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### RESEARCH ARTICLE

## NATURAL IMMUNOMODULATORS AS ALTERNATIVES TO ANTIMICROBIALS: EVIDENCE FROM ZEBRAFISH MODELS

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#### Abstract

Antimicrobial resistance is one of the emerging issues of global concern. The declining efficacy of conventional antimicrobial treatment highlights the urgent need for novel interventions. Natural alternatives, specifically immunomodulators, present a promising solution by empowering the host's inherent immune responses rather than directly targeting the pathogens. This approach offers a distinct advantage, introducing new mechanisms of action to which pathogens have yet to develop resistance. Oxidative stress and inflammation are strongly linked to each other, where each can exacerbate the other. Therefore, developing drugs that can decrease the production of these molecules presents a viable treatment strategy for microbial infections. In this context, immunomodulation emerges as a key player. By enhancing the host's broad defensive capabilities against viral, bacterial, and fungal pathogens, immunomodulation can lead to a reduction in reactive oxygen species (ROS), thereby offering valuable therapeutic solutions. Natural compounds derived from plants, animals, and microbes are increasingly being explored for their immunomodulatory properties. This mini-review summarises current research on their potential to mitigate the harmful effects of infections in the host. We aim to present promising, environmentally friendly, and sustainable alternatives to traditional antimicrobial therapies, thereby supporting a healthy environment. Using zebrafish models, which are widely preferred due to their genetic similarity to humans and well-characterised immune system, we highlight key natural immunomodulators, emphasizing their mechanisms, applications, and potential as sustainable substitutes.

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#### Introduction: -

Microorganisms are available naturally in the surrounding environment; hence, it is obvious that they can easily create infections in almost every group of vertebrates. An antimicrobial agent is a substance that destroys or inhibits the growth of microorganisms, including bacteria, fungi, viruses, and parasites. These agents are widely used in medicine and healthcare to prevent and treat infectious diseases by targeting harmful microbes and reducing their

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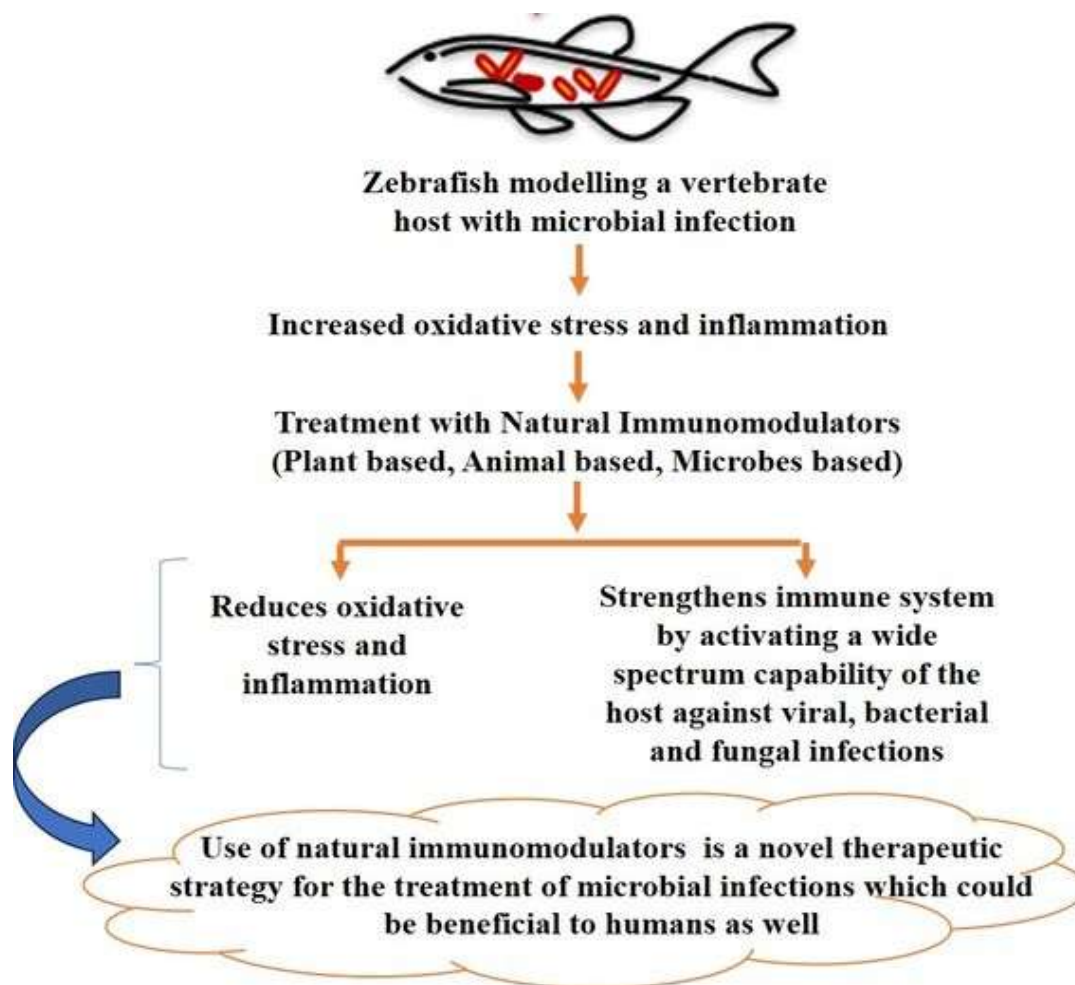
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ability to survive and multiply in the body (Madigan et al., 2018). For decades, antimicrobial drugs have been the primary strategy for controlling infectious diseases. However, the excessive and improper use of antibiotics in human healthcare, veterinary practice, and animal husbandry has accelerated the emergence of antimicrobial resistance (AMR), thereby reducing the effectiveness of conventional antimicrobial therapies (O'Neill, 2016; Holmes et al., 2016; Chiş et al., 2022). In addition, many synthetic antimicrobial agents are associated with high costs and adverse effects on the host. These challenges have prompted growing interest in alternative and sustainable therapeutic approaches.

