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### RESEARCH ARTICLE

## BRASSINOSTEROIDS A INTER KINGDOM SIGNALING MOLECULES MODULATING STERIOIDogenesis IN POLYCYSTIC OVARY SYNDROME AN IN SILICO STUDY

Velan Athithan

1. Department of Biochemistry, Jawaharlal Institute of Post Graduate Medical Education and Research, Puducherry-605006, India.

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### Abstract

Understanding the influence of ubiquitously present plant hormone brassinosteroids on ovarian steroidogenesis in polycystic ovarian syndrome (PCOS) is currently of interest. Ovarian tissue steroidogenesis depends upon the availability of cholesterol besides the catalytic activity of  $17\beta$ -dehydrogenases and aromatase is a major regulatory step in ovarian steroidogenic pathway. Ovarian testosterone and estradiol biosynthesis is feedback regulated by GnRH, FSH and LH acting through membrane bound hormone receptors. Brassinosteroids is a polyoxygenated derivative of cholesterol, showing structural similarities with animal oxysterol and available to human through diet, exhibiting antihyperglycemic, anticholesterolemic and antiviral effects. Present study intends to investigate brassinosteroids against aromatase,  $17\beta$  hydroxysteroid dehydrogenase, androgen and estrogen receptors as a therapeutic target. ADME properties of brassinosteroids molecules were evaluated using swiss ADME tool. In Silico molecular docking study were performed via AutoDock version 4.0. Brassinosteroids molecules were exhibit high docking score against the aromatase,  $17\beta$  hydroxysteroid dehydrogenase, androgen and estrogen receptors as compared to standard ligand. Dietary intake of brassinosteroids can be potentially down regulating ovarian steroidogenesis in PCOS.

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### Introduction:-

Polycystic ovary syndrome (PCOS) is a multifactorial endocrine disorder affecting 8-20% women worldwide at child bearing age. Being a heterogeneous in nature of PCOS are represented multifaceted symptoms, which predominantly the hyperandrogenism, multiple fluid-filled cystic ovarian morphology, anovulation, hirsutism, androgenic alopecia, acne, clitoromegaly and infertility [13]. PCOS, in general, altered the metabolic steady states such as insulin resistance, increased blood triglyceride, glucose intolerance, obesity, type 2 diabetes, hypertension and cardiovascular disease therefore affecting tissue function [19]. Stien and Leventhal in 1935 proposed the PCOS a state associated with amenorrhoea, enlarged ovary, hirsutism and infertility. However, still there is no diagnostic test

undeniably determines PCOS, nevertheless, in 2013 Rotterdam diagnostic criteria proposed and 2018 firm international evidence based guidelines for the assessment and management of PCOS agreed Rotterdam diagnostic criteria based PCOS diagnosis [5,6]. Rotterdam criteria of PCOS subject at least having two of the three clinical presentations are polycystic ovarian morphology (PCOM), anovulation and hyperandrogenism. While, based on Rotterdam criteria standard the PCOS subject divided into four phenotypes: (1) hyperandrogenism, anovulation and PCOM, (2) anovulation, PCOM and non hyperandrogenism (3) anovulation, hyperandrogenism and normal ovaries, (4) PCOM and hyperandrogenism and normal ovulation. Nevertheless, the molecular pathogenesis underlying in PCOS was unclear, however, suggestive of hyperandrogenism causes high level of estrogen triggered release of increased level of luteinizing hormone (LH) and other hand decrease the follicular stimulating hormone (FSH) secretion from the anterior pituitary.

The low levels of FSH impede to stimulation, maturation and ovulation and along with increased estrogen level induce endometrial hyperplasia at the end [5]. Hence, FSH secretion can be enhanced by reducing estrogen level with inhibition of aromatase enzyme which converting testosterone to estrogen. Blocking the estrogen receptors are (ER $\alpha$ ) another target for the FSH secretions enhancement. 17 $\beta$  dehydrogenases type 1 (17 $\beta$ HSD) catalyzes the androstenedione to testosterone conversion and inhibiting 17 $\beta$ HSD can be reduced androgen level [7, 18, 20]. Now clear that those drug target on PCOS can be preventing further complications. Currently using drug molecules showed unwanted side effects importantly osteoporosis, impotence and hepatotoxicity [21]. Hence, searching the medicine from plant source is continuously being investigated in PCOS treatment.

Brassinosteroid (BR) is a polyoxygenated sterol comes under the class six of phyto hormones. In 1979 reported first active form hormone termed brassinolide, followed by 70 hormones in the class identified [16]. BRs ubiquitously existing in all the plants and regulates seedling, growth, flowering and present to withstand biotic and abiotic stress effects [14]. However, BRs mimic structural similarity with animal hormones estrogen, androgen and insect ecdysteroid. Brassinosteroids are consumed by human through food and herbal based folk medicine [12]. Assimilation of brassinosteroids and enter into organs through general blood circulation resulted to down regulation of glucose, cholesterol, triglycerides, LDH, proinflammatory cytokines IL-1, TNF- $\alpha$ , COX-2 and up-regulation of glycogen, HDL and transactivation of nuclear receptor in animal cells [3,4].

Conventional pharmacological methods of drug discovery are time-consuming, labour-intensive and expensive. An alternative concept of reverse pharmacology could be a major breakthrough in the field of drug discovery. The basis of reverse pharmacology includes In Silico analogue designing and ligand-receptor interaction and ADMET studies recommendation of chemical nature of molecules [15]. Nevertheless, in the present In Silico study, we intend to investigate the anti-androgenic and anti-estrogenic potential of specific brassinosteroids against aromatase, 17 $\beta$ -hydroxysteroid dehydrogenase type 1, androgen receptor and estrogen receptor as an ideal drug target in PCOS.

## Material and Methodology:-

### ADMET properties evaluation:

Brassinosteroid molecules were evaluated pharmacokinetic and pharmacodynamic properties of Absorption, Distribution, Metabolism, Excretion and Toxicity (ADMET) were determined using ADMET prediction online server (<http://admet.scdbb.com>) [15].

### Ligand preparation:

Brassinosteroid and standard compound three-dimensional structures were downloaded from PubChem database (<http://pubchem.ncbi.nlm.nih.gov/>) as .SDF file format of Brassinolide (CID: 115196), 28-Homobrassinolide (CID: 11038340), 24-Epibrassinolide (CID: 443055), Castasterone (CID: 133534), 28-Homocastasterone (CID: 5487654), 28-Norbrassinolide (CID: 13845880), 28-Norcastasterone (CID: 13982110), 24-Epicastasterone (CID: 11812633), 3,24-Diepicastasterone (CID: 10961603), 6 $\alpha$ -Hydroxycastasterone (CID: 15542699), 6-Deoxocastasterone (CID: 13870433), 6-Deoxo-28-norcastasterone (CID: 101682290), Abiraterone (CID: 132971), Androstenedione (CID: 6128), Letrozole (CID: 3902), Anastrozole (CID: 2187), Testosterone (CID: 6013), Flutamide (CID: 3397), Testosterone (CID: 6013), Tamoxifen (CID: 2733526) and Estradiol (CID: 5757) followed by converted into .mol2 file format using Open Babel User Interface software version 2.4.1 and ligands were optimized by means of ligand preparation script in AutoDock ver. 4.0. program. Ligands were prepared for docking as torsion tree, root detection, torsion number were set and saved in .pdbqt file format (Figure 1) [23].

**Protein preparation:**

X-ray crystallographic three-dimensional structures of aromatase (PDB ID: 5JKV), 17 $\beta$ -hydroxysteroid dehydrogenase type1 (PDB ID:1FDS), androgen receptor (PDB ID: 1E3G) and estrogen receptor (PDB ID: 3ERT) were retrieved from RCSB Protein Data Bank (<http://www.rcsb.org>). Consequently, AutoDockver. 4.0.script employs the removal of water molecules, addition of polar hydrogen atoms, assignment of Kollman charges and conversion of the protein files in .pdb format for further molecular docking study (Figure 2)[23].

**Grid box generation:**

3D structures of selected protein and ligand structure were together to form a grid. Therefore, centroid of the ligand molecule in complex protein structure were chosen to generate grid points X = 60, Y = 60 and Z = 60 axis set for molecular docking. The grid file generated by means of "grid generation panel" in AutoDock software version 4.0 [23].

**Molecular docking simulation:**

Protein-ligand molecular docking performed using AutoDock software version 4.0. For each ligand (chemical structure), 100 docking runs with default parameters were performed by treating protein as rigid and the ligand as flexible. The results were visualized using PyMol (The PyMOL Molecular Graphics System, Version 1.5.0.4 Schrodinger, LLC), wherein all the conformations for each of the ligand was found to be within the cavity of protein indicating that the docking run was free from errors. The conformational clusters with lowest binding energy were considered for further analysis [23].

**Evaluation of the total binding energy:**

The AutoDock ver. 4.0. algorithms were applied to evaluate the total binding energy of ligand against target proteins. Various docked conformations were obtained and one with lowest binding energy towards ligand binding cavity of protein were selected as possible binding conformation and considered for further protein-ligand interaction analysis. The final evaluations of the interactions between the target ligand and amino acid residues of the ligand binding cavity of protein were analyzed using BIOVIA Discovery Studio 2021 script[23].

**Results:-**

Brassinosteroids has shown acceptable water solubility (Log S) score with the human intestinal absorption indicative of 90%. While, the Blood-Brain barrier permeability shown acceptable range score thus denotes no ligands molecules have the blood-brain barrier permeability. Also, observed that no brassinosteroids molecules shown CYP450 enzymes inhibition, denoted that ligands metabolized by CYP450 enzyme in liver. Although, evaluation of brassinosteroid molecules bioavailability in blood and tissues noted that acceptable result indicative of potential bioactive molecule induce cellular effects and similarly, observed good renal clearance scores. The AMES test shown there was no mutagenicity property associated with the brassinosteroids molecules (Table 1 and 2). The 17 $\beta$ HSD binding interactions with dietary phyto oxysterol brassinosteroids were studied by In Silico docking simulations.

Based on docking energies, 17 $\beta$ -Hydroxysteroid dehydrogenase and brassinolide, 28-Homobrassinolide, 24-Epibrassinolide, castasterone, 28-Homocastasterone, 28-Norbrassinolide, 28-Norcastasterone, 24-Epicastasterone, 3,24-Diepicastasterone, 6 $\alpha$ -Hydroxycastasterone, 6-Deoxocastasterone and 6-Deoxo-28-norcastasterone exhibited binding affinity -6.82, -6.60, -8.78, -9.02, -8.42, -7.75, -8.69, -8.39, -6.78, -9.16, -8.97 and -8.68 Kcal/mole respectively as compared to abiraterone and Androstenedione presented binding affinity -7.68, -7.04 Kcal/mol with 17 $\beta$ HSD (Table 3 and 7).

