

# STUDY OF VIRULENCE FACTORS AND ANTIMICROBIAL SUSCEPTIBILITY PATTERN OF UROPATHOGENIC ESCHERICHIA COLI IN A TERTIARY CARE HOSPITAL

## ABSTRACT

## INTRODUCTION

Escherichia coli is the major causative agent of urinary tract infections. UPEC possess a variety of virulence factors that enable it to colonize, invade and persist within the urinary tract, leading to both complicated and uncomplicated UTIs. Knowledge of virulence factors of Escherichia coli is responsible for pathogenesis of UTIs and their antimicrobial susceptibility pattern will help in better understanding of the treatment of UTI. Emergence and spread of multi drug resistant strains of Escherichia coli have raised considerable interest in understanding their diversity and epidemiology of infections in humans.

## AIM

To determine the virulence factors and antimicrobial susceptibility pattern of Uro Pathogenic Escherichia coli.

## MATERIALS AND METHODS

A prospective study is done on urine samples over a period of 3 months from May 2025 to July 2025 in a tertiary care hospital to detect virulence factors like Haemolysis, Hemagglutination and gelatin hydrolysis.

Antibiotic susceptibility test was done by Kirby Baur disc diffusion method as per the CLSI guidelines.

## RESULTS

Out of total 350 urine samples tested 114 were Escherichia coli isolates. Out of the 114 Escherichia coli isolates 69 were females and 45 males. Among the 114 isolates -43 isolates showed haemolysis virulence factor, 31 isolates showed hemagglutination factor- 22 were mannose resistant and 9 mannose sensitive hemagglutination and, 11 isolates showed gelatin hydrolysis and 29 isolates did not show any of the virulence factor.

Antibiotic susceptibility testing revealed that UPEC strains showed maximum resistance to **Ampicillin (55.2%)**, followed by **Cotrimoxazole (48.2%)** and **Norfloxacin (44.7%)**. Most isolates were sensitive to **Meropenem (82.4%)**, Amikacin (79.8%) and followed by **Nitrofurantoin (74.5%)**.

## CONCLUSION

Detection of virulence factors of Uro-pathogenic Escherichia coli shows a strong association to urinary tract infection. And presence of multiple virulence factors leads to drug resistance.

## KEY WORDS

Urinary tract infection, Uropathogenic Escherichia coli, Virulence factors, Haemolysis, Hemagglutination, Gelatin hydrolysis

## **INTRODUCTION**

Urinary tract infections (UTIs) are one of the most common bacterial infections affecting humans throughout their life span. They can be symptomatic or asymptomatic. *Escherichia coli* is the most common cause of UTIs, accounting for about 85% of community acquired and 50% of hospital-acquired infections, it predominates strongly at most ages. UPEC is the main cause of community-acquired UTIs (about 80-90%).

*Escherichia coli* is a commensal in the human intestinal tract, when enters into unnatural sites, it can cause a variety of infections, e.g., UTIs, sepsis, pyelonephritis etc. Serotypes which lead to UTIs are designated as uropathogenic *Escherichia coli* (UPEC)<sup>1</sup>. It has been known that certain serotypes of *E. coli* are consistently associated with uro-pathogenicity and are designated as uropathogenic *E. coli* that expresses chromosomally encoded virulence markers<sup>3</sup>. Uropathogenic strains account for 90% of all UTIs among ambulatory patients and upto 50% of all nosocomial UTIs<sup>2</sup>.

The common virulence factors include surface hydrophobicity, colonization factor, capsule, serum resistance, resistance to phagocytosis, hemolysin production, enterotoxin and siderophore, fimbriae and hemagglutination. The ability of *E. coli* to adhere to the uroepithelium is mediated by fimbriae, thereby resisting elimination by the flow of urine. Adhesion is therefore measured to be important step in the pathogenesis of UTI<sup>4</sup>.

During UTIs, UPEC pathogenesis includes:

- (a) UPEC colonization of the periurethral and vaginal areas with colonization of the urethra;
- (b) ascending into the bladder lumen and growth as planktonic cells in urine;
- (c) adherence to the surface and interaction with the bladder epithelium defense system (see below);
- (d) biofilm formation;
- (e) invasion and replication by forming bladder Intracellular Bacterial Communities (IBCs) where quiescent intracellular reservoirs (QIRs) form and reside in the underlying urothelium;
- (f) kidney colonization and host tissue damage with increased risk for bacteremia/septicemia.