Oxidative stress is the imbalance between reactive oxygen species (ROS) production and antioxidant defenses during immune responses. Excessive oxidative stress can damage immune cells and tissues, leading to inflammation and immune dysfunction. Inflammation is the protective response of the body against injury, infection, or harmful stimuli to eliminate the cause and start healing. It is characterized by redness, heat, swelling, pain, and sometimes loss of function (Halliwell and Gutteridge, 2015). Oxidative stress and inflammation are two important and well-studied pathological hallmarks of any infectious disease. These are two coexisting phenomena that influence each other (Tejchman et al., 2021). The onset of oxidative stress induces the overproduction of reactive oxygen species that can lead to inflammation (Biswas, 2016). Prolonged inflammation may further enhance ROS generation, resulting in tissue damage. Therefore, therapeutic strategies capable of regulating oxidative stress and inflammatory responses may help control infection-associated pathologies more effectively.

In this context, immunomodulation has emerged as a promising approach for combating infectious diseases. Unlike conventional antimicrobial agents that directly target pathogens, immunomodulators enhance or regulate the host immune response against a broad spectrum of viral, bacterial, and fungal infections (Pirofski and Casadevall, 2006). This provides a broad range of acute care strategies in case of emerging pathogens or biowarfare agents (Mahmoudi and Baradaran, 2024). Natural immunomodulators are naturally occurring substances that regulate or modify the activity of the immune system by either enhancing or suppressing immune responses. They help maintain immune homeostasis, improve host defense mechanisms, and reduce excessive inflammatory reactions, making them important in the prevention and management of various diseases (Patwardhan and Gautam, 2005). Natural bioactive compounds derived from plants, animals, and microorganisms have gained considerable attention due to their antioxidant, anti-inflammatory, and immunomodulatory properties. These natural compounds may provide sustainable alternatives to traditional antimicrobial therapies while reducing the risk of antimicrobial resistance.

Zebrafish (*Danio rerio*) is a small freshwater fish native to South Asia, especially India, Bangladesh, Nepal, and Pakistan. It is called “zebrafish” because of the dark horizontal stripes on its body that resemble the stripes of a zebra (Lawrence, 2007). Zebrafish is widely used as a model organism in immunology because it has rapid development, transparent embryos, easy genetic manipulation, and an immune system that shares many similarities with humans, making it useful for studying immunity, inflammation, and diseases. Zebrafish has emerged as a valuable experimental model for studying immune responses and evaluating natural immunomodulators because of its genetic similarity to humans, transparent embryonic stages, and well-characterised immune system (Howe et al., 2013). In this mini-review, we summarise recent evidence regarding plant-, animal-, and microbe-derived natural immunomodulators investigated in zebrafish models as illustrated in Figure 1 with particular emphasis on their ability to counteract infection-induced oxidative stress and inflammation, enhance host immunity, and serve as potential alternatives to conventional antimicrobial therapies.