Brassinolide forms hydrogen bond with GLU163, GLU167, ARG266 amino acid residues, while it forms Van der Waals interaction with LEU146, THR250, LEU251, LEU260, LEU263 amino acid residues and amino acid residues ARG252, PHE254, LEU267 form Alkyl/ $\pi$ -Alkyl interactions with 17 $\beta$ HSD enzyme. 28-Homobrassinolide forms hydrogen bond with ARG252, ARG266 residues, while it forms Van der Waals interaction with GLU163, GLY164, THR250, LEU251 amino acid residues and amino acid residues LEU146, PHE254, LEU267 form Alkyl/ $\pi$ -Alkyl interactions with enzyme. 28-Homocastasterone forms hydrogen bond with residues GLY145, while it forms Van der Waals interaction with amino acid residues MET147, PHE160, GLU163, GLU167, THR250, LEU251, TYR253 and amino acid residues LEU146, ARG252, PHE254, LEU263, LEU267 form Alkyl/ $\pi$ -Alkyl interactions. 28-Norbrassinolide forms hydrogen bond with residues LEU146, ARG266, while it forms Van der Waals interaction with amino acid residues GLY145, MET147, GLU163, GLY164, THR250, LEU251, ARG252, PHE254 and

amino acid residues PHE160, LEU263, LEU267 form Alkyl/ $\pi$ -Alkyl interactions. 28-Norcastasterone forms hydrogen bond with GLY145 residues, while it forms Van der Waals interaction with MET147, PHE160, GLU163, GLU167, THR250, LEU251, LEU260, ARG266 amino acid residues and LEU146, ARG252, PHE254, LEU263 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 24-Epicastasterone forms hydrogen bond with GLY145 residues, while it forms Van der Waals interaction with MET147, PHE160, GLU163, GLU167, THR250, LEU260, ARG266 amino acid residues and LEU146, ARG252, PHE254, LEU263, LEU267 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions.

3,24-Diepicastasterone forms hydrogen bond with PHE160 residues, while it forms Van der Waals interaction with GLY145, GLU163, GLY164, GLU167, ARG252, LEU260, ARG264, ARG266 amino acid residues and LEU146, PHE254, LEU263, LEU267 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 6 $\alpha$ -Hydroxycastasterone forms hydrogen bond with GLY145, GLU167 residues, while it forms Van der Waals interaction with MET147, PHE160, GLU163, THR220, LEU251, LEU260, ARG266 amino acid residues and LEU146, ARG252, PHE254, LEU263, LEU267 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 6-Deoxocastasterone forms hydrogen bond with GLY145, ARG252 residues, while it forms Van der Waals interaction with GLU163, GLY164, GLU167, LEU251, ARG266, LEU267 amino acid residues and LEU146, ARG160, PHE254, LEU263 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions.

Likewise, 6-Deoxo-28-norcastasterone forms hydrogen bond with GLY145 residues, while it forms Van der Waals interaction with MET147, PHE160, GLU163, GLU167, THR250, LEU251, LEU260, ARG266 amino acid residues and LEU146, ARG252, PHE254, LEU263, LEU267 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions with 17 $\beta$ HSD enzyme (Table 3 and Fig 4, 8).

Likewise, the binding studies performed between abiraterone and androstenedione and 17 $\beta$ HSD enzyme, indicates abiraterone interacting with amino acid residues PHE160 via hydrogen bond and with amino acid residues GLY145, GLU163, GLY164, ARG252, PHE254, ARG264, ARG266 via Van der Waals interaction and LEU146, LEU260, LEU267 residues via Alkyl/ $\pi$ -Alkyl interactions and amino acid LEU263 via  $\pi$ -sigma. On the other hand, androstenedione interacts with 17 $\beta$ HSD amino acid residues GLY164 via hydrogen bond, while GLY145, PHE160, GLU163, GLU167, ARG252, PHE254, ARG266 amino acid residues via Van der Waals interactions and amino acid residues LEU146, LEU263, LEU267 via Alkyl/ $\pi$ -Alkyl interactions with lowest binding affinity with 17 $\beta$ HSD enzyme (Table 7 and Fig 3, 8).

The aromatase binding interactions with dietary phyto oxysterol brassinosteroids were studied by In Silico docking simulations. Based on docking energies obtained, aromatase exhibited binding affinity towards Brassinolide, 28-Homobrassinolide, 24-Epibrassinolide, Castasterone, 28-Homocastasterone, 28-Norbrassinolide, 28-Norcastasterone, 24-Epicastasterone, 3,24-Diepicastasterone, 6 $\alpha$ -Hydroxycastasterone, 6-Deoxocastasterone and 6-Deoxo-28-norcastasterone is -12.51, -11.54, -12.26, -12.44, -12.36, -12.25, -10.23, -10.99, -11.86, -10.24, -10.53 and -11.10 Kcal/mol respectively as compared to letrozole, anastrozole and testosterone presented binding affinity -7.93, -9.6 and -9.85 Kcal/mol with aromatase (Table 4 and 7).

In aromatase enzyme interaction, ligand brassinolide interacted with ARG115 and LEU477 residues via hydrogen bond and ILE132, TRP141, ARG145, ALA306, ASP309, THR310, LEU372, VAL373, ARG435, GLY439, SER478 residues via Van der Waals interaction and ILE133, PHE134, PHE221, TRP224, VAL370, MET374, CYS437, ALA438 residues via Alkyl/ $\pi$ -Alkyl interactions (Table). Similarly, in aromatase-28-Homobrassinolide interaction, ARG115, LEU372, MET374, ALA438, GLY439 amino acid residues form hydrogen bonds with ligand, while PHE221, TRP224, ALA307, THR310, MET311, VAL370, VAL373, ALA443, ILE442, MET446, LEU477, SER478 residues form Van der Waals interactions.

Ligand 24-Epibrassinolide interacted with ARG115, LEU477 residues via hydrogen bond and ILE132, ILE133, TRP141, ARG145, ALA306, ASP309, THR310, VAL370, LEU372, VAL373, ARG435, CYS437, ALA438, GLY439, SER478 residues via Van der Waals interaction and PHE134, PHE221, TRP224, MET374 residues via Alkyl/ $\pi$ -Alkyl interactions. Ligand Castasterone interacted with LEU372, MET374, CYS437 residues via hydrogen bond and ILE132, LEU152, PHE221, GLU302, THR310, VAL373, GLY439, SER478 residues via Van der Waals interaction and ILE133, PHE134, PHE148, TRP224, ALA306, MET303, VAL370, ALA438, LEU477 residues via Alkyl/ $\pi$ -Alkyl interactions. Ligand 28-Homocastasterone interacted with LEU372, MET374, GLY439 residues via hydrogen bond and ARG115, LEU151, PHE221, GLU302, MET303, THR310, VAL373, LEU477 residues via Van

der waals interaction and ILE132, ILE133, PHE134, PHE148, TRP224, ALA306, VAL370, CYS437, ALA438 residues via Alkyl/ $\pi$ -Alkyl interactions. Ligand 28-Norbrassinolide interacted with ARG115, LEU372, LEU477 residues via hydrogen bond and ILE132, TRP141, ARG145, PHE221, ALA306, THR310, VAL373, SER478, ARG435, ALA438, GLY439 residues via Van der waals interaction and ILE133, PHE134, TRP224, MET374, CYS437 residues via Alkyl/ $\pi$ -Alkyl interactions. Ligand 3,24-Diepicasterone interacted with ARG115, LEU477 residues via hydrogen bond and ILE132, TRP141, ARG145, ALA306, ASP310, VAL370, LEU372, VAL373, ARG435, GLY436, SER478 residues via Van der waals interaction and ILE133, PHE134, PHE221, TRP224, MET374, CYS437, ALA438 residues via Alkyl/ $\pi$ -Alkyl interactions (Table 4 and Fig 5, 9).

Likewise, the binding studies performed between letrozole, anastrozole, testosterone and aromatase enzyme, indicates testosterone interacting with amino acid residues ARG 115, ALA 306, MET 374 via hydrogen bond and amino acid residues PHE134, PHE221, ILE305, ASP309, THR310, ILE133, LEU372, VAL373, LEU477, SER478 via Van der waals interaction and TRP224, VAL370 residues via Alkyl/ $\pi$ -Alkyl interactions. On the other hand, letrozole interacts with aromatase amino acid residues ARGY115, TRP141 via hydrogen bond, while ILE132, ILE133, ARG145, LEU152, MET303, ALA306, VAL373, PHE430, GLY431, GLY439 amino acid residues via Van der waals interactions and amino acid residues ARG435 via Alkyl/ $\pi$ -Alkyl interactions and amino acid residues CYS437, ALA438 via  $\pi$ -sigma interactions and amino acid residues CYS437 form  $\pi$ -sulfur interaction with lowest binding affinity. Similarly, the anastrozole interacting with amino acid MET311 via hydrogen bond and amino acid residues SER314, THR310, MET364, PRO368, VAL369, PRO429, CYS437, ALA443 via Van der waals interaction and PHE430, VAL370 via Alkyl/ $\pi$ -Alkyl interactions with aromatase (Table 7 and Fig 3, 9).

In the current In Silico study, androgen receptor was docked with the dietary phytosterols brassinosteroids. The docking scores of Brassinolide, 28-Homobrassinolide, 28-Homocasterone, 28-Norbrassinolide, 2-Deoxybrassinolide, 28-Norcastasterone, 24-Epicasterone, 2-Epicasterone, 3,24-Diepicasterone, 6- $\alpha$ -14-Hydroxycasterone, 6-Deoxocasterone, 6-Deoxo-28-norcastasterone against androgen receptor is -5.65, -4.12, -4.42, -5.05, -5.89, -4.25, -4.88, -3.09, -6.66 and -6.54 Kcal/mol respectively as compared to flutamide and testosterone presented binding affinity -0.83 and -1.11 Kcal/mol with androgen receptor (Table 5 and 7).

In androgen receptor, the brassinolide forms hydrogen bond with amino acid residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions with androgen receptor. 28-Homobrassinolide forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions with androgen receptor. 28-Homocasterone forms hydrogen bond with residues while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 28-Norbrassinolide forms hydrogen bond with residues while it forms Van der waals interaction with amino acid residues and amino acid residues PHE160, LEU263, LEU267 form Alkyl/ $\pi$ -Alkyl interactions.

28-Norcastasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 24-Epicasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 3,24-Diepicasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 6- $\alpha$ -Hydroxycasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 6-Deoxocasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. Likewise, 6-Deoxo-28-norcastasterone forms hydrogen bond with residues, while it forms Van der waals interaction with amino acid residues and amino acid residues form Alkyl/ $\pi$ -Alkyl interactions with androgen receptor (Table 5 and Fig 6, 3, 10).

Likewise, the binding studies performed between flutamide, testosterone and androgen receptor, indicates flutamide interacting with amino acid residues SER740, SER814, GLN867 via hydrogen bond and with amino acid residues THR739, MET742, GLY743, LEU744, LEU812, VAL866 via Van der waals interaction and LEU811, ILE815, ALA870, ARG871, HIS874, ILE906, PRO913 residues via Alkyl/ $\pi$ -Alkyl interactions. On the other hand, testosterone interacts with androgen receptor amino acid residues MET745 via hydrogen bond, while GLN711, ALA748, MET780, LEU873, PHE876, THR877 amino acid residues via Van der waals interactions and amino acid

residues LEU704, LEU707, MET742, VAL746, MET749, PHE764 via Alkyl/ $\pi$ -Alkyl interactions with lowest binding affinity as compared to the dietary brassinosteroids (Table 7 and Fig 3, 10).