Considering the high degree of morbidity and mortality due to UTIs caused by uropathogenic *E. coli* and also the drug resistance among strains has further aggravated the problem of UTI's. Therefore, the present study was carried out with aim to know the prevalence of various virulence factors and the antimicrobial susceptibility pattern in UPEC<sup>5</sup>.

## **MATERIALS AND METHODS**

This is a prospective study which was conducted in the Department of Microbiology in a tertiary care hospital over a period of 3 months i.e from May 2025-July 2025. Patients of all age group were included. A total of 350 urine samples were tested out of which 114 were *Escherichia coli* isolates. The samples were processed immediately as per the standard guidelines in the lab. The isolates were taken for the detection of virulence factors.

The virulence factors tested were

1. Haemolysin production:
2. Haemagglutination
3. Gelatin hydrolysis

### **1. Haemolysin production**

The *Escherichia coli* isolates were inoculated onto 5 % sheep blood agar and incubated overnight at 37 degree Celsius and observed for a zone of complete lysis around the colony. *Escherichia coli* ATCC 25922 was used as a negative control.

Presence of clear zone of complete hemolysis indicates hemolysin production.

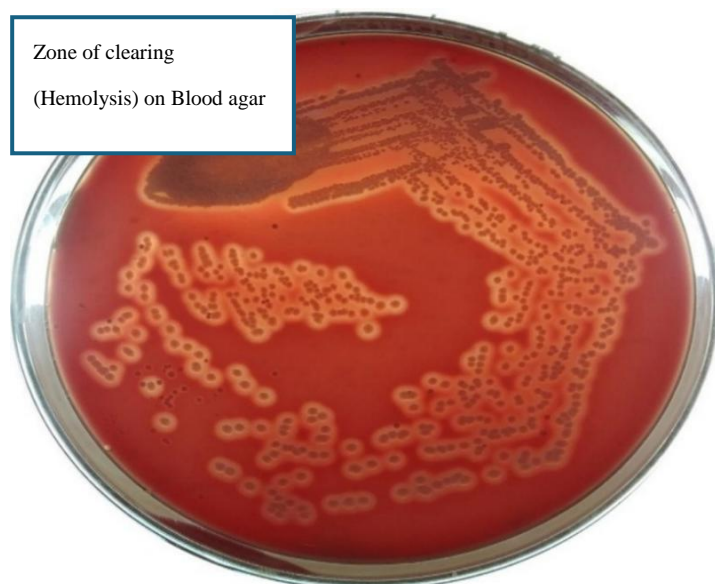


FIG 1: HEMOLYSIS

## 2.HAEMAGGLUTINATION

The test was carried out as per the direct bacterial hemagglutination test-slide method. One drop of red blood cell (RBC) suspension was added to a drop of broth culture and the slide was rocked at room temperature for 5 min. Presence of clumping was taken as positive for hemagglutination. Mannose-sensitive hemagglutination was detected by the absence of hemagglutination in a parallel set of test in which a drop of 2% W/V D-mannose was added to the red cells and a drop of broth culture. Mannose resistant hemagglutinating (MRHA) was detected by the presence of hemagglutination of 3% 'O' blood group human RBCs in the presence of 2% W/V D-mannose.

