

## 1 Hepatoprotective Effect of *Kalmegha* (*Andrographis paniculata*): Insights 2 from an Animal Experimental Study

3

## 4 Abstract-

**Background:** Liver diseases are a global health burden. *Kalmegha* (*Andrographis paniculata*), a traditional medicinal plant, has been used for liver ailments, but scientific validation in experimental models is essential. **Objective:** To evaluate the hepatoprotective potential of *Kalmegha* in experimentally induced hepatic injury in animal models. **Methods:** [Briefly mention model used, e.g., carbon tetrachloride/paracetamol-induced hepatotoxicity, animal group division, dose of extract, duration, biochemical & histopathological assessment. **Results:** *Kalmegha* significantly improved biochemical parameters (ALT, AST, Bilirubin) and restored antioxidant levels compared to toxicant control. Histopathological examination supported biochemical findings. **Conclusion:** Findings suggest *Kalmegha* possesses promising hepatoprotective activity and may serve as a potential natural hepatoprotective agent.

15 **Key Words-***Kalmegha, Andrographis paniculata*, hepatoprotective, liver injury, animal study,  
16 herbal medicine.

17

18 INTRODUCTION

19 Liver diseases are a major cause of morbidity and mortality worldwide, accounting for  
20 nearly two million deaths annually <sup>1</sup>. Current pharmacological therapies are limited and  
21 often associated with side effects, which has increased interest in natural hepatoprotective  
22 agents<sup>2</sup>.

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Acharya Priyavrat Sharma states in *Priye Nighantu* that *Kalmegha* possesses Deepana and Pittasaraka properties and is used in *Yakrit Roga*, highlighting its hepatoprotective potential<sup>3</sup>

31 *Kalmegha* is cited for liver disorders in *Dravyaguna Hastamalak*<sup>4</sup>, *Vanoshadhi Nidarsika*<sup>5</sup>,  
32 and Vaidya V.M. Gagte's *Ayurvedic Pharmacology*<sup>6</sup>.  
33 .

34 *Kalmegha* (*Andrographis paniculata*, family: Acanthaceae), known as "King of Bitters,"  
35 is widely used in Ayurveda, Siddha, and traditional Chinese medicine for treating fever,  
36 jaundice, and liver disorders<sup>7</sup>. Its bioactive compound, andrographolide, has been  
37 reported to possess antioxidant, anti-inflammatory, and hepatoprotective properties<sup>8</sup>.  
38 However, experimental validation of its hepatoprotective efficacy in animal models  
39 remains crucial.

40 The present study was designed to evaluate the hepatoprotective activity of *Kalmegha*  
41 against paracetamol induced hepatotoxicity in Wistar rats.

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## 43 AIM AND OBJECTIVES

44

### Aim-

45 To study the effect of *Kalmegha* on hepatic disorders.

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### Objective-

47  To study about *Yakrit Roga* (liver disorder) and hepatotoxicity and determine the role of  
48 hepatoprotective property of *Kalmegha* in *Yakrit Roga*.

49  To serve humanity by providing safe, economical and effective hepatoprotective drug and by  
50 treating liver diseases without producing toxicity.

51  To prove that Ayurveda cures diseases in natural way without disturbing equilibrium of the  
52 body, and is superior therapy than other modern therapies.

53

## 54 MATERIALS AND METHODS

55

### 56 Plant Material

57 *Kalmegha* leaves were collected, shade-dried, and authenticated by a botanist. To  
58 investigate anti-hepatotoxic substances, it is customary to subject animal experimentation  
59 to a range of standard protocols of hepatoprotective activity by merging certain *in vivo*

60 and *in vitro* models. In these models certain toxic substances or toxicant have been used  
61 to produce hepatic injury resembling the different diseases and then anti hepatotoxic  
62 activity is evaluated.

63 The hepatoprotective activity is assessed by noting the effect of test drug on toxicants  
64 induced changes in different parameters like weight, volume and cytoarchitecture of  
65 liver, viability of hepatocytes after perfusion with test drug, chemical constituents and  
66 enzyme activity in liver and serum, especially those that are related to secretory  
67 metabolic and excretory functions of liver.

