary.org/doi/full/10.5555/19980304075>

- 426 16. Shamim Anwar SA, Khan NA, Amin KMY, Ghufraan Ahmad GA. Effects of BanadequlBuzoor  
427 in some renal disorders. 1999 [cited 2025 Jun 14]; Available from:  
428 <https://www.cabidigitallibrary.org/doi/full/10.5555/20000314091>
- 429 17. Samiulla DD. Comparitive evaluation of polyherbal formulation for its nephroprotective  
430 activity. In: Proceedings of International Congress on Ayurveda. 2000. p. 193.
- 431 18. MohdAfzal. Nephroprotective effects and standardization of Unani compound formulation –  
432 JawarishZaroonisada. AMU Aligarh: Dept of IImulAdvia; 2000.
- 433 19. MohdAzam Khan. Moheet e Azam. Vols. 4, Part-1. Kanpur: MatbaNizami; 1313. 4–7 p.
- 434 20. Lim BO, Yu BP, Oh JH, Park DK. The inhibitory effects of ginsenoside and quercetin on  
435 oxidative damage by puromycinaminonucleoside in rat. *Phytother Res.* 1998 Aug;12(5):375–7.
- 436 21. Yuka N, Michinori K, Takuo O. Effect of Geraniin on AminonucleosideNephrosis in Rats. *Nat*  
437 *Med.* 1999 Apr;53(2):94–100.
- 438 22. Kirtikar, K.R,Basu,B.D.Indianmedicinalplantswithillustrations.2nded.Vol.  
439 5.Dehradun:OrientalEnterprises;2003.1640p.
- 440 23. MominHM.TohfatulMomineem.Ed.1st.Lucknow:MatbaHasni;YNM.98p.
- 441 24. Karnick CR. Some useful crude-drug plants of the Hindu system of medicine. *ActaPhytother.*  
442 1970;17(10):181–4.
- 443 25. Nawaz HR, Malik A, Ali MS. Trianthenol: an antifungal tetraterpenoid from  
444 *Trianthemaportulacastrum*(Aizoaceae). *Phytochemistry.* 2001 Jan 1;56(1):99– 102.
- 445 26. Irfan S, Ranjha MMAN, Nadeem M, Safdar MN, Jabbar S, Mahmood S, et al. Antioxidant  
446 Activity and Phenolic Content of Sonication- and Maceration- Assisted Ethanol and Acetone  
447 Extracts of *Cymbopogon citratus* Leaves. *Separations.* 2022 Sep;9(9):244.

- 448 27. Perše M, Večerić-Haler Ž. Cisplatin-Induced Rodent Model of Kidney Injury: Characteristics  
449 and Challenges. *BioMed Res Int*. 2018;2018:1462802.
- 450 28. Yao X, Panichpisal K, Kurtzman N, Nugent K. *Cisplatin nephrotoxicity: a review*. *Am J Med*  
451 *Sci*. 2007;334(2):115–24.
- 452 29. Bhandary SK, N SK, Bhat VS, P SK, Bekal MP. PRELIMINARY PHYTOCHEMICAL  
453 SCREENING OF VARIOUS EXTRACTS OF PUNICA GRANATUM PEEL, WHOLE  
454 FRUIT AND SEEDS. *J Health Allied Sci NU*. 2020 Apr 29;02:34–8.
- 455 30. Kumar G, Banu GS, Pandian MR. Evaluation of the antioxidant activity of  
456 *Trianthemaportulacastrum L*. *Indian J Pharmacol*. 2005;37(5):331–3.
- 457 31. IbnHubl. *KitabulMukhtaraatFilTibb* (Urdu translation). Ed. 1st. Vol. vol 1st, vol 2nd. New  
458 Delhi: CCRUM, Ministry of Health & Family Welfare, Gov. of India; 2005. 70, 99–100, 141  
459 p.
- 460 32. Karim MS, Ashraf N, Kalam A, Jahan N, Jafri MA, Ahmad G. Effects of Biskhapra  
461 (*Trianthemaportulacastrum Linn.*) leaves extract in adriamycin induced nephrotic syndrome.  
462 *Int J Green Pharm IJGP* [Internet]. 2011 [cited2025 Jun 14]; 5(4). Availablefrom:  
463 <http://www.greenpharmacy.info/index.php/ijgp/article/view/223>

464

465

466

467

468



469

470

471

472

473

474

475

476

477

UNDER PEER REVIEW IN IJAR