**Figure 1.** Conceptual framework illustrating the role of natural immunomodulators as therapeutic alternatives in zebrafish models of microbial infection. Zebrafish (*Danio rerio*), serving as a vertebrate host model, respond to microbial infection with increased oxidative stress and inflammation. Treatment with plant-, animal-, and microbe-derived natural immunomodulators counteracts these responses by reducing oxidative stress and inflammation while simultaneously strengthening broad-spectrum host immunity against viral, bacterial, and fungal pathogens. Together, these dual outcomes support the use of natural immunomodulators as a novel and potentially human-relevant therapeutic strategy for the management of microbial infections.

### I] Plant-Based Natural Immunomodulators

Plant-derived immunomodulators are natural compounds obtained from diverse botanicals that can regulate, enhance, or suppress immune responses, offering therapeutic potential against inflammation, infections, autoimmune disorders, and cancer. Their effects are mediated by various phytochemicals, including polysaccharides, flavonoids, alkaloids, and terpenoids. Studies using zebrafish models have demonstrated that several plant-based natural compounds and commercial products enhance survival against bacterial infections and modulate important inflammatory pathways. The following plant-based commercial products further emphasise the immunomodulatory potential of natural compounds for therapeutic applications.

#### **Immusante® (IM-133N): -**

Immusante®, a patent product of the Himalaya Drug Company, Bengaluru, India, consists of a combination of aqueous extracts of *Symplocos racemosa* and *Prosopis glandulosa*. Venkata et al. (2021) reported that Immusante® reduces the

upregulated levels of pro-inflammatory cytokine genes - IL1 $\beta$ , IFN $\gamma$ , and TNF $\alpha$  in zebrafish kidney infected by *A. hydrophila*. In addition, it did not alter the levels of anti-inflammatory cytokine IL-4, suggesting that it may help in reducing the inflammation without affecting the overall immune response.

#### **BMSL-cN16E:-**

BMSL-cN16E, a combination of Butea monosperma seed lectin (BMSL) and N-palmitoylethanolamine-derived cationic lipid (cN16E), proved to be effective against *Escherichia coli* infection in zebrafish (Subramaniyan et al., 2024). Zebrafish treated with BMSL-cN16E showed significant upregulation in the expression of certain immune genes - TNF $\alpha$ , IFN $\gamma$ , IL-1 $\beta$ , IL -4, IL -10, TLR-2, etc., as compared to those of untreated fish or the fish treated singly with BMSL or cN16E. The results revealed that BMSL-cN16E not only rescued infected zebrafish but also conferred long-lasting protection in terms of immunomodulation that could be protective against multiple reinfections.

#### **Rice Husk Silica (RHS):-**

RHS is shown to improve the innate immune response of zebrafish. The fish infected with *Aeromonas hydrophila* and *Streptococcus iniae* and then treated with different concentrations of rice husk silica showed a significant increase in the expression levels of certain genes - IL1 $\beta$ , IL6, IL15, TNF $\alpha$ , COX2a, TLR4a, lysozyme, and complement C3b (Hong et al., 2019). This finding indicates that RHS can be used as an immunostimulant to improve the health of the organism.

#### **Orange Juice extract (OJe):-**

OJe is reported to have a protective role against inflammation if it is properly consumed in the diet. Cirimi et al. (2021) found that prior treatment with OJe led to reduced intestinal inflammation and decreased expression of inflammatory genes such as IL1 $\beta$ , IL6, and TNF $\alpha$  in zebrafish infected with *Vibrio anguillarum*.

#### **Astragalus Polysaccharides (APS):-**

*Astragalus membranaceus*, a traditional medicinal herb, has demonstrated immunomodulatory effects in zebrafish models, primarily through its polysaccharides (APS). Studies have shown that APS can enhance innate immunity by upregulating immune-related genes such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$ , while also improving resistance to bacterial infections like *Aeromonas hydrophila*. Additionally, *Astragalus* modulates inflammatory responses via the NF- $\kappa$ B pathway and exhibits antioxidant properties, contributing to reduced oxidative stress. At suitable doses, it is well-tolerated in zebrafish, making it a promising natural immunostimulant for biomedical and aquaculture applications (Li et al, 2021; Zhang et al., 2019).