In the current *In Silico* study, estrogen receptor was docked with the dietary phytosterols brassinosteroids. The docking scores of Brassinolide, 28-Homobrassinolide, 28-Homocastasterone, 28-Norbrassinolide, 2-Deoxybrassinolide, 28-Norcastasterone, 24-Epicastasterone, 2-Epicastasterone, 3,24-Diepicastasterone, 6- $\alpha$ -14-Hydroxycastasterone, 6-Deoxocastasterone, 6-Deoxo-28-norcastasterone against estrogen receptor is -12.96, -9.56, -9.63, -13.33, -10.66, -10.19, -10.98, -12.93, -11.99, -10.66, -12.78 and -11.36 Kcal/mole respectively as compared to tamoxifen and estradiol presented binding affinity and -11.03 and -9.69 Kcal/mol with estrogen receptor (Table 6 and 7).

Brassinolide forms hydrogen bond with amino acid residues ASP351 and ASN532, while it forms Van der Waals interaction with amino acid residues THR347, LEU384, LEU387, MET388, LEU391, MET421, PHE422, LEU428, VAL534 and amino acid residues LEU346, ALA350, LEU354, TRP383, PHE404, LEU525, MET528, PRO535 form Alkyl/ $\pi$ -Alkyl interactions with estrogen receptor. 28-Homobrassinolide forms hydrogen bond with residues THR347, ASP351, GLU353, while it forms Van der Waals interaction with amino acid residues LEU349, LEU391, ARG394, PHE404, MET421, ILE424, PHE425, LEU428, ASN532, VAL534 and amino acid residues MET343, LEU346, ALA350, LEU354, TRP383, LEU387, LEU525, MET528, PRO535 form Alkyl/ $\pi$ -Alkyl interactions with estrogen receptor.

Ligand 24-Epibrassinolide interacted with residue ASP351 via hydrogen bond and residues MET343, LEU346, THR347, LEU387, LEU391, PHE404, MET421, ILE424, PHE425, LEU428, HIS524, VAL534 via Van der Waals interaction and ALA350, LEU354, LEU525, VAL533, PRO535 residues via Alkyl/ $\pi$ -Alkyl interactions. Ligand Castasterone interacted with residues ASP351, ASN532 via hydrogen bond and residues THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, PHE425, LEU428, VAL534 via Van der Waals interaction and residues LEU346, ALA350, LEU354, PHE404, LEU525, MET528, VAL533, PRO535 via Alkyl/ $\pi$ -Alkyl interactions. 28-Homocastasterone forms hydrogen bond with residue THR347 while it forms Van der Waals interaction with amino acid residues MET343, GLU353, TRP383, LEU384, MET388, ARG394 and LEU346, LEU349, ALA350, LEU387, LEU391, PHE404, LEU428, LEU525 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions.

28-Norbrassinolide forms hydrogen bond with ASP351, ASN532 residues while it forms Van der Waals interaction with amino acid residues MET343, THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, PHE425, LEU428, VAL534 and LEU346, ALA350, LEU354, TRP383, PHE404, LEU525, MET528, VAL533, PRO535 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions. 28-Norcastasterone forms hydrogen bond with THR347 residue, while it forms Van der Waals interaction with amino acid residues ASP351, GLU353, LEU354, LEU384, MET388, LEU391, ARG394, PHE404, PHE425, LEU428, MET528, ASN532, VAL533, VAL534, PRO535 and amino acid residues MET343, LEU346, ALA350, TRP383, LEU387, MET421, LEU525 form Alkyl/ $\pi$ -Alkyl interactions. 24-Epicastasterone forms hydrogen bond with residues ASP351 and ASN532, while it forms Van der Waals interaction with amino acid residues THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, LEU428, VAL534 and amino acid residues LEU346, ALA350, LEU354, PHE404, LEU525, MET528, VAL533, PRO535 form Alkyl/ $\pi$ -Alkyl interactions.

3,24-Diepicastasterone forms hydrogen bond with residue ASP351, while it forms Van der Waals interaction with amino acid residues THR347, GLU353, LEU354, LEU391, ARG394, PHE404, PHE425, LEU428, ASN532, VAL533, VAL534, THR347, ARG394, PHE404, MET421, PHE425, LEU428, VAL428, VAL534 and amino acid residues MET343, LEU346, ALA350, TRP383, LEU384, LEU387, MET388, MET421, LEU525, PRO535 form Alkyl/ $\pi$ -Alkyl interactions. 6- $\alpha$ -Hydroxycastasterone forms hydrogen bond with ASN532 residue, while it forms Van der Waals interaction with LEU346, THR347, LEU349, ASP351, GLU353, LEU354, MET421, ILE424, PHE425, LEU428, GLY521, VAL534 amino acid residues and amino acid residues ALA350, TRP383, LEU384, LEU387, MET388, LEU391, PHE404, VAL533, PRO535 form Alkyl/ $\pi$ -Alkyl interactions. 6-Deoxocastasterone forms Van der Waals interaction with MET343, THR347, ASP351, GLU353, TRP383, LEU384, MET388, ARG394, MET421, PHE425, ASN532, PRO535 amino acid residues and LEU346, LEU349, ALA350, LEU387, LEU391, PHE404, LEU525, VAL533 amino acid residues form Alkyl/ $\pi$ -Alkyl interactions.

Likewise, 6-Deoxo-28-norcastasterone forms LEU346, LEU347, GLU353, TRP383, MET388, ARG394, PHE404, PRO535 Van der waals interaction with amino acid residues and amino acid residues MET343, ALA350, LEU384, LEU387, LEU391, MET421, ILE424, PHE425, LEU428, LEU525, VAL533 form Alkyl/ $\pi$ -Alkyl interactions with estrogen receptor (Table 6 and Fig 7, 3, 11).

Likewise, the binding studies performed between thetamoxifen, estradiol and estrogen receptor, indicates interacting with amino acid residues ASP351, VAL534 via hydrogen bond and with amino acid residues GLU353, LEU354, TRP383, LEU384, MET388, MET421, ILE424, PHE425, GLY521, HIS524, VAL533, PRO535 via Van der waals interaction and LEU346, LEU349, ALA350, LEU387, PHE404, LEU525 residues via Alkyl/ $\pi$ -Alkyl interactions and amino acid residue LEU525 form  $\pi$ -sigma interaction. On the other hand, estradiol interacts with estrogen receptor amino acid residues GLU353, ARG394, GLY521 via hydrogen bond, while amino acid residues LEU346, LEU349, MET421, LEU428, HIS524 via Van der waals interactions and amino acid residues ALA350, LEU384, LEU387, MET388, LEU391, ILE424, LEU525 via Alkyl/ $\pi$ -Alkyl interactions and amino acid residue PHE404 form  $\pi$ -sigma interaction with lowest binding affinity as compared to the dietary brassinosteroids (Table 7 and Fig 3, 11).

### Discussion:-

Understanding on enzyme and cellular receptors functions in ovarian steroidogenic pathways provides several occurrences of feedback regulation. Proof of ovarian steroidogenic pathway modulator through In Silico methods have accepted exogenous molecules act as active site modifiers of specific enzymatic function in steroidogenic metabolic pathways, indicating rate limiting regulatory phenomena in cells. The important enzymatic regulatory step in the steroidogenic biosynthetic pathway leading to the synthesis of testosterone, estrogen in ovarian cells is that involving the transformation of cholesterol to pregnenolone (enzyme P450<sub>scc</sub>) into androstenedione to testosterone (enzyme 17 $\beta$ HSD) and testosterone to estradiol (aromatase) in the theca and granulosa cells. Further, androgen and estrogen biosynthesis and homeostasis are under the regulation of GnRH, LH and FSH signaling mediators [22].

Human 17 $\beta$ -hydroxysteroid dehydrogenase type 1 is comes under the steroid dehydrogenase reductase family. 17 $\beta$ HSD catalyzes reduction of androstenedione to testosterone, in the presence of NADPH as a cofactor in the ovarian granulosa cell [18]. The hyperandrogenism a main hallmark in PCOS resulted to anovulation, polycystic ovarian morphology, which can be over expression of 17 $\beta$ HSD enzyme. Hence 17 $\beta$ HSD enzyme inhibition can be ideal therapeutic management in PCOS individual [2]. In the present In Silico study observed that valuable insights between brassinosteroids ligands with 17 $\beta$ HSD a key enzyme involved in ovarian steroidogenesis pathway. The brassinosteroids compounds 6 $\alpha$ -Hydroxycastasterone, castasterone, 6-Deoxocastasterone, 24-Epibrassinolide, 6-Deoxo-28-norcastasterone shown high binding avidity towards 17 $\beta$ HSD enzyme compared to the reference compounds abiraterone (-7.68 kcal/mol) and androstenedione (-7.04 kcal/mol) highlights their promising inhibitors for the 17 $\beta$ HSD enzyme (Table 3 and 7).

Individual with PCOS causes the oligomenorrhoea, amenorrhoea and anovulation resulted by higher level of estrogen in PCOS, that affecting normal ovarian physiology. The aromatase enzyme inhibitoris make ovulation happen in PCOS. The aromatase enzyme comes under the cytochrome p450 family, secreted by granulosa cell of ovary and catalyzes the conversion of testosterone to estrogen irreversibly. This catalytic reaction is the final and rate limiting step in the ovarian estrogen synthesis [9, 10]. In the present In Silico study observed that brassinolide, castasterone, 28-Homocastasterone, 24-Epibrassinolide, 28-Norbrassinolide, 3,24-Diepicastasterone, 28-Homobrassinolide, 6-Deoxo-28-norcastasterone shown highest binding affinities towards aromatase enzyme compared to reference compounds testosterone (-9.85 kcal/mol), anastrozole (-9.60 kcal/mol) and letrozole (-7.93 kcal/mol) that highlights their promising inhibitors for the aromatase enzyme (Table 4 and 7). However, daily intake of brassinosteroids may be down regulate estrogen level resulted to preventing the PCOS effects on ovulation.

Androgen receptor mediated signaling mechanism in PCOS resulted by altered phenotype traits that down regulation of follicles development leading to multiple small cysts in ovary caused the anovulation, hirsutism, acne and alopecia in peripheral tissue [17]. Therefore, inhibition of androgen with androgen receptor interaction is idyllic phenomenon to managing PCOS complications. Although in the present work the brassinosteroids compounds 6-Deoxocastasterone, 6-Deoxo-28-norcastasterone, 28-Norcastasterone, brassinolide, 28-Norbrassinolide, 3,24-Diepicastasterone, 28-Homocastasterone, 28-Homobrassinolide shown highest binding avidity towards androgen receptor compared to reference compounds testosterone (-1.11 kcal/mol) and flutamide (-0.83 kcal/mol) (Table 5 and 7) to inhibition of androgen receptor potentially. However, the brassinosteroids may be down regulate androgen receptor impact on the PCOS individual and prevent complications.