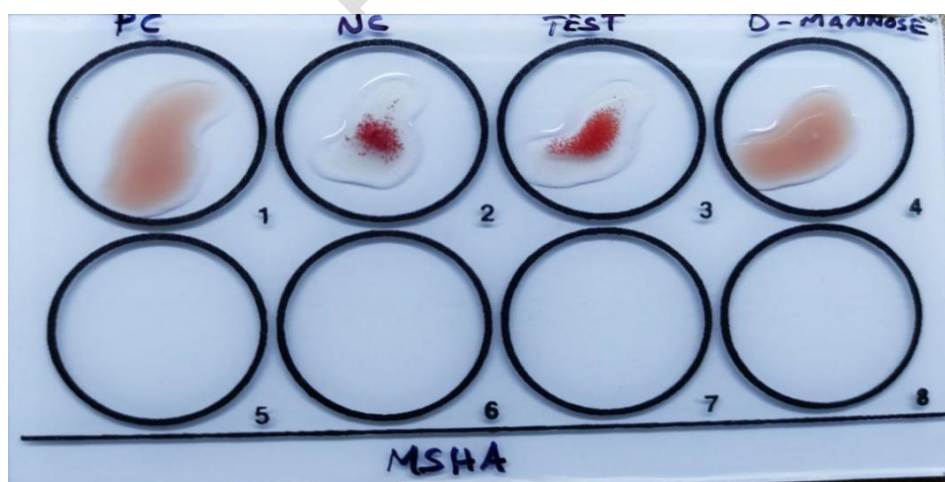


FIGURE 2: MANNOSE SENSITIVE HAEMAGGLUTINATION

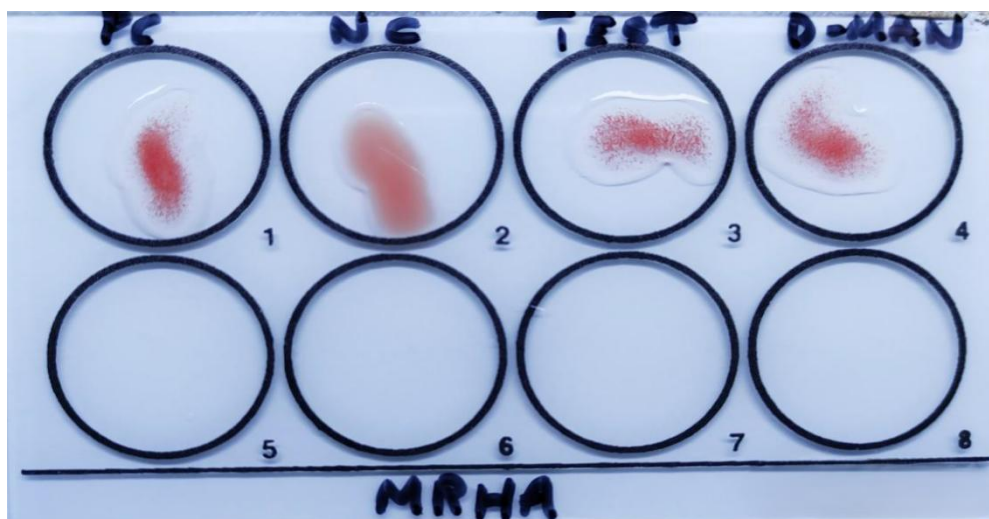


FIGURE 3: MANNOSE RESISTANT HAEMAGGLUTINATION

### 3. GELATIN HYDROLYSIS.

Gelatinase production was tested using gelatin agar. *Escherichia coli* isolated was inoculated on gelatin agar and incubated overnight at 37 degrees celsius for 24 hrs. Later the gelatin agar plate was flooded with 1% tannic acid. Development of opacity around colonies shows gelatinase production.

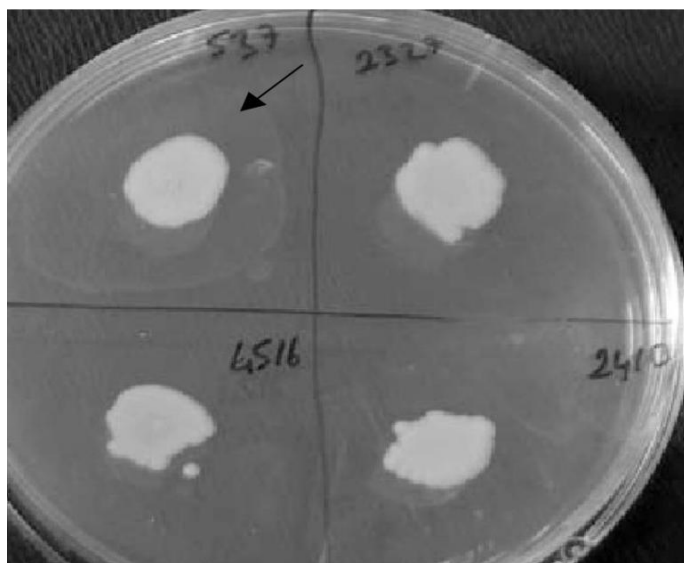


FIGURE 4: GELATIN HYDROLYSIS

### 4. Antimicrobial susceptibility

Antimicrobial susceptibility testing was done on Mueller Hinton Agar by Kirby Bauer disc diffusion method as per CLSI guidelines.