#### **Ginger (*Zingiber officinale*):-**

Ginger has demonstrated immunomodulatory effects in zebrafish models, attributed to its active constituents such as gingerols and shogaols. Studies show that ginger enhances the expression of key immune cytokines (IL-1 $\beta$ , TNF- $\alpha$ , IFN- $\gamma$ ), boosts resistance to pathogens like *Aeromonas hydrophila*, and modulates inflammation via the NF- $\kappa$ B pathway. Additionally, its antioxidant properties reduce oxidative stress and support immune homeostasis. Ginger is well-tolerated in zebrafish embryos, highlighting its potential as a natural immunostimulant in health and aquaculture applications (Rahman et al., 2020).

#### **Curcumin:-**

Curcumin exhibits significant immunomodulatory effects in zebrafish models by downregulating pro-inflammatory cytokines such as IL-1 $\beta$  and TNF- $\alpha$ , enhancing antioxidant enzyme activity, and modulating immune gene expression. It protects against bacterial infections and oxidative stress while maintaining a favourable safety profile at appropriate doses. These findings support curcumin's potential as a natural immunotherapeutic agent in aquatic and biomedical research (Cui et al., 2016; Zou et al., 2020).

### **II] Animal-Based Natural Immunomodulators**

Beyond well-known plant-derived compounds, various animal-based natural immunomodulators are gaining attention for their ability to regulate host immune responses. For instance, beta-glucans from yeast and fungi (often associated with animal products like probiotics), polysaccharides and peptides from marine organisms, and components derived from colostrum and milk are offering valuable natural alternatives for immune system modulation. The following are some of the commercial products that have been studied using zebrafish as the animal model.

**Phosvitin-Derived Peptide (Pt5):-**

Pt5 is a peptide derived from zebrafish egg yolk protein phosvitin. It has an affinity with different microbial signature molecules, lipopolysaccharide, lipoteichoic acid and peptidoglycan and hence can display a remarkable antimicrobial activity (Ding et al., 2012). They reported that in the spleen and kidneys of zebrafish infected with *A. hydrophila*, Pt5 suppresses the production of proinflammatory cytokine genes (IL1 $\beta$ , IL6, TNF $\alpha$  and IFN $\gamma$ ) while upregulating the expression of anti-inflammatory cytokine genes (IL10 and IL4), thereby preventing *Aeromonas hydrophila* infection in zebrafish.

**NK-lysin A:-**

NK-lysin A, an antimicrobial peptide derived from natural killer cells, has demonstrated immunomodulatory and antimicrobial effects in zebrafish. It enhances the expression of immune cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$ , reduces bacterial burden during infections with *Aeromonas hydrophila*, and improves survival rates. NK-lysin A also regulates inflammatory signalling pathways like NF- $\kappa$ B, promoting immune balance without excessive inflammation. Importantly, it is well tolerated in zebrafish, suggesting potential for therapeutic applications in infectious diseases and immune modulation (Wang et al., 2022; Liu et al., 2023).

**Chitosan:-**

Chitosan, generated from chitin through deacetylation, is a well-known animal-based antibacterial agent. Chitosan, a natural polysaccharide derived from chitin, exhibits significant immunomodulatory effects in zebrafish models. It enhances innate immune responses by upregulating cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$ , and increases resistance to bacterial infections like *Aeromonas hydrophila*. Chitosan also modulates inflammatory signalling through NF- $\kappa$ B and reduces oxidative stress by boosting antioxidant enzyme activity. Importantly, it is biocompatible and well-tolerated in zebrafish embryos, making it a promising agent for immune enhancement in both medical and aquaculture contexts (Liu et al., 2020; Zhang et al., 2021).

**Black Soldier Fly Larvae (BSFL) derived products:-**

Black Soldier Fly Larvae (BSFL) derived products, including protein hydrolysates and lipid extracts, exhibit immunomodulatory effects in zebrafish models. These products enhance innate immunity by upregulating cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$ , improve resistance to bacterial infections like *Aeromonas hydrophila*, and modulate inflammatory and antioxidant pathways such as NF- $\kappa$ B signalling. Additionally, BSFL-derived compounds positively influence gut microbiota and are well tolerated during zebrafish development, highlighting their potential as sustainable natural immunostimulants in aquaculture and biomedical research (Martinez et al., 2022; Singh et al., 2023).