Estrogen receptor can be localized in cell membrane, cytoplasm and nucleus. The estrogen interact with estrogen receptor resulted to genomic or non-genomic effects such as transcriptional and cell division effects on target tissues. In PCOS estrogen receptor inducing estrogen synthesis that causes increased level of estrogen resulted to anovulation. Therefore, targeting estrogen receptor in PCOS individual is idyllic phenomenon [1, 24]. Therefore, in the present investigation carried out estrogen receptor inhibition effects of the brassinosteroids compounds castasterone, brassinolide, 24-Epicastasterone, 6-Deoxocastasterone, 3,24-Diepicastasterone, 6-Deoxo-28-norcastasterone shown highest binding affinities towards estrogen receptor compared to reference compounds tamoxifen (-11.03kcal/mol) and estradiol (-9.69kcal/mol) (Table 6 and 7) highlights their promising inhibitors of the estrogen receptor in PCOS.

Drug molecules with poor pharmacokinetic and pharmacodynamic properties can have adverse impact on human biological system such as alter organ function, immunological reaction, and dermatological issues. However, present investigation the brassinosteroids ADMET properties evaluated. 12 brassinosteroids was evaluated for the water solubility, human intestinal absorption, blood brain barrier permeability, CYP450 isoforms inhibitor or substrate, bioavailability, renal clearance and mutagenic effect using the ADMET online tool. Brassinosteroids has shown the acceptable ADMET results in order intestinal absorption into circulatory system to reach target tissues followed by bio-physiological effects and metabolized in liver than excreted in urine.

Ovarian steroidogenesis a rate limiting steps in testosterone and estrogen biosynthesis had been recognized as catalytic regulation of 17 $\beta$ HSD and aromatase enzyme, the effects of brassinosteroids on enzyme catalytic function was studied by In Silico analysis of interaction between 17 $\beta$ HSD and aromatase enzyme, androgen and estrogen receptor with brassinosteroid ligands. The significant binding avidity between the testosterone, estrogen and brassinosteroid on enzyme catalytic side, suggestive a modulatory effect by brassinosteroids on ovarian steroidogenesis in cell. In the present investigation highlights the effects of brassinosteroids in its capability to down regulate testosterone and estrogen levels and inhibition of androgen and estrogen receptors in PCOS. Further, an In Vivo study is needed to understanding the possible influences contributing brassinosteroids to ovarian steroidogenesis in normal and PCOS subjects.

#### Tables and Figures:

**Table 1. Evaluation of drug-likeness of brassinosteroids molecules using Lipinski rule of five**

Compounds	Molecular Weight (Da)	H-bond donors	H-bond acceptors	LogP Values
Brassinolide	480.686	4	6	3.390
28-Homobrassinolide	494.713	4	6	3.780
24-Epibrassinolide	480.686	4	6	3.390
Castasterone	464.687	4	5	3.806
28-Homocastasterone	478.714	4	5	4.190
28-Norbrassinolide	466.659	4	6	3.140
28-Norcastasterone	450.660	4	5	3.560
24-Epicastasterone	464.687	4	5	3.806
3,24-Diepicastasterone	464.687	4	5	3.806
6 $\alpha$ -	466.703	5	5	3.598



Hydroxycastasterone				
6-Deoxocastasterone	450.704	4	4	4.627
6-Deoxo-28-norcastasterone	436.677	4	4	4.381

**Table 2.ADME properties of Brassinosteroids compounds**

ADME Properties	1	2	3	4	5	6	7	8	9	10
<b>Absorption Properties</b>										
Caco-2 Permeability Optimal: higher than -5.15 Log unit or -4.70 or -4.80	- 4.79 1 cm/s	- 4.827 cm/s	- 4.791 cm/s	- 4.791 cm/s	- 4.793 cm/s	- 4.846 cm/s	- 4.842	- 4.793	- 4.793	- 4.836
Human Intestinal Absorption ≥30%: HIA+; <30%: HIA-	0.61 1	0.599	0.611	0.611	0.689	0.592	0.684	0.689	0.689	0.685
P-glycoprotein Substrate	0.10 1	0.166	0.101	0.101	0.08	0.151	0.123	0.080	0.080	0.167
P- glycoprotein Inhibitor	0.38 4	0.619	0.384	0.384	0.36	0.622	0.603	0.360	0.360	0.330
<b>Distribution Properties</b>										
Plasma Protein Binding90%	84.5 29	84.50 2	84.52 9	84.52 9	83.78 4	83.93 1	83.93 5	83.78 4	83.78 4	82.01 2
Blood brain barrier (BBB) BB ratio ≥0.1: BBB+; BB ratio <0.1: BBB-	0.40 6	0.407	0.418	0.418	0.808	0.388	0.764	0.808	0.808	0.818
Volume Distribution 0.04-20 L/kg	- 0.46 8	- 0.523	- 0.468	- 0.468	- 0.297	- 0.411	- 0.247	- 0.297	- 0.297	- 0.243
<b>Metabolism Properties</b>										
P450 CYP1A2-inhibitor	0.03 9	0.057	0.039	0.039	0.028	0.050	0.039	0.028	0.028	0.026
P450 CYP1A2-substrate	0.31 8	0.370	0.318	0.318	0.330	0.362	0.370	0.330	0.330	0.293
P450 CYP3A4-inhibitor	0.20 3	0.319	0.203	0.203	0.109	0.355	0.216	0.109	0.109	0.133
P450 CYP3A4-substrate	0.66 7	0.683	0.667	0.667	0.687	0.658	0.688	0.687	0.687	0.587
P450 CYP2C9-inhibitor	0.19 5	0.289	0.195	0.195	0.116	0.234	0.151	0.116	0.116	0.137
P450 CYP2C9-substrate	0.18 5	0.214	0.185	0.185	0.194	0.194	0.193	0.194	0.194	0.217
P450 CYP2C19-inhibitor	0.09 5	0.108	0.095	0.095	0.057	0.106	0.059	0.057	0.057	0.107
P450 CYP2C19substrate	0.55 2	0.536	0.552	0.552	0.470	0.567	0.481	0.470	0.470	0.465
P450 CYP2D6-inhibitor	0.26 7	0.289	0.267	0.095	0.245	0.279	0.261	0.245	0.245	0.301
P450 CYP2D6-substrate	0.22 2	0.207	0.222	0.222	0.255	0.240	0.270	0.255	0.255	0.337
<b>Excretion Properties</b>										
T 1/2 (Half Life Time)	1.39	1.447	1.392	1.392	1.551	1.347	1.543	1.551	1.551	1.565

>8h: high, 3h<Cl< 8h: moderate,<3h: low	2									
ClearancemL/min/kg >15 mL/min/kg: high; 5mL/min/kg<Cl< 15mL/min/kg: moderate; <5 mL/min/kg: low	1.38 2	1.32	1.382	1.382	1.385	1.436	1.447	1.385	1.385	1.355
<b>Toxicity properties</b>										
hERG Blockers	-	-	-	0.353	0.401	0.345	0.387	0.401	0.401	0.417
Category 0: Non-blockers	0.35	0.361	0.353							
Category 1: Blockers	3									
AMES test	0.18	0.182	0.186	0.186	0.224	0.212	0.288	0.224	0.224	0.302
Category 0: Ames test -ve	6									
Category 1: Ames test +ve										
Drug Induced Liver Injury	-	-	-	-	0.226	0.244	0.112	0.226	0.226	0.124
Category 0: DILI -ve	0.36	0.242	0.360	0.360						
Category 1: DILI +ve	0									
<b>Physicochemical Properties</b>										
Log S (Solubility)	-	-	-	-	-5.19	-	-	-5.19	-5.19	-
Optimal: higher than -4 log mol/L	4.83 6	5.083	4.836	4.836		4.388	5.012			5.056
Distribution Coefficient	3.31	3.489	3.318	3.318	3.521	3.294	3.565	3.521	3.521	3.516
1 to 3: Solubility moderate, Permeability moderate, Metabolism low.	8									
LogP (LogP<0: poor lipid bilayer permeability. LogP>3: poor aqueous.	3.39	3.78	3.39	3.39	3.806	3.144	3.56	3.806	3.806	3.598

**Compounds:** 1.Brassinolide, 2.28-Homobrassinolide, 3. 4-Epibrassinolide, 4. Castasterone, 5. 28- Homocastasterone, 6.28 Norbrassinolide, 7.28 Norcastasterone, 8.24 Epicastasterone, 9.3,24 Diepicastasterone, 10.6 $\alpha$  Hydroxycastasterone.

**Table.3. Brassinosteroids and 17 $\beta$ -Hydroxysteroid-dehydrogenase enzyme interaction**

17 $\beta$ Hydroxysteroid dehydrogenase. (1FDS)	G-Score kcal/mol	Van der waals	Hydrogen bond	Alkyl/ $\pi$ -Alkyl
<b>Brassinolide</b>	-6.82	LEU146, THR250, LEU251, LEU260, LEU263.	GLU163, GLU167, ARG266.	ARG252, PHE254, LEU267.
<b>28-Homobrassinolide</b>	-6.60	GLU163, GLY164, THR250, LEU251.	ARG252, ARG266.	LEU146, PHE254, LEU267.
<b>24-Epibrassinolide</b>	-8.78	GLU163, GLY164, GLU167, LEU251, ARG252, PHE254, ARG266, LEU267.	GLY145	LEU146, PHE160, LEU263.
<b>Castasterone</b>	-9.02	MET147, PHE160, GLU163, GLU167, THR250, LEU251, ARG266.	GLY145	LEU146, ARG252, PHE254, LEU263, LEU267.
<b>28-Homocastasterone</b>	-8.42	MET147, PHE160, GLU163, GLU167, THR250, LEU251, TYR253.	GLY145	LEU146, ARG252, PHE254, LEU263, LEU267.
<b>28-Norbrassinolide</b>	-7.75	GLY145, MET147,	LEU146,	PHE160, LEU263,

		GLU163, GLY164, ARG252, PHE254.	ARG266.	LEU267.
<b>28-Norcastasterone</b>	-8.69	MET147, PHE160, GLU163, GLU167, THR250, LEU251, LEU260, ARG266.	GLY145	LEU146, ARG252, PHE254, LEU263.
<b>24-Epicastasterone</b>	-8.39	MET147, PHE160, GLU163, GLU167, THR250, LEU260, ARG266.	GLY145	LEU146, ARG252, PHE254, LEU263, LEU267.
<b>3,24-Diepicastasterone</b>	-6.78	GLY145, GLU163, GLY, 164, GLU167, ARG252, LEU260, ARG264, ARG266.	PHE160	LEU146, PHE254, LEU263, LEU267.
<b>6<math>\alpha</math>-Hydroxycastasterone</b>	-9.16	MET147, PHE160, GLU163, THR220, LEU251, LEU260, ARG266.	GLY145, GLU167.	LEU146, ARG252, PHE254, LEU263, LEU267.
<b>6-Deoxocastasterone</b>	-8.97	GLU163, GLY164, GLU167, LEU251, ARG266, LEU267.	GLY145, ARG252.	LEU146, ARG160, PHE254, LEU263.
<b>6-Deoxo-28-norcastasterone</b>	-8.68	MET147, PHE160, GLU163, GLU167, THR250, LEU251, LEU260, ARG266.	GLY145.	LEU146, ARG252, PHE254, LEU263, LEU267.