68 **Study design-**

69 **(a) Chemicals**

70 The chemicals used in hematology, biochemistry and histopathology are listed  
71 below.

- 72 • Acetone (Make: Molychem, 21040)
- 73 • Alanine amino transferase kit (Cat loguErba Transaia Bio-Medicals)
- 74 • Aspartate amino transferase kit (CatT, Erba ransaia Bio-Medicals )
- 75 • Eosin-Y (Merck, 230251)
- 76 • Geimsa stain (Fisher Scientific, 38723)
- 77 • Hematology reagents (Ark diagnostics)
- 78 • Harris hematoxylin (Hi-Media, SO34)
- 79 • Paraffin wax (Make: Hi-Media, GRM1042)

80 **(b) Equipment**

- 81 • Blood cell analyzer (Make: Abacus junior vet 5)
- 82 • Centrifuge machine (Make: REMI, R-8C)

83           ● Microscope with image capturing facility (Make: Leica)

84           ● Rotary microtome (Make: Yorco, Spencer type)

85           ● Semi-automatic biochemical analyzer (Make: Erba Mannheim)

86           ● Wax incubator (Make: Yorco scientific)

87 **Experimental animals**

88           The experiment was conducted on Wistar rats of either sex weighing 150-200 g.

89           These rats were housed in the Central Laboratory Animal House, College of Veterinary Science

90           & A.H., Jabalpur, as per the guidelines of CPCSEA. Rats were provided with *ad-libitum*

91           commercial pelleted feed (Nutrivet life sciences) and water. Environmental conditions such as

92            $22\pm3^{\circ}\text{C}$  temperature and 12 hours light and dark cycle were given to the rats. The protocol of the

93           study was approved by the IAEC (IAEC protocol no. 09./IAEC/Vety/20)

94           After 05 days of acclimatization, the rats were randomly divided into four groups,

95           consisting of 08 animals each. The treatment protocol is summarized in following Table .

96 **Experimental design**

<b>Group</b>	<b>Treatment</b>	<b>Number of animals</b>
I	Positive Control,	08
II	Negative Control	08
III	2 ml decoction of <i>Kalmegha</i>	08
IV	2 ml dilution of <i>Kalmegha</i>	08

97

98           ● Hepatic damage in rats from group I, III, IV, were produced by administration of single

99           dose of Paracetamol @1500mg/kg orally.

100          ● Rats of group I served as Positive control of hepatic damage.

101          ● Rats of group III, and IV, also administered with treatment protocol as mentioned in

102           above table.

103       ● Rats of group II were provided with standard feed and water, served as Negative control.

104   **Clinical observation**

105       ● All the rats belonging to various groups were closely observed on a daily basis for the  
106       development of any clinical signs during the entire experimental period.

107   **Body weight estimation**

108       ● The body weight of individual rats was recorded from the first to fourth week of the study  
109       to assess the weekly body weight gain using the weighing balance (Aczet, CY223C)  
110       during the experimental period.

111   **Collection of blood samples**

112       ● Approximately 1.0 ml of blood was collected aseptically from retro orbital sinus of rats  
113       on 30<sup>th</sup> day of study. Then, the blood was transferred into two sterilized Eppendorf tubes,  
114       one coated with heparin as anticoagulant was used for hematological examination, while  
115       the other without anticoagulant was used for serum separation (stored at -20°C).

116   **Serum Biochemistry**

117       ● Serum samples were analyzed for biochemical parameters namely, ALT, AST and total  
118       bilirubin using semi-automatic biochemical analyzer (Erba mannheim) by using  
119       commercially available kits (Erba- Transaia Bio-Medicals LTD).

120   **Collection of tissue samples**

121       ● Rats belonging to different groups were humanely sacrificed at end of study period. All  
122       the rats were subjected for detailed post mortem examination. Liver from different groups  
123       were collected. A portion of liver is collected for histopathological study.

124   **Gross Pathological Examination**

125       ● Rats belonging to various experimental groups were subjected to detail pathological  
126       examination. Gross pathological lesions were closely observed and recorded in various  
127       organs especially liver of rats.

128   **Histopathology**

129     ● Representative tissue samples of liver from rats of different groups were collected and  
130       fixed in 10% formalin for a minimum period of 24 hours and processed for  
131       histopathological examination. Tissue pieces of approximately  $0.5 \times 0.5$  cm in size were  
132       dehydrated in three changes of acetone and cleared in three changes of benzene, followed  
133       by impregnation was done in four chambers of wax. The paraffin tissue blocks were made  
134       by embedding tissue in wax using L-molds or cassettes as per the method described by  
135       Gridley (1960) with slight modification.