**Silkworm (*Bombyx mori*) Polysaccharides:-**

Polysaccharides derived from silkworm (*Bombyx mori*) demonstrate immunomodulatory effects in zebrafish by upregulating cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$  and enhancing antioxidant enzyme activities. These compounds improve zebrafish resistance to bacterial infections, modulate inflammatory pathways including NF- $\kappa$ B, and reduce oxidative stress. Importantly, silkworm polysaccharides are well-tolerated in zebrafish embryos, suggesting their potential as safe natural immunostimulants in aquaculture and biomedical research (Li et al., 2021; Zhang et al., 2023).

**Sp-LECin:-**

Sp-LECin, a synthetic peptide derived from *Scylla paramamosain*, demonstrates potent immunomodulatory and antimicrobial effects in zebrafish models. It enhances the expression of cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$ , improves survival rates against bacterial infections, including *Aeromonas hydrophila*, and modulates inflammatory signalling pathways like NF- $\kappa$ B. Furthermore, Sp-LECin is well tolerated in zebrafish embryos, supporting its potential as a novel immunostimulant in aquatic health and biomedical research (Chen et al., 2021; Li et al., 2022).

**III] Microbes-Based Natural Immunomodulators**

Microbes (including bacteria, fungi, and yeasts) are an incredibly rich and diverse source of natural immunomodulators, offering a sustainable and eco-friendly avenue for enhancing human health. These microbes produce specific metabolites that regulate cytokine production, resulting in their anti-inflammatory activities. Researchers are keenly focused on these microbial-derived compounds, continuously developing new products that leverage their ability to interact with and regulate the immune system. Here are some examples of microbe-based immunomodulators that have been studied using the zebrafish model, highlighting its potential in preclinical screening:

**Probiotics:-**

Zebrafish were treated orally with *Bacillus coagulans* and then infected with *Vibrio vulnificus*. The expression of immune-related genes was assessed 30 days post-infection, and the researchers found a significant increase in the expression of TLR4, TNF $\alpha$ , TRAM1, and NF $\kappa$ B (Pan et al., 2012). In another experiment performed by Wang et al. (2016), probiotic treatment using *Bacillus coagulans* 9-712 and *Lactobacillus plantarum* 08-923 significantly affected the cytokine expression in zebrafish. TNF $\alpha$  and IL10 levels were significantly increased, and IL1 $\beta$  levels decreased over time following infection, suggesting that these bacterial strains have immunoregulatory and protective properties, and they could effectively stimulate the regeneration of the mucosal barrier and promote an anti-inflammatory response in zebrafish. Yi et al. (2019) evaluated the immune-relevant targets in zebrafish infected with *Aeromonas hydrophila* and *Streptococcus iniae*. The fish was pretreated with *Chromobacterium aquaticum*, which resulted in increasing the levels of IL1 $\beta$ , IL6, TNF $\alpha$ , IL10, IL21, NF $\kappa$ B, lysozyme, and complement C3b in fish. This modulated the innate immunity of zebrafish against infection with *A. hydrophila* and *S. iniae*. Li et al. (2022) investigated the effect of *Lactobacillus plantarum* WCFS1, which was found to prevent inflammation and muscle atrophy in zebrafish infected with *Aeromonas hydrophila* NJ-1. Ehsannia et al. (2022) studied the immunomodulatory effects of two probiotic strains on zebrafish. The fish were infected with *A. hydrophila* and then treated with *L. bulgaricus*, *L. acidophilus*, and a combination of both probiotics. The untreated group had the greatest levels of TNF $\alpha$  and IL1 $\beta$  expression, and the groups that received *Lactobacillus bulgaricus* treatment solely had the best survival rate.

**Heat-killed *Listeria monocytogenes*:-**

Heat-killed *Listeria monocytogenes* was used to induce immune responses in adult zebrafish that led to increased clearance of mycobacterial infection in them (Luukinen et al., 2017). The response induces TNF $\alpha$  and NOS2b, and downregulates SOD2, likely leading to increased production of radical nitrogen and oxygen species and enhanced intracellular killing of mycobacteria.

**CM11:-**

CM11, a short antimicrobial peptide, was used against *Streptococcus iniae* and *Yersinia ruckeri* infection in zebrafish. The AMP significantly improved the antioxidant and immune responses in the fish, revealed by the upregulation of TNF- $\alpha$ , Lys, IL-1 $\beta$ , IL-8, SOD, and CAT (Rashidian et al., 2021).