Table.4. Brassinosteroids and aromatase enzyme interaction

Aromatase (5JKV )	G-Score kcal/mol	Van der waals	Hydrogen bond	Alkyl/ $\pi$ -Alkyl
<b>Brassinolide</b>	-12.51	ILE132, TRP141, ARG145, ALA306, ASP309, THR310, LEU372, VAL373, ARG435, GLY439, SER478.	ARG115, LEU477	ILE133, PHE134, PHE221, TRP224, VAL370, MET374, CYS437, ALA438.
<b>28-Homobrassinolide</b>	-11.54	PHE221, TRP224, ALA307, THR310, MET311, VAL370, VAL373, ALA443, ILE442, MET446, LEU477, SER478.	ARG115, LEU372, MET374, ALA438, GLY439	-
<b>24-Epibrassinolide</b>	-12.26	ILE132, ILE133, TRP141, ARG145, ALA306, ASP309, THR310, VAL370, LEU372, VAL373, ARG435, CYS437, ALA438, GLY439, SER478.	ARG115, LEU477	PHE134, PHE221, TRP224, MET374.
<b>Castasterone</b>	-12.44	ILE132, LEU152, PHE221, GLU302, THR310, VAL373, GLY439, SER478.	LEU372, MET374, CYS437.	ILE133, PHE134, PHE148, TRP224, ALA306, MET303, VAL370, ALA438,

				LEU477.
<b>28-Homocastasterone</b>	-12.36	ARG115, LEU151, PHE221, GLU302, MET303, THR310, VAL373, LEU477.	LEU372, MET374, GLY439.	ILE132, PHE134, TRP224, VAL370, ALA438.
<b>28-Norbrassinolide</b>	-12.25	ILE132, TRP141, ARG145, PHE221, ALA306, THR310, VAL373, SER478, ARG435, ALA438, GLY439.	ARG115, LEU372, LEU477	ILE133, PHE134, TRP224, MET374, CYS437.
<b>28-Norcastasterone</b>	-10.23	ARG115, ILE132, PHE134, LEU152, PHE221, ALA307, THR310, MET443, MET446, LEU477, SER478.	GLY439	ILE133, PHE148, TRP224, MET303, VAL370, ALA438.
<b>24-Epicastasterone</b>	-10.99	ARG115, ILE132, PHE134, ARG145, ASP309, LEU372, MET374, ALA443, SER478.	CYS437, ALA438, GLY439, LEU477.	ILE133, PHE221, TRP224, MET303, VAL370, ALA438.
<b>3,24-Diepicastasterone</b>	-11.86	ILE132, TRP141, ARG145, ALA306, ASP310, VAL370, LEU372, VAL373, ARG435, GLY436, SER478.	ARG115, LEU477.	ILE133, PHE134, PHE221, TRP224, MET374, CYS437, ALA438.
<b>6<math>\alpha</math>-Hydroxycastasterone</b>	-10.24	ARG115, ILE132, PHE221, ALA307, THR310, MET311, VAL373, CYS437, GLY439, LEU477, SER478.	-	ILE133, PHE148, LEU152, TRP224, MET303, ALA306, VAL370, ALA438, ILE442, ALA443, MET446.
<b>6-Deoxocastasterone</b>	-10.53	ARG115, ILE133, PHE221, MET311, MET364, MET374, PRO429, MET447.	THR310, LEU372.	PHE134, TRP224, VAL370, VAL373, PHE430, CYS437, ALA443, LEU477.
<b>6-Deoxo-28-norcastasterone</b>	-11.10	ARG115, TRP224, ASP309, THR310, VAL369, VAL370, LEU372, MET374, GLY439, SER478.	CYS437, ALA438, LEU477.	ILE133, PHE134, PHE221, ALA306, ALA438.

Table 5. Brassinosteroids and Androgen receptor amino acid residues interaction

Androgen receptor (1E3G)	G-Score kcal/mol	Van der waals	Hydrogen bond	Alkyl/ $\pi$ -Alkyl
<b>Brassinolide</b>	-5.65	LEU701, ASN705, LEU707, GLY708, TRP741, MET745, ARG752, PHE764, LEU768, GLN783, MET787, THR877,	VAL746	LEU704, MET742, MET780, LEU873, PHE876, MET895

		LEU880, ILE899.	PHE891,		
<b>28-Homobrassinolide</b>	-4.12	LEU701, LEU707, TRP741, MET749, GLN783, MET787, PHE891, ILE899.	ASN705, GLY708, MET745, ARG752, PHE764, LEU880,	VAL746, THR877	LEU704, MET780, MET895. MET742, PHE876,
<b>28-Homocastasterone</b>	-4.42	LEU701, GLY708, TRP741, PHE764, THR877, ILE899	LEU707, GLN711, MET749, MET787,	ASN705, VAL746, ARG752.	LEU704, MET745, LEU873, MET895. MET742, MET780, PHE876,
<b>28-Norbrassinolide</b>	-5.05	LEU701, LEU707, MET745, ARG752, MET787, LEU880, PHE891.	LEU704, GLY708, MET749, LEU768, THR877,	ASN705, VAL746.	MET742, LEU873, MET895. MET780, PHE876,
<b>28-Norcastasterone</b>	-5.89	GLU681, VAL685, LEU744, TRP751, ASN756, LYS808.	GLY683, HIS714, MET745, THR755, PRO766,	GLN711	PRO682, VAL715, ARG752 VAL684, ALA748,
<b>24-Epicastasterone</b>	-4.25	GLY708, LEU880, PHE891, MET895.	TRP741, VAL889,	LEU701, ASN705, GLN711, PHE764, THR877.	LEU704, MET742, VAL746, MET780, LEU823, PHE876. LEU707, MET745, MET749, MET787,
<b>3,24-Diepicastasterone</b>	-4.88	LEU701, GLY708, ALA748, PHE764, THR877, PHE891.	LEU707, TRP741, MET749, MET787, LEU880,	ASN705, GLN711, VAL746, ARG752.	LEU704, MET745, LEU873, MET895. MET742, MET780, PHE876,
<b>6<math>\alpha</math> Hydroxycastasterone</b>	-3.09	LEU701, GLY708, TRP741, THR877, PHR891, ILE899.	LEU707, GLN711, ARG752, LEU880,	ASN705	LEU705, MET745, MET749, MET780, LEU873, MET895. MET742, VAL746, PHE764, MET787, PHE876,
<b>6-Deoxocastasterone</b>	- 6.66	LEU701, LEU707, MET742, GLN783, LEU880, ILE899.	ASN705, TRP741, ALA748, THR877, PHE891,	GLN711, ARG752, PHE762.	LEU704, VAL746, MET780, LEU873, PHE876, MET895. MET745, MET749, MET787, MET780,
<b>6-Deoxo-28-norcastasterone</b>	-6.54	LEU701, ASN705, GLY708, ARG752, THR877, PHE891.	LEU704, LEU707, TRP741, GLN783, LEU880,	-	MET742, VAL746, PHE764, MET787, LEU873, PHE876, MET895. MET745, MET749, MET780,

**Table 6. Brassinosteroids and Estrogen receptor amino acid residues interaction**

Estrogen receptor (3ERT)	G-Score kcal/mol	Van der waals	Hydrogen bond	Alkyl/ $\pi$ -Alkyl
<b>Brassinolide</b>	-12.96	THR347, LEU384, LEU387, MET388, LEU391, MET421, PHE422, LEU428, VAL534.	ASP351, ASN532.	LEU346, ALA350, LEU354, TRP383, PHE404, LEU525, MET528, PRO535.
<b>28-Homobrassinolide</b>	-9.56	LEU349, LEU391, ARG394, PHE404, MET421, ILE424, PHE425, LEU428, ASN532, VAL534.	THR347, ASP351, GLU353.	MET343, LEU346, ALA350, LEU354, TRP383, LEU387, LEU525, MET528, PRO535.
<b>24-Epibrassinolide</b>	-9.63	MET343, LEU346, THR347, LEU387, LEU391, PHE404, MET421, ILE424, PHE425, LEU428, HIS524, VAL534.	ASP351	ALA350, LEU354, LEU525, VAL533, PRO535.
<b>Castasterone</b>	-13.33	THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, PHE425, LEU428, VAL534.	ASP351, ASN532.	LEU346, ALA350, LEU354, PHE404, LEU525, MET528, VAL533, PRO535.
<b>28-Homocastasterone</b>	-10.66	MET343, GLU353, TRP383, LEU384, MET388, ARG394.	THR347	LEU346, LEU349, ALA350, LEU387, LEU391, PHE404, LEU428, LEU525.
<b>28-Norbrassinolide</b>	-10.19	MET343, THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, PHE425, LEU428, VAL534.	ASP351, ASN532.	LEU346, ALA350, LEU354, TRP383, PHE404, LEU525, MET528, VAL533, PRO535.
<b>28-Norcastasterone</b>	-10.98	ASP351, GLU353, LEU354, LEU384, MET388, LEU391, ARG394, PHE404, PHE425, LEU428, MET528, ASN532, VAL533, VAL534, PRO535.	THR347.	MET343, LEU346, ALA350, TRP383, LEU387, MET421, LEU525.
<b>24-Epicastasterone</b>	-12.93	THR347, LEU384, LEU387, MET388, LEU391, MET421, ILE424, LEU428, VAL534.	ASP351, ASN532.	LEU346, ALA350, LEU354, PHE404, LEU525, MET528, VAL533, PRO535.
<b>3,24-Diepicastasterone</b>	-11.99	THR347, GLU353, LEU354, LEU391, ARG394, PHE404, PHE425, LEU428, ASN532, VAL533, VAL534, THR347,	ASP351.	MET343, LEU346, ALA350, TRP383, LEU384, LEU387, MET388, MET421, LEU525, PRO535.