## **RESULTS**

A total of 350 urine samples received from symptomatic cases of urinary tract infection with significant bacteriuria were taken. Out of these, 114 *Escherichia coli* isolates were processed and studied for the virulence factors and their antibiotic susceptibility pattern. Out of the 114 *Escherichia coli* isolated samples, 60.5% were from females and 39.47% were from males. Among 114 isolates, 37.7% showed haemolysis, 19.3% showed mannose resistant haemagglutination, 7.9% showed mannose sensitive haemagglutination, 9.7% showed gelatin hydrolysis and 25.4% did not show any of the virulence factors.

Antibiotic susceptibility testing revealed that UPEC strains showed maximum resistance to Ampicillin (55.2%), followed by Cotrimoxazole (48.2%) and Norfloxacin (44.7%). Most isolates were sensitive to Meropenem (82.4%), Amikacin (79.8%) and followed by Nitrofurantoin (74.5%).

TABLE 1: GENDER WISE DISTRIBUTION OF ESCHERICHIA COLI ISOLATES

GENDER	TOTAL	PERCENTAGE
FEMALES	69	60.5%
MALES	45	39.4%
TOTAL	114	100%

TABLE 2: VIRULENCE FACTORS AMONG THE UROPATHOGENIC ESCHERICHIA COLI ISOLATES(n=114)

VIRULENCE FACTOR	NUMBER	PERCENTAGE
HAEMOLYSIS	43	37.7%
HEMAGGLUTINATION	(31)	
MANNOSE RESISTANT HEMAGGLUTINATION	22	19.3%
MANNOSE SENSITIVE HEMAGGLUTINATION	9	7.9%
GELATIN HYDROLYSIS	11	9.7%

TABLE 3: ANTIBIOTIC SUSCEPTIBILITY PATTERN OF UPEC ISOLATES

ANTIBIOTIC	SENSITIVE	PERCENTAGE	RESISTANT	PERCENTAGE
AMPICILLIN	51	44.8%	63	55.2%
TRIMETHOPRIM SULFAMETHOXAZOLE	59	51.8%	55	48.2%
NORFLOXACIN	63	55.3%	51	44.7%
CEFTRIAXONE	67	58.8%	47	41.2%
MEROPENEM	94	82.4%	20	17.6%
AMIKACIN	91	79.8%	23	20.2%
NITROFURANTOIN	85	74.5%	29	25.5%

## DISCUSSION

UPEC are the most important group of micro-organisms responsible for UTI. UPEC differ from non-pathogenic *E. coli* by the production of specific virulence factors which enable the bacteria to adhere to uroepithelial cells and to establish UTI<sup>5</sup>. UTIs which are not properly treated from their onset can become a renal threat in time, finally leading to renal failure<sup>8</sup>.

These virulence factors enable some members of the normal flora to elicit an infection by overcoming the host defence mechanisms<sup>1</sup>.

The capacity of *E. coli* to produce many virulence factors contributes to its pathogenicity<sup>1,7</sup>.

Incidence of UTI was more common in females 60.5% than in males in our study. Piatti et al<sup>5,6</sup> also reported a higher prevalence of UTI in female (77%). The reasons for the high prevalence of the UTIs in females can be due to the anatomical structure of the urogenital tract having short urethra, presence of normal flora in vagina, menstrual cycle and pregnancy<sup>5</sup>.

Priscilla et al<sup>1</sup> also reported a higher incidence of UTI was more common in females (66.72%) than in males (33.27%) in our study.

Our study also correlates with Sanjay Singh Kaira et al<sup>4</sup> who reported 56.09%, Mittal et al<sup>1,8</sup> (53.3%) and Chhaya et al<sup>8,9</sup> (53%).

Hemolysin production is associated with human pathogenic strains of *E. coli*, especially those causing more clinically severe forms of UTI<sup>5,10</sup>. It is toxic to a range of host cells in ways that probably contribute to inflammation, tissue injury and impaired host defenses<sup>1,11</sup>. In the present study, 37.7% *E. coli* isolates produced hemolysin. In other studies conducted by Raksha et al<sup>5,12</sup>, Siegfried et al<sup>5,13</sup>, Hughes et al<sup>5,14</sup>, Shruthi et al<sup>1,15</sup> hemolysin production was detected in 41.36% and 59.6%, 59.7% and 41.9% isolates respectively.