**Microalga:-**

In a study by Nayak et al. (2018), fish were fed diets containing two different strains of microalgae for one month prior to the infection with *Streptococcus iniae*. The researchers found that the diet containing the mutant strain (P127) led to significantly higher levels of catalase and glutathione peroxidase, enzymes potentially contributing to the improved survival observed in these fish following bacterial challenge.

**Aeromonas Immune Modulator A (AimA):-**

Aeromonas Immune Modulator A (AimA), a secreted protein from *Aeromonas* species, functions as an immunomodulator in zebrafish by dampening excessive inflammatory responses. It reduces pro-inflammatory cytokines such as IL-1 $\beta$  and TNF- $\alpha$ , regulates NF- $\kappa$ B signalling, and promotes host tolerance to infection, resulting in improved survival without necessarily clearing the pathogen. AimA is well tolerated in zebrafish embryos, highlighting its role as a modulator of immune homeostasis during the bacterial challenge (Rolig et al., 2018).

To facilitate comparison across all three categories discussed in this review, Table 1 provides a consolidated summary of the natural immunomodulators investigated in zebrafish models, highlighting their origin, the pathogen models employed, key immune genes and pathways modulated, and relevant references.

Table 1. Summary of natural immunomodulators investigated in zebrafish (*Danio rerio*) models, categorised by their origin

Category	Immunomodulator	Source	Key Immunomodulatory Effects in Zebrafish	Target Pathogens/ Applications	Key References
Plant-based	Immusante®(IM-133N)	Symplocos racemosa + Prosopis glandulosa extracts	Reduced IL1 $\beta$ , IFN $\gamma$ , TNF $\alpha$ ; maintained IL-4 levels	Aeromonas hydrophila infection	Venkata et al., 2021
Plant-based	BMSL-cN16E	Butea monosperma seed lectin + cationic lipid	Upregulated TNF $\alpha$ , IFN $\gamma$ , IL-1 $\beta$ , IL-4, IL-10, TLR-2	Escherichia coli infection	Subramanian et al., 2024
Plant-based	Rice Husk Silica (RHS)	Rice husk-derived silica	Increased IL1 $\beta$ , IL6, IL15, TNF $\alpha$ , COX2a, TLR4a, lysozyme, C3b	Aeromonas hydrophila, Streptococcus iniae	Hong et al., 2019
Plant-based	Orange Juice extract (OJe)	Citrus flavonoid extract	Reduced intestinal inflammation and IL1 $\beta$ , IL6, TNF $\alpha$ expression	Vibrio anguillarum infection	Cirmi et al., 2021
Plant-based	Astragalus Polysaccharides (APS)	Astragalus membranaceus	Enhanced IL-1 $\beta$ , TNF- $\alpha$ , IFN- $\gamma$ ; modulated NF- $\kappa$ B; antioxidant activity	Aeromonas hydrophila resistance	Liet al., 2021; Zhang et al., 2019
Plant-based	Ginger (Zingiber officinale)	Gingerols and shogaols	Enhanced cytokine expression; reduced oxidative stress; modulated NF- $\kappa$ B	Aeromonas hydrophila infection	Rahman et al., 2020
Plant-based	Curcumin	Curcuminoid from turmeric	Downregulated IL-1 $\beta$ and TNF- $\alpha$ ; enhanced antioxidant enzymes	Oxidative stress and bacterial infections	Cui et al., 2016; Zhou et al., 2020
Animal-based	Pt5 peptide	Zebrafish egg yolk phosphovitin	Suppressed IL1 $\beta$ , IL6, TNF $\alpha$ , IFN $\gamma$ ; increased IL10 and IL4	Aeromonas hydrophila infection	Ding et al., 2012
Animal-based	NK-lysin A	Natural killer cell peptide	Enhanced IL-1 $\beta$ , TNF- $\alpha$ , IFN- $\gamma$ ; regulated NF- $\kappa$ B	Aeromonas hydrophila infection	Wan et al., 2022; Liu et al., 2023
Animal-based	Chitosan	Chitin-derived polysaccharide	Enhanced cytokine expression; antioxidant and NF- $\kappa$ B modulation	Aeromonas hydrophila resistance	Liu et al., 2020; Zhang et al., 2021
Animal-based	Black Soldier Fly Larva eproducts	Protein hydrolysates / lipid extracts	Increased IL-1 $\beta$ , TNF- $\alpha$ , IFN- $\gamma$ ; improved gut microbiota	Bacterial infection resistance	Martinez et al., 2022; Singh et al., 2023
Animal-based	Silkworm polysaccharides	Bombyx mori polysaccharides	Enhanced cytokines and antioxidant enzymes; modulated NF- $\kappa$ B	Bacterial infection resistance	Liet al., 2021; Zhang et al., 2023