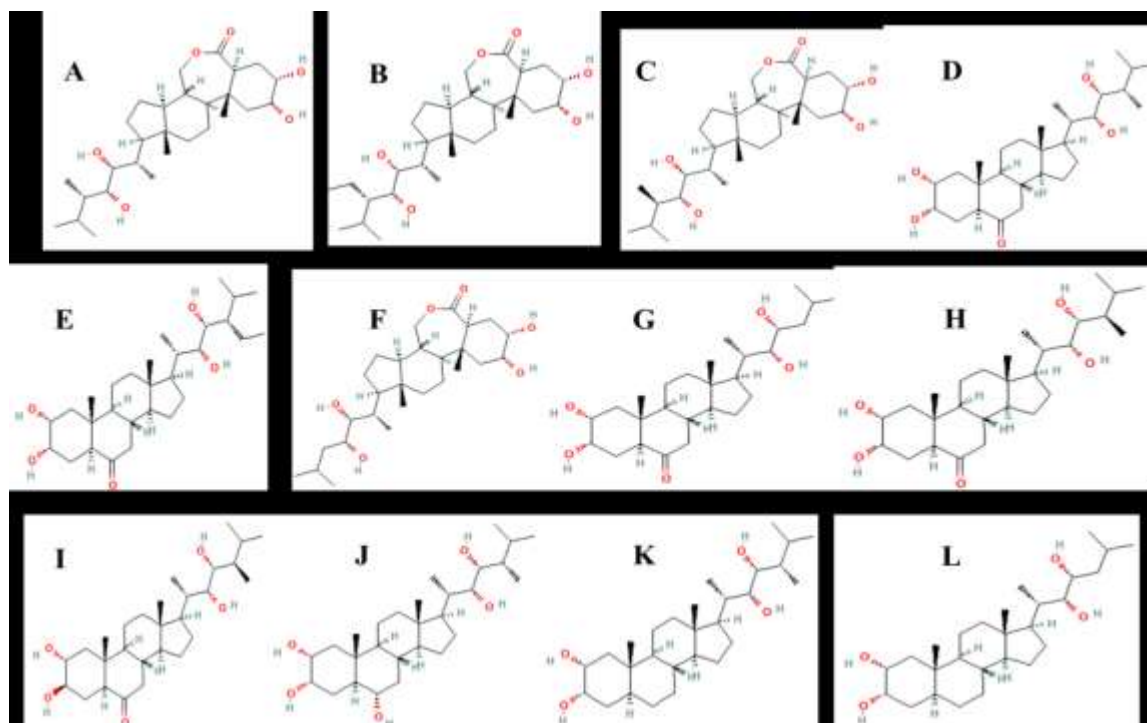
		ARG394, PHE404, MET421, PHE425, LEU428, VAL428, VAL534.		
<b>6<math>\alpha</math></b> Hydroxycastasterone	-10.66	LEU346, THR347, LEU349, ASP351, GLU353, LEU354, MET421, ILE424, PHE425, LEU428, GLY521, VAL534.	ASN532	ALA350, TRP383, LEU384, LEU387, MET388, LEU391, PHE404, VAL533, PRO535.
<b>6-Deoxocastasterone</b>	-12.78	MET343, THR347, ASP351, GLU353, TRP383, LEU384, MET388, ARG394, MET421, PHE425, ASN532, PRO535.	-	LEU346, LEU349, ALA350, LEU387, LEU391, PHE404, LEU525, VAL533.
<b>6-Deoxo-28-norcastasterone</b>	-11.36	LEU346, LEU347, GLU353, TRP383, MET388, ARG394, PHE404, PRO535.	-	MET343, ALA350, LEU384, LEU387, LEU391, MET421, ILE424, PHE425, LEU428, LEU525, VAL533.

Table 7. Standard ligands and steroidogenesis proteins amino acid residues interaction

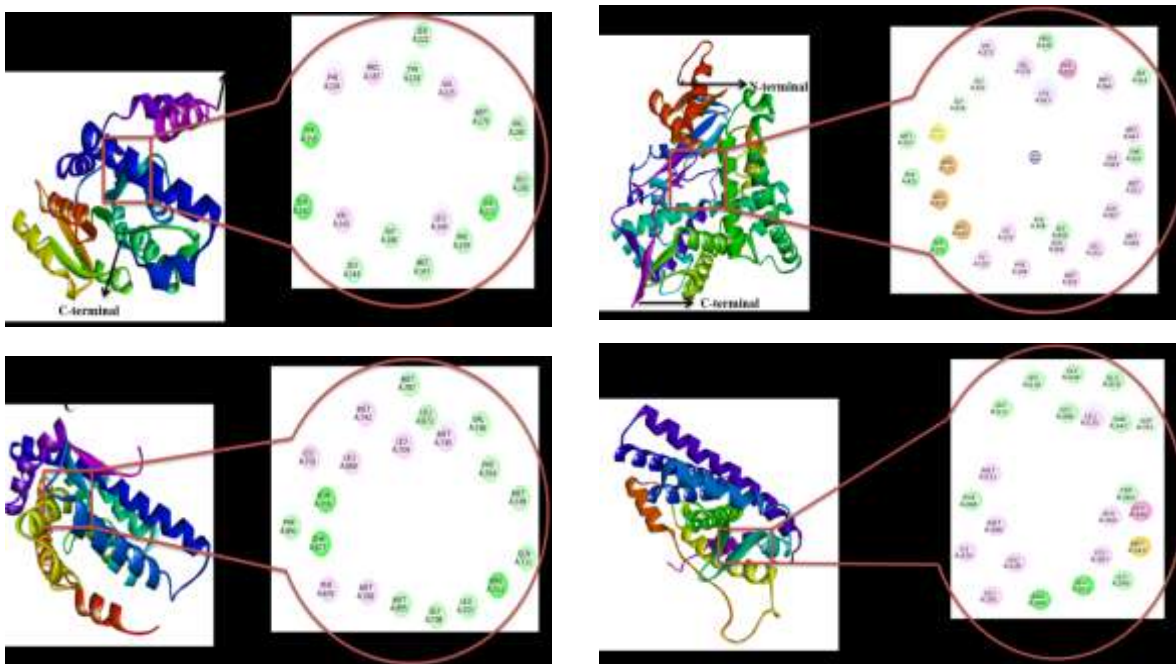
Proteins	Compounds	G-Score kcal/mol	Van der waals	Hydrogen bond	Alkyl/ $\pi$ -Alkyl	$\pi$ -sigma
<b>17<math>\beta</math></b> hydroxysteroid dehydrogenase (1FDS)	<b>Abiraterone</b>	-7.68	GLY145, GLU163, GLY164, ARG252, PHE254, ARG264, ARG266.	PHE160	LEU146, LEU260, LEU267.	LEU263
	<b>Androstenedione</b>	-7.04	GLY145, PHE160, GLU163, GLU167, ARG252, PHE254, ARG266.	GLY164	LEU146, LEU263, LEU267.	-
<b>Aromatase (5JKV)</b>	<b>Letrozole</b>	-7.93	ILE132, ILE133, ARG145, LEU152, MET303, ALA306, VAL373, PHE430, GLY431, GLY439	ARGY115, TRP141	ARG435	CYS437
	<b>Anastrozole</b>	-9.6	SER314, THR310, MET364, PRO368,	MET311	PHE430, VAL370	-

			VAL369, PRO429, CYS437, ALA443.			
	<b>Testosterone</b>	-9.85	PHE134, PHE221, ILE305, ASP309, THR310, ILE133, LEU372, VAL373, LEU477, SER478.	ARG 115, ALA 306, MET 374.	TRP224, VAL370	-
<b>Androgen receptor (1E3G)</b>	<b>Flutamide</b>	-0.83	THR739, MET742, GLY743, LEU744, LEU812, VAL866.	SER740, SER814, GLN867.	LEU811, ILE815, ALA870,ARG 871 HIS874, ILE906, PRO913.	-
	<b>Testosterone</b>	-1.11	GLN711, ALA748, MET780, LEU873, PHE876, THR877.	MET745	LEU704, LEU707, MET742, VAL746, MET749, PHE764.	-
<b>Estrogen receptor (3ERT)</b>	<b>Tamoxifen</b>	-11.03	GLU353, LEU354, TRP383, LEU384, MET388, MET421, ILE424, PHE425, GLY521, HIS524, VAL533, PRO535.	ASP351, VAL534	LEU346, LEU349, ALA350, LEU387, PHE404, LEU525	LEU525
	<b>Estradiol</b>	-9.69	LEU346, LEU349, MET421, LEU428, HIS524.	GLU353, ARG394, GLY521.	ALA350, LEU384, LEU387, MET388, LEU391, ILE424, LEU525.	PHE404