The role of bacterial adherence in the pathogenesis of urinary tract infection is that colonization of the urogenital epithelium of susceptible individuals by specific bacteria is associated with successful microbial invasion of the urinary tract<sup>16</sup> and lead to UTIs. Thus, possession of MRHA by UPEC can be considered as one of the important virulence factor in the pathogenesis of UTIs. My study correlates with Seigfried et al<sup>1,13</sup>, Vagarali et al<sup>13</sup>, Raksha et al<sup>1,12</sup>, Kauser et al<sup>3,16</sup> have reported the incidence of MRHA *E. coli* isolates as 23%, 25%, 30.9%, 30% respectively. In the present study also the rate of MRHA positive *E. coli* isolates was 19.3 %

Gelatinase, an important virulence factor which is capable of hydrolyzing gelatin, collagen, and is associated with inflammation. Shetty et al<sup>17,18</sup> observed that gelatinase is not an important virulence factor. While Mittal et al<sup>5</sup> observed that gelatinase producing strains were multidrug resistant. In the present study Gelatinase production was seen in 25.4 % of isolates which is similar to Mittal et al<sup>5</sup> where it was 67.5%. But Vaish et al<sup>18,19</sup> & Jayanthi et al<sup>18,20</sup> showed lesser production of gelatinase which was 2% & 6% respectively.

Antibiotic susceptibility pattern was studied for all *E. coli* isolates. These isolates were most commonly resistant to Ampicillin, Cotrimoxazole and Norfloxacin. And the maximum sensitivity was shown to Meropenem, Amikacin and then followed by Nitrofurantoin. The present study has shown the production of various virulent factors and developing drug resistance in UPEC.

The antibiotic susceptibility pattern observed in our study correlates with Tabasi et al<sup>8,21</sup>, Karam et al<sup>8,22</sup> and Chhaya et al<sup>8,9</sup>.

The present study also correlates with Kauser Y et al<sup>16,18</sup>, Vaish et al<sup>18,19</sup> & Jayanthi et al<sup>18,20</sup>.

Antibiotic resistance may provide a substantial advantage to the survival of the pathogen. The drug resistance among UPEC is on rise therefore the selection of appropriate antibiotics after antibiotic susceptibility testing is must for proper treatment of patients and to avoid emergence of drug resistance.

Resistance to commonly used antibiotics is because of excessive use and misuse of the antibiotics by the healthcare personnel and dissemination of multidrug resistance among hospital strains<sup>18,23,24</sup>.

## CONCLUSION

Detection of virulence factors of Uro-pathogenic *Escherichia coli* shows a strong association to urinary tract infection. And presence of multiple virulence factors leads to drug resistance. So this helps in better understanding and treatment of Urinary tract infection. Since most Urovirulent strains express multiple virulent factors simultaneously, further studies at molecular level are necessary.

## REFERENCES

1. Priscilla, P., Tiwari, A., & Kumari, P. (2025). Study of virulence factors of uropathogenic *Escherichia coli* and its antibiotic susceptibility pattern. *International Journal of Health Sciences*, 9(S1), 636-642. <https://doi.org/10.53730/ijhs.v9nS1.15820>
2. Steadman R, Topley N (1998) The virulence of *Escherichia coli* in urinary tract, In: Urinary tract infections. Chapman and Hall publication, London.
3. Vagarali MA, Karadesai SG, Patil CS, Metgud SC, Mutnal MB. Haemagglutination and Siderophore production as the urovirulence markers of Uropathogenic *Escherichia coli*. *Indian J Med Microbiol*. 2008;26(1):68-70
4. Kaira SS, Pai C. Study of uropathogenic *Escherichia coli* with special reference to its virulence factors. *Int J Community Med Public Health* 2018;5:177-81.
5. Mittal S, Sharma M, Chaudhary U. Study of virulence factors of uropathogenic *Escherichia coli* and its antibiotic susceptibility pattern. *Indian J Pathol Microbiol* 2014;57:61-4.
6. Piatti G, Mannini A, Balistreri M, Schito AM. Virulence factors in urinary *Escherichia coli* strains: Phylogenetic background and quinolone and fluoroquinolone resistance. *J Clin Microbiol* 2008;46:480-
7. Biswas D, Gupta P, Prasad R, Singh V, Arya M, Kumar A. Choice of antibiotics for empirical therapy of acute cystitis in a setting of high antimicrobial resistance. *Indian J Med Sci*. 2006;60(2):53-8.
8. K. Pavani, Ramalakshmi. K, T. Kanakadurgamba et.al. Study of virulence factors and antimicrobial susceptibility pattern of uropathogenic *Escherichia coli* in a tertiary care hospital. *International Journal of Research and Review*. 2021; 8(2): 591-596.
9. Shah Chhaya, Baral R, Bartaula B, Shrestha LB. Virulence factors of uropathogenic *Escherichia coli* (UPEC) and correlation with antimicrobial resistance. *BMC Microbiol*. 2019 Sep 2;19(1):204. doi: 10.1186/s12866-019-1587-3.
10. Slavchev G, Pisareva E, Markova N. Virulence of uropathogenic *Escherichia coli*. *J Cult Collect* 2008-2009;62:3-9.
11. Stanley P, Koronakis V, Hughes C. Acylation of *Escherichia coli* hemolysin: A unique protein lipidation mechanism underlying toxin function. *Microbiol Mol Biol Rev* 1998;62:309-33.