136     **Section cutting**

137     ● Approximately 4-5 mm thin sections were cut by rotary microtome and ribbon section  
138       was placed in water bath. Floating sections were taken on clean glass slides smeared with  
139       egg albumin as adhesive for histopathology.

140

141     **Staining of tissues**

142     ● **Hematoxylin & Eosin (H&E)**

143       Hematoxylin & Eosin staining was performed as per the method described by  
144       Gridley (1960) with slight modification. The sectioned slides were stained with  
145       hematoxylin and eosin, mounted with DPX (Distyrene plasticizer xylene) and covered  
146       with coverslips for further histopathological examination.

147       **Duration of study-** The study was conducted for the period of six months.

148     **RESULTS-**

149       In the present study, serum samples of rats were subjected to biochemical  
150       examination including, ALT, AST and Total bilirubin (TB). Results are presented in following  
151       table:

152     **Biochemical parameters of rats of different groups**

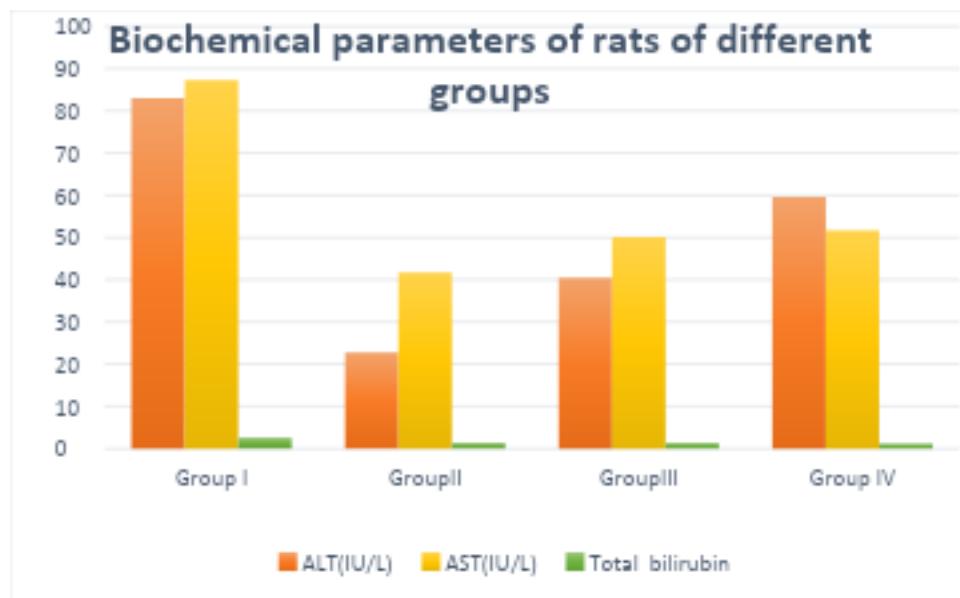
<b>Groups/ Parameters</b>	<b>ALT (IU/L)</b>	<b>AST (U/L)</b>	<b>Total Bilirubin(mg/dL)</b>
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<b>Group I</b>	82.97 <sup>a</sup> ±0.63	87.33. <sup>a</sup> ±1.07	02.60 <sup>a</sup> ±0..22
<b>Group II</b>	22.76 <sup>f</sup> ±0.32	41.66 <sup>d</sup> ±0.33	01.36 <sup>b</sup> ±0.05
<b>Group III</b>	40.46 <sup>d</sup> ±2.00	50.17 <sup>b</sup> ±1.61	01.35 <sup>b</sup> ±0.03
<b>Group IV</b>	59.64 <sup>b</sup> ±0.31	51.64. <sup>b</sup> ±0.28	01.21 <sup>b</sup> ±0.03

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157 Mean value with different superscript differs significantly( $P<0.05$ ) in row

158 Biochemical examination showed, significant increase in value of liver enzymes like  
 159 ALT, AST and total bilirubin rats of positive control as compared to negative control and other  
 160 groups. There was significant difference in the values between the groups.