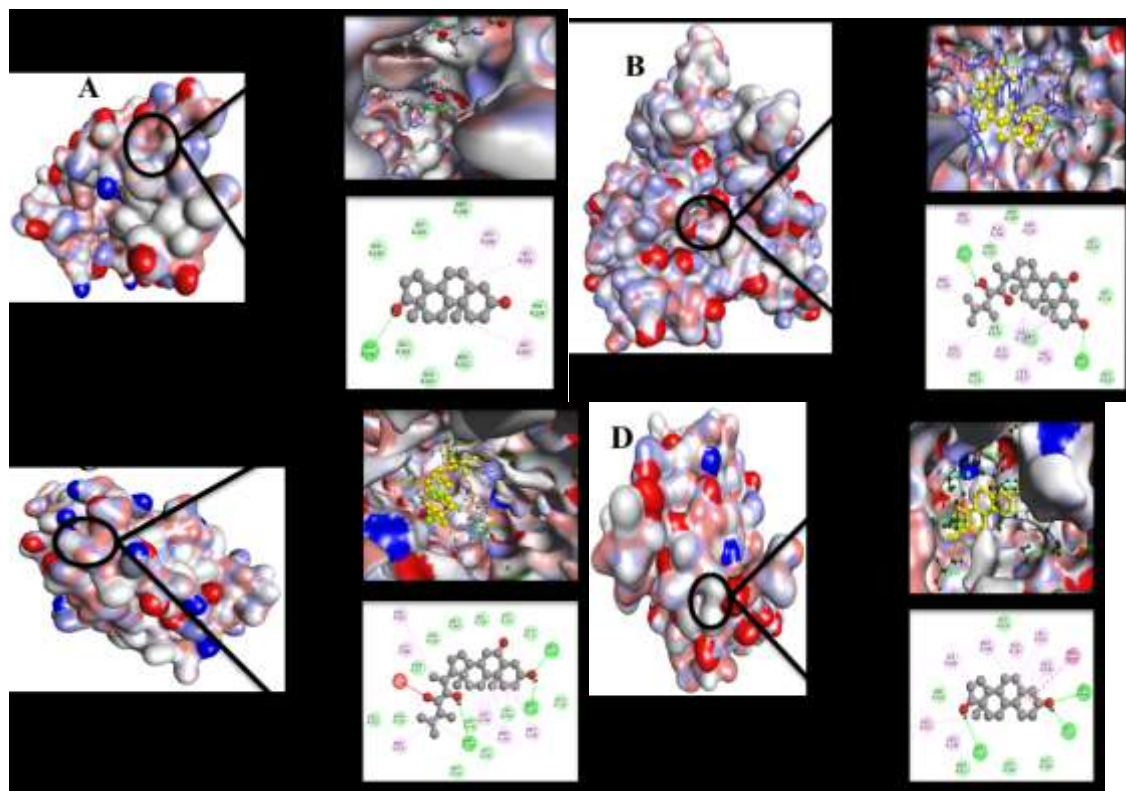




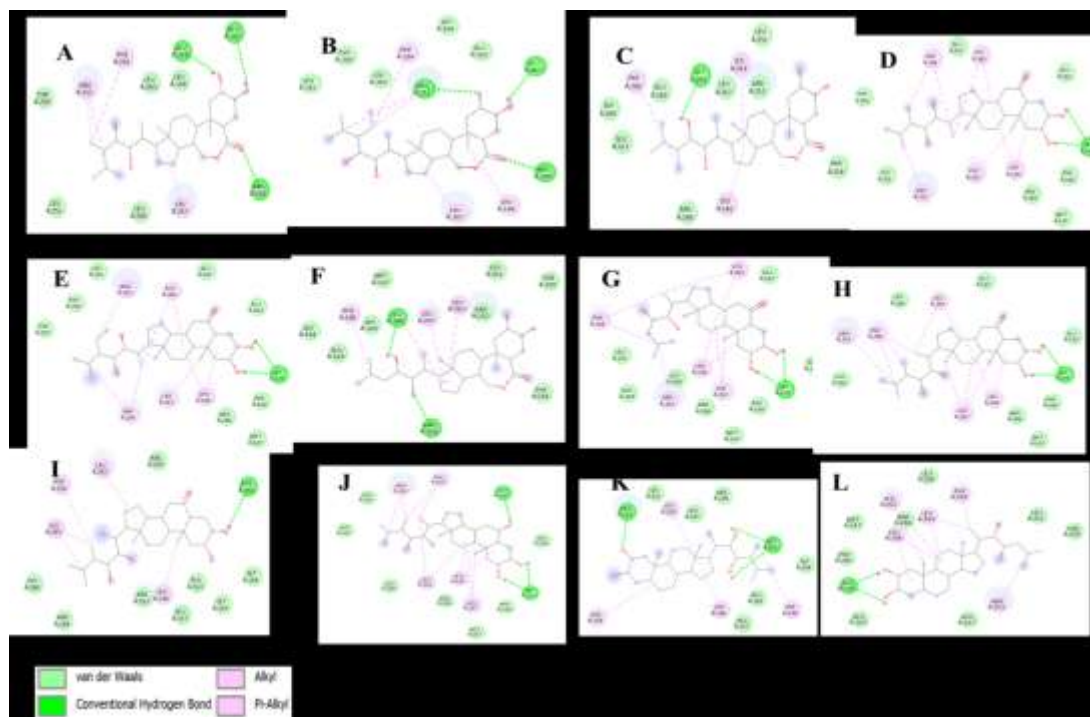
**Figure 1.** Shows 2D structure of ligands A. Brassinolide, B. 28-Homobrassinolide, C. 24-Epibrassinolide, D. Castasterone, E. 28-Homocastasterone, F. 28-Norbrassinolide, G. 28-Norcastasterone, H. 24-Epicasterone, I. 3,24-Diepicasterone, J. 6 $\alpha$ -Hydroxycastasterone, K. 6-Deoxocastasterone, L. 6-Deoxo-28-norcastasterone.



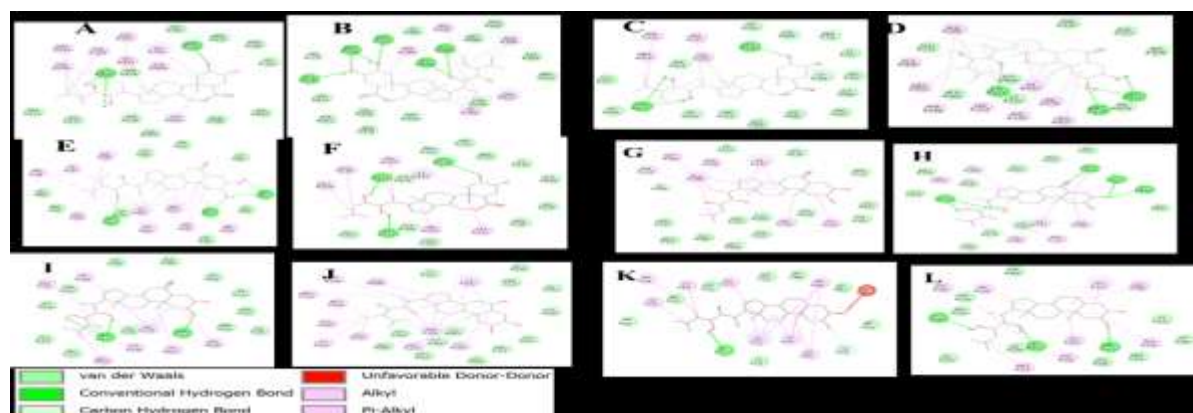
**Figure 2.** Shows the ligand binding side and the amino acid residues interacting ligand, A. 17 $\beta$ HSD enzyme, B. Aromatase enzyme, C. Androgen receptor, D. Estrogen receptor.



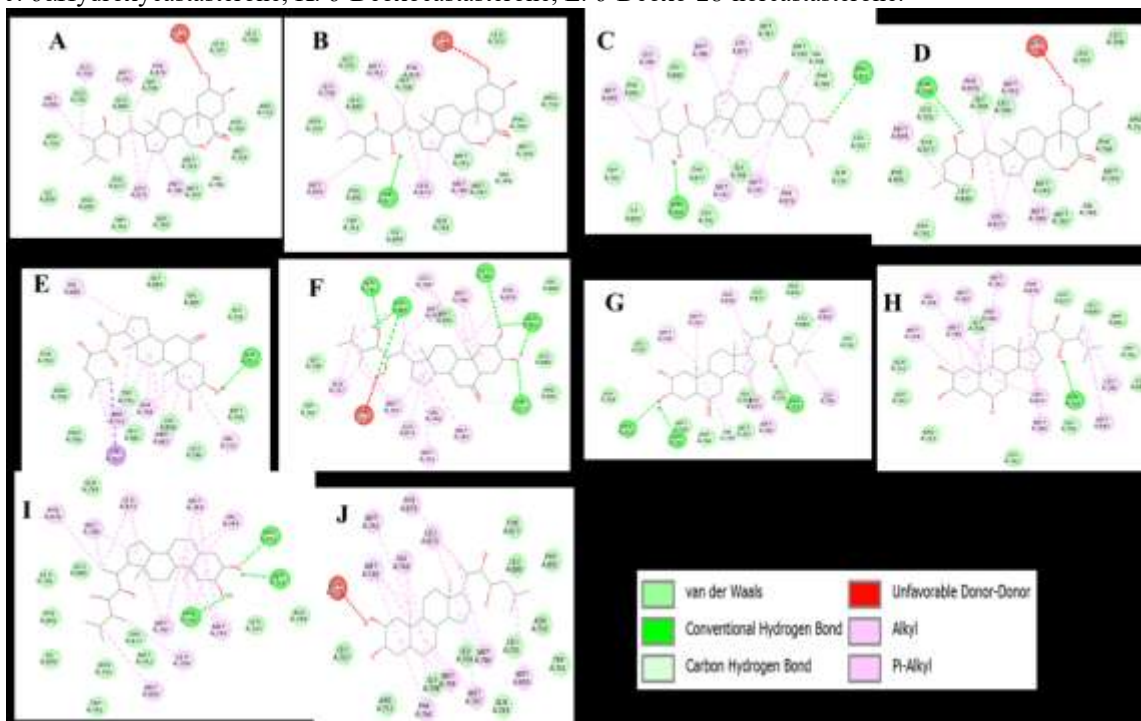
**Figure 3.** Shows the 2D and 3D interaction of A. 17 $\beta$ HSD enzyme and Androstenedione, B. Aromatase enzyme and testosterone, C. Androgen receptor and testosterone, D. Estrogen receptor and estradiol.



**Figure 4.** Shows A. Brassinolide, B. 28-Homobrassinolide, C. 24-Epibrassinolide, D. Castasterone, E. 28-Homocastasterone, F. 28-Norbrassinolide, G. 28-Norcastasterone, H. 24-Epicastasterone, I. 3,24-Diepicastasterone, J. 6 $\alpha$ Hydroxycastasterone, K. 6-Deoxocastasterone, L. 6-Deoxo-28-norcastasterone.

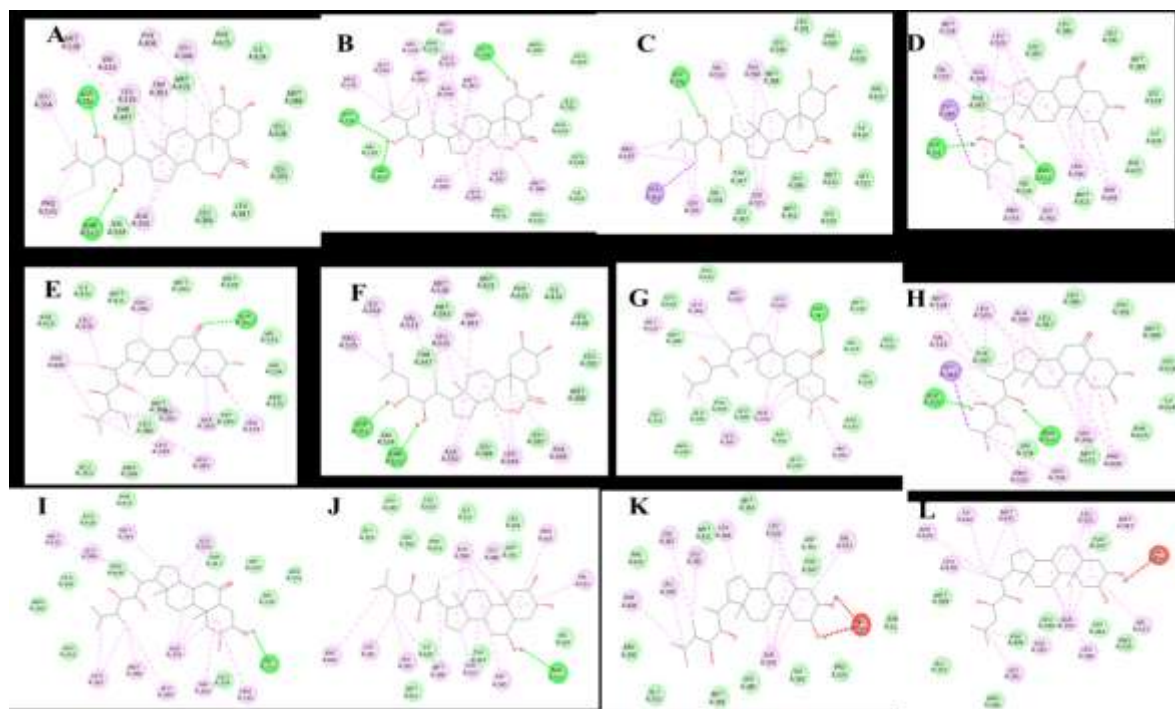


**Figure 5.** Shows A. Brassinolide, B. 28-Homobrassinolide, C. 24-Epibrassinolide, D. Castasterone, E. 28-Homocastasterone, F. 28-Norbrassinolide, G. 28-Norcastasterone, H. 24-Epicastasterone, I. 3,24-Diepicastasterone, J. 6 $\alpha$ Hydroxycastasterone, K. 6-Deoxocastasterone, L. 6-Deoxo-28-norcastasterone.