12. Raksha R, Srinivasa H, Macaden RS. Occurrence and characterisation of uropathogenic *Escherichia coli* in urinary tract infections. *Indian J Med Microbiol* 2003;21:102-7.
13. Siegfried L, Kmetová M, Janigová V, Sasinka M, Takácová V. Serum response of *Escherichia coli* strains causing dyspepsia and urinary tract infection: Relation to alpha-hemolysin production and O type. *Infect Immun* 1995;63:4543-5.
14. Hughes C, Phillips R, Roberts AP. Serum resistance among *Escherichia coli* strains causing urinary tract infection in relation to O type and the carriage of hemolysin, colicin, and antibiotic resistance determinants. *Infect Immun* 1982;35:270-5
15. Shruthi N, Kumar R, Kumar R. Phenotypic study of virulence factors in *Escherichia coli* isolated from antenatal cases, catheterized patients, and faecal flora. *J Clin Diagn Res* 2012;6:1699-703.
16. Kauser Y, Chunchanur SK, Nadagir SD, Halesh LH, Chandrashekhar MR. Virulence factors, serotypes and antimicrobial susceptibility pattern of *Escherichia coli* in urinary tract infections. *Al Ameen J Med Sci* 2009; 2:47-1.
17. Shetty SK, Rao SP, Subbannayya K, Janakiram K. Study of prevalence of virulence factors in extraintestinal pathogenic *Escherichia coli* isolated from a tertiary care hospital. *Int J Curr Microbiol App Sci*. 2014;3(7):1055-61.
18. Hiremath MB, Lava R. Study of virulence factors and antibiotic susceptibility pattern of extraintestinal pathogenic *Escherichia coli*. *Indian J Microbiol Res* 2020;7(4):330-334.
19. Kandi V, Vaish R, Pradeep MSS, Setty CR. Evaluation of Virulence Factors and Antibiotic Sensitivity Pattern of *Escherichia Coli* Isolated from Extraintestinal Infections. *Cureus*. 2016;8(5):604. doi:10.7759/cureus.604.
20. Jayanthi RS, Soumya K. Study of Virulence Factors in *Escherichia coli* Isolated from Skin and Soft Tissue Infections. *Int J Curr Microbiol Appl Sci*. 2017;6(7):2288-94. doi:10.20546/ijcmas.2017.607.269.
21. Tabasi M, Asadi Karam MR, Habibi M, Yekaninejad MS, Bouzari S. Phenotypic Assays to Determine Virulence Factors of Uropathogenic *Escherichia coli* (UPEC) Isolates and their Correlation with Antibiotic Resistance Pattern. *Osong Public Health Res Perspect*. 2015;6(4):261-268. doi:10.1016/j.phrp.2015.08.002



- 22.Karam MRA, Habibi M, Bouzari S. Relationships between Virulence Factors and Antimicrobial Resistance among Escherichia coli Isolated from Urinary Tract Infections and Commensal Isolates in Tehran, Iran. Osong Public Health Res Perspect. 2018 Oct;9(5):217-224. doi: 10.24171/j.phrp.2018.9.5.02.
- 23.Chitnis SV, Chitris V, Sharma N, Chitnis DS. Current status of drug resistance among gram negative bacilli isolated from admitted cases in a tertiary care centre. J Assoc Physicians India. 2003;51:28-31.
24. Wiener J, Quinn JP, Bradford PA, Goering RV, Nathan C, Bush K. Multiple antibiotic resistant Klebsiella and Escherichia coli in a nursing home. JAMA. 1999;281:517-23.