161 There was significant improvement recorded on ALT and AST and total bilirubin  
 162 in all treated rats who were received decoction of *Kalmegha* (group III ) and dilution of  
 163 *Kalmegha* (group IV).

## 164 **DISCUSSION**

### 165 **Serum biochemistry**

166 Biochemical examination showed, significant increase in value of liver enzymes like ALT, AST  
167 and total bilirubin in rats of positive control as compared to negative control and other groups.

168 There was significant difference in the values between the groups. There was significant  
169 improvement recorded on level of ALT and AST and total bilirubin in all treated rats who  
170 received decoction of *Kalmegha* and dilution of *Kalmegha* · Though this improvement was more  
171 pronounced in group group III, then group V, with statistical difference at level of both the liver  
172 enzymes.

173 The above result aligns with Gupta et al. (2022)<sup>9</sup> and Thomas et al. (2023)<sup>10</sup> with improved level  
174 of ALT, AST and total bilirubin with use of *Kalmegha* due to its antioxidant, anti-inflammatory  
175 and hepatoprotective actions · However, Verma et al. (2013)<sup>11</sup> compared the hepatoprotective  
176 action of *Kalmegha* and *Chirayata*, in which extract of *A. Paniculata* showed significant better  
177 hepatoprotective as compare to *S. chirayita* . Shrivastava and Gilhotra. (2017) also evaluated the  
178 hepatoprotective activity in Kalmegha in CCl4 treated rats induced liver damage and found  
179 significant reduction in ALT value<sup>12</sup> .

180

## 181 **Gross and HistoPathology**

182 At the end of study, post mortem examination of rats pointed that dark coloured congestion and  
183 whitish-yellowish discolored foci in few rats of positive control group.

184 Histopathological examination of liver of rats of positive control, who received single dose of  
185 Paracetamol showed moderate necrosis and vacuolation of hepatocytes pointing towards hepatic  
186 damage, as also evident by the increased liver enzymes (ALT and AST ) in group I rats. Rats  
187 who received the decoction of *Kalmegha* after hepatic damage (paracetamol administration)  
188 showed normal histology and hepatocytes pointing towards improvement in liver  
189 histoarchitecture on administration of the drugs. Where as this improvement on liver histology  
190 was milder in rats who received dilution of *Kalmegha* after hepatic damage.

191 The findings of our research work aligns with work done Verma et al. (2013), paracetamol  
192 group shows severe centrilobular necrosis characterized by nuclear pyknosis, karyolysis, and  
193 eosinophilic infiltration, confirming extensive hepatocellular damage and found hepatoprotective

194 effects demonstrated by both the plants ( *Kalmegha* and *Chirayata* ) . Similarly, in present study  
195 we observed improvement on hepatic damage on administration of *Kalmegha* with better  
196 preserved histology in decoction of *Kalmegha* than dilution of *Kalmegha* . As *Kalmegha*  
197 prevents the lipid peroxidation of hepatocytes additionally anti-inflammatory effects protects the  
198 hepatocytes from necrosis and vacuolation.

199 **Overall Effect Of Therapy**

200 The study demonstrated that paracetamol administration at 1500 mg/kg produced significant  
201 biochemical disturbances, histopathological alterations, and pronounced hepatic injury in rats.  
202 *Kalmegha* exhibited clear hepatoprotective effects against this induced liver damage. Notably,  
203 the decoction form provided greater therapeutic benefit compared to dilution, indicating superior  
204 efficacy in restoring liver function and mitigating tissue injury.

205 The results demonstrate that *Kalmegha* possesses potent hepatoprotective activity against  
206 paracetamol induced hepatotoxicity. Restoration of liver function markers and antioxidant  
207 enzymes suggests its protective mechanism involves free radical scavenging and stabilization of  
208 hepatocyte membranes.

209 Andrographolide, the major active compound of *Kalmegha*, has been previously shown to  
210 modulate cytochrome P450 enzymes, suppress lipid peroxidation, and enhance antioxidant  
211 defence. Our findings align with earlier studies reporting hepatoprotective effects of  
212 *Andrographis paniculata* extracts in animal models .