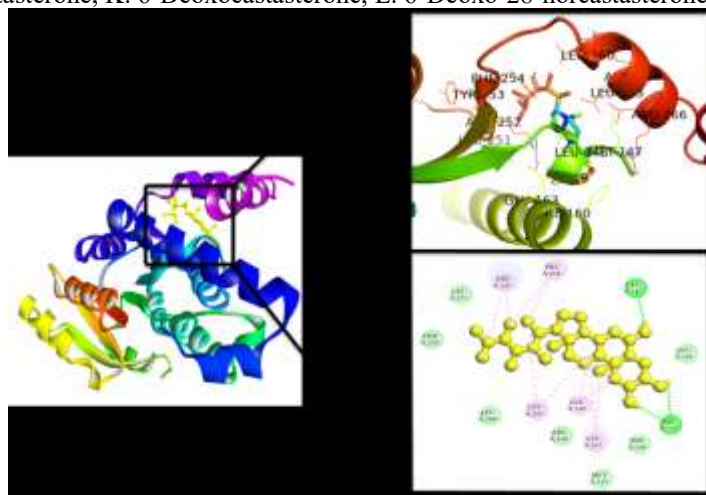


**Figure 6.** Shows A. Brassinolide, B. 28-Homobrassinolide, C. 28-Homocastasterone, D. 28-Norbrassinolide, E. 28-Norcastasterone, F. 24-Epicastasterone, G. 3,24-Diepicastasterone, H. 6 $\alpha$ Hydroxycastasterone, I. 6-Deoxocastasterone, J. 6-Deoxo-28-norcastasterone.

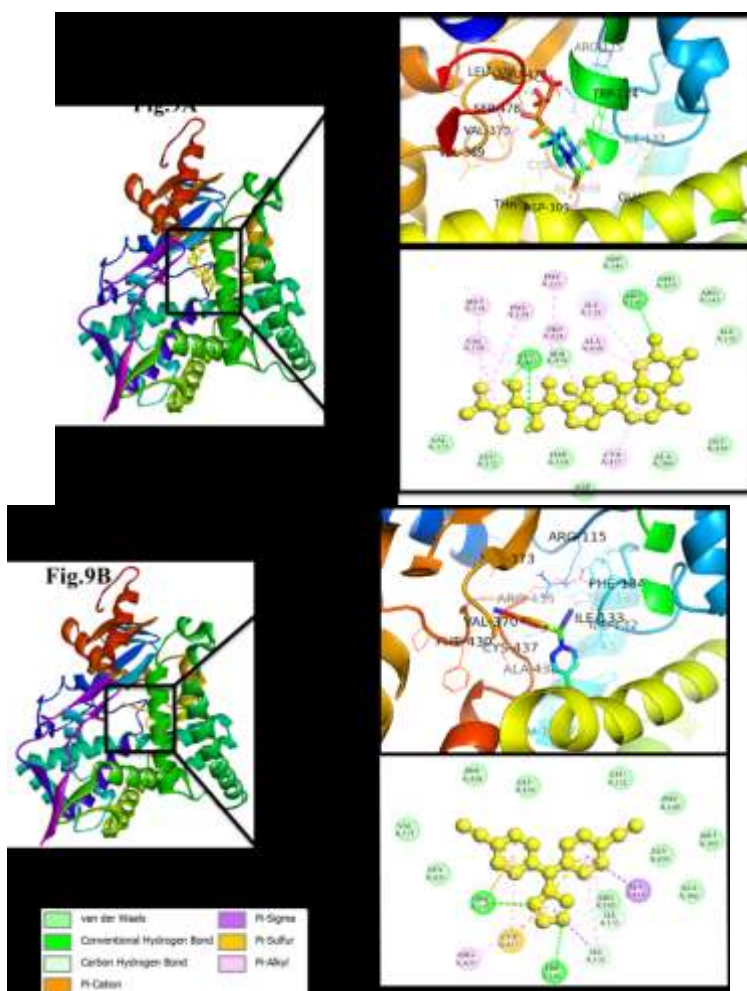




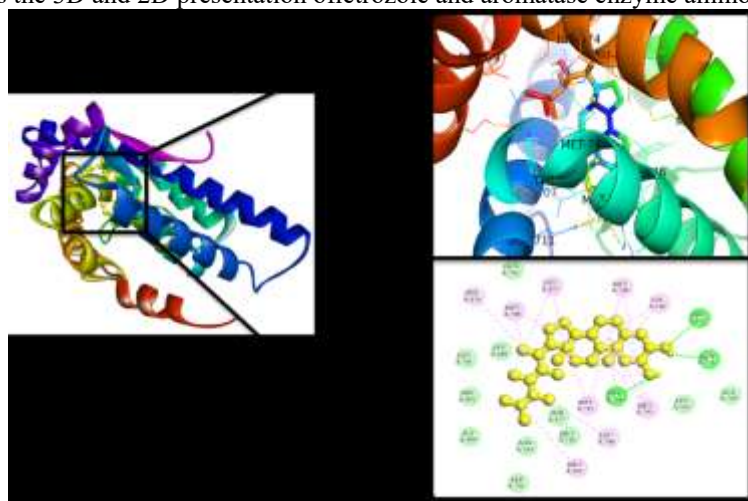
**Figure 7.**Shows A. Brassinolide, B. 28-Homobrassinolide, C. 24-Epibrassinolide, D. Castasterone, E. 28-Homocastasterone, F. 28-Norbrassinolide, G. 28-Norcastasterone, H. 24-Epicastasterone, I. 3,24-Diepicastasterone, J. 6 $\alpha$ -Hydroxycastasterone, K. 6-Deoxocastasterone, L. 6-Deoxy-28-norcastasterone.

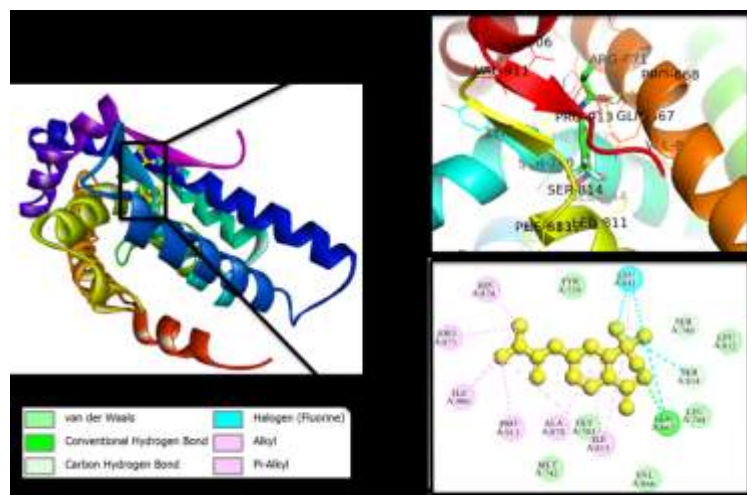


**Figure 8A.**Showsthe3D and 2D presentation of abiraterone and 17 $\beta$ HSD enzyme amino acid interactions, **Figure.8B.**Shows the 3D and 2D presentation of 6 $\alpha$ -Hydroxycastasterone and 17 $\beta$ HSD enzyme amino acid interactions.

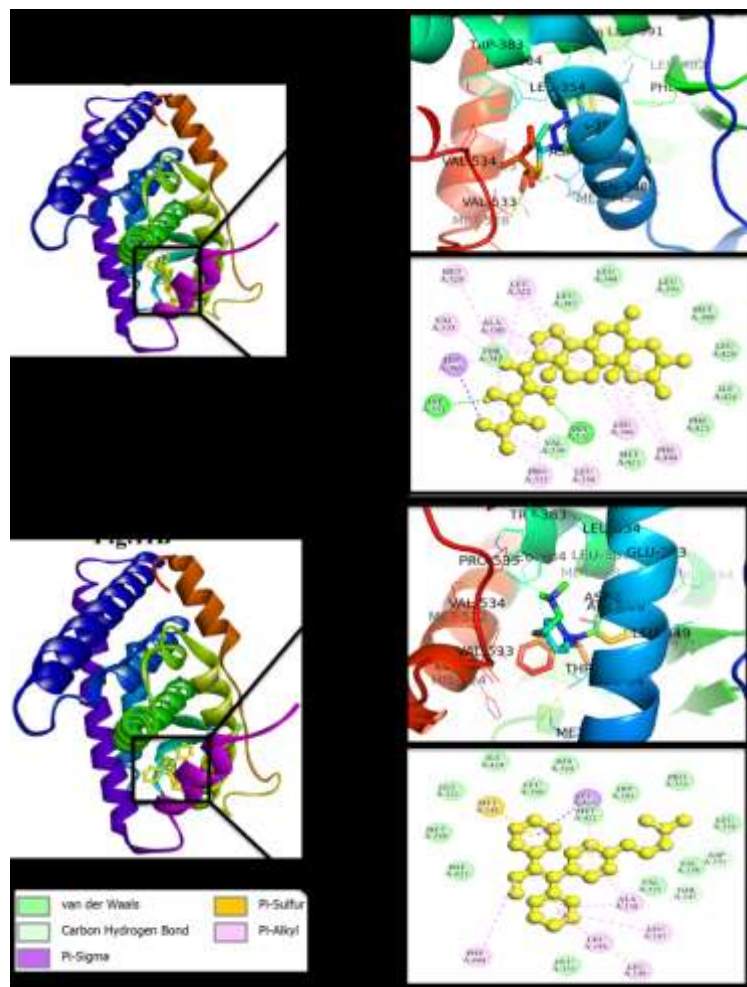


**Figure 9A.**Shows the 3D and 2D presentation of brassinolide and aromatase enzyme amino acid interactions,  
**Figure.9B.**Shows the 3D and 2D presentation of ofletrozole and aromatase enzyme amino acid interactions.





**Figure 10A.**Shows the 3D and 2D presentation of 6-Deoxocastasterone and androgen receptor amino acid interactions, **Figure.10B.**Shows the 3D and 2D presentation of Flutamide and androgen receptor amino acid interactions.



**Figure 11A.**Shows the 3D and 2D presentation of castasterone and androgen receptor amino acid interactions, **Figure.11B.**Shows the 3D and 2D presentation of tamoxifen and androgen receptor amino acid interactions.

## Conclusion:-

Present In Silico study identified 6 $\alpha$ -Hydroxycastasterone, castasterone, 28-Homocastasterone, 6-Deoxocastasterone, 24-Epibrassinolide, 6-Deoxo-28-norcastasterone, brassinolide, 28-Norbrassinolide, 3,24-Diepicastasterone, and 28-Homobrassinolide as potential modulator of ovarian steroidogenesis through 17 $\beta$ HSD, aromatase, androgen and estrogen receptors inhibition. Among them 6-Deoxo-28-norcastasterone, castasterone, 24-Epibrassinolide, 6-Deoxocastasterone, 3,24-Diepicastasterone, 28-Norbrassinolide and brassinolide are exhibited superior putative ovarian steroidogenesis inhibitor. These results suggest a novel phytomolecule based managing PCOS complications. Remarkably, brassinosteroids shown better binding avidity with 17 $\beta$ HSD, aromatase, androgen and estrogen receptors than conventional drug molecules.

In summary, BRs are now known that to inhibit 17 $\beta$ HSD, aromatase enzymes and androgen and estrogen receptors transactivation in PCOS, it is established that BRs shows structural similarity with human oxysterol function. The outcome of our In Silico data may be the basis for In Vivo and In Vitro studies against PCOS with phyto molecule brassinosteroids.

**Conflicts of interest:** Declare that there are no conflicts of interest, whatsoever, among themselves.

**Animal and human ethics clearance:** Not applicable

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