213

214 Its *Ushna virya* supports *Agnivardhana*, facilitating detoxification and regeneration of *Yakrit*  
215 *dhatu*.

216 **Guna–Karma Relationship**

- 217 • *Tikta rasa* → Detoxifies liver, enhances bile flow, and clears *Ama*.
- 218 • *Laghu-Rukshaguna* → Reduces *Medodushti* and *Kleda* accumulation in hepatic tissue.
- 219 • *Ushna virya* → Stimulates metabolism (*Agni*).
- 220 • *Katu vipaka* → Ensures clearance of metabolic wastes (*Mala nissarana*).

221 Hence, the hepatoprotective effect observed experimentally is a physiological manifestation of  
222 these *gunas* acting in synergy.

223

224 The experimental findings scientifically validate traditional Ayurvedic reputation of Kalmegha'  
225 as a potent hepatoprotective agent, demonstrating its ability to effectively prevent and reverse  
226 paracetamol-induced liver injury in Wistar rats. Notably, the decoction form outperformed the  
227 diluted powder suspension, likely due to enhanced solubility and bioavailability of key  
228 phytoconstituents like andrographolide.

## 229 **Conclusion**

230 • Paracetamol successfully induced hepatotoxicity, evidenced by elevated ALT, AST,  
231 bilirubin, and histopathological damage.

232 • Kalmegha treatment significantly normalized biochemical markers and restored liver  
233 architecture.

234 • Decoction exhibited superior efficacy compared to diluted powder form.

## 235 **Clinical Implications**

236 These results affirm Ayurveda's *Samadosha Samagnischa Samadhatu Mala Kriya* principle,  
237 wherein Kalmegha restores doshic equilibrium and physiological homeostasis. As a safe,  
238 economical, and effective natural remedy, Kalmegha decoction holds substantial promise for  
239 integration into modern hepatology protocols for both prevention and management of drug-  
240 induced liver injury.

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## 242 **REFERENCES**

243

- 244 1. GBD 2019 Diseases and Injuries Collaborators. Global burden of 632 diseases and  
245 injuries in 204 countries and territories, 1990-2019. *Lancet.* 2020;396(10258):1204-22.
- 246 2. Trivedi KP, Rawal BK. *Introduction to hepatology*. Ahmedabad: NB Patel; 1999.
- 247 3. Priyavrat Sharma A. *Priye Nighantu*. Varanasi: Chaukhamba Orientalia; 2001. p. 136  
248 (Priyanguvarga).
- 249 4. Sharma PC, Yelne MB, Dennis TJ. *Dravyaguna Vijnana*. Varanasi: Chaukhamba  
250 Bharati Academy; 2005.
- 251 5. Ambike VN, Bhide MB. *Vanoushadhi Nidarsika*. Pune: Saptarishi Prakashan; 1970.

252 6. Garge VM. *Ayurvedic Pharmacology*. Mumbai: Vaidyasagar Prakashan; 1995.

253 7. Hossain MS, Urbi Z, Sule A, Hafizur Rahman KM. *Andrographis paniculata* (Burm.f.)  
254 Wall. ex Nees: a review of its ethnopharmacology, phytochemistry, and pharmacological  
255 activities. *J Pharm Pharmacol.* 2014;66(12):1371-95.

256 8. Jayakumar T, Hsieh CY, Lee JJ, Sheu JR. Experimental and Clinical Pharmacology of  
257 *Andrographis paniculata* and Its Major Bioactive Constituent Andrographolide. *Front  
258 Pharmacol.* 2013;4:125.

259 9. Gupta P, Yadav D, Bisen PS. Evaluation of hepatoprotective activity of *Andrographis  
260 paniculata* against paracetamol-induced hepatotoxicity. *J Pharmacogn Phytochem.*  
261 2022;11(11S):467-72.

262 10. Thomas T, Sabu MC, Kuttan R. Hepatoprotective and anti-inflammatory activities  
263 of *Andrographis paniculata*. *Pharm J.* 2023;1(1):295-300.

264 11. Verma VK, Saraf SK, Tripathi P. Comparison of hepatoprotective activity of *Swertia  
265 chirayita* and *Andrographis paniculata* against CCl induced hepatotoxicity in  
266 rats. *Pharmacologyonline.* 2013;3:104-13.

267 12. Shrivastava S, Gilhotra R. Hepatoprotective activity of *Andrographis paniculata* in  
268 CCl induced liver damage in Wistar rats. *Int J Pharm Sci Res.* 2017;8(10):4321-7.

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