			F-κB		
<b>Animal-based</b>	Sp-LECIn	PeptidefromScylla paramamosain	EnhancedIL-1β, TNF-α, IFN-γ; improved survival; NF-κB modulation	Aeromonashydrophila infection	Chen et al., 2021; Liet al., 2022
<b>Microbe-based</b>	Probiotics	Bacillus, Lactobacillus, Chromobacterium spp.	Modulated TNFα, IL10, IL1β, NFκB, lysozyme, complement C3b	Vibriovulnificus, Aeromonashydrophila, S.iniae	Pan et al., 2012; Wang et al., 2016; Yi et al., 2019
<b>Microbe-based</b>	Heat-killed <i>Listeria monocytogenes</i>	Inactivated bacteria	Increased TNFα and iNOS2b; enhance radical species production	Mycobacterial infection	Luukinen et al., 2017
<b>Microbe-based</b>	CM11 peptide	Antimicrobial peptide	Upregulated TNF-α, Lys, IL-1β, IL-8, SOD, CAT	<i>Streptococcus iniae</i> , <i>Yersinia ruckeri</i>	Rashidian et al., 2021
<b>Microbe-based</b>	Microalga	ω6 LC-PUFA-rich algae	Increased catalase and glutathione peroxidase	<i>Streptococcus iniae</i> infection	Nayak et al., 2018
<b>Microbe-based</b>	<i>Aeromonas</i> Immune Modulator A (AimA)	<i>Aeromonas</i> secreted protein	Reduced IL-1β and TNF-α; regulated NF-κB; promoted tolerance	Bacterial infection tolerance	Rolig et al., 2018

### Conclusion: -

This mini review highlights the potential of natural immunomodulators as a promising avenue for developing sustainable and environmentally friendly antimicrobial therapies. Natural immunomodulators, derived from plants, animals, and microbes, offer a promising alternative to conventional antimicrobial therapies. Zebrafish-based studies provide valuable insights into the therapeutic potential of natural immunomodulators for developing safer, environmentally friendly, and sustainable strategies for infection management. Future research should focus on specific mechanisms of action of these natural immunomodulators, optimising their delivery and efficacy, and rigorously assessing their safety profiles. In addition, advances in genome editing, omics technologies, and high-throughput screening using the zebrafish (*Danio rerio*) model may further accelerate the discovery and development of novel natural immunotherapeutic agents for biomedical and aquaculture applications.

### Ethical Declaration: -

This article is a review based on previously published studies and does not involve any new experiments on human participants or animals performed by the authors. Therefore, ethical approval and informed consent were not required.

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### Conflict of Interest: -

The authors declare that they have no conflict of interest.

**References: -**

1. Biswas, S. K. (2016). Does the Interdependence between Oxidative Stress and Inflammation Explain the Antioxidant Paradox? *Oxidative Medicine and Cellular Longevity*, 2016, 1–9. <https://doi.org/10.1155/2016/569893>
2. Chen, Y., Wang, S., & Zhang, X. (2021). Immunomodulatory effects of Sp-LECin peptide in zebrafish (*Danio rerio*): Activation of innate immunity and antimicrobial activity. *Fish & Shellfish Immunology*, 109, 101–109. <https://doi.org/10.1016/j.fsi.2020.12.012>,
3. Chiş, A. A., Rus, L. L., Morgovan, C., Arseniu, A. M., Frum, A., Vonica-Țincu, A. L., Gligor, F. G., Mureşan, M. L., & Dobrea, C. M. (2022). Microbial Resistance to Antibiotics and Effective Antibiotherapy. *Biomedicines*, 10(5), 1121. Cirmi, S., Randazzo, B., Russo, C., Musumeci, L., Maugeri, A., Montalbano, G., Guerrero, M. C., Lombardo, G. E., & Levanti, M. (2020). Anti-inflammatory effect of a flavonoid-rich extract of orange juice in adult zebrafish subjected to *Vibrio anguillarum*-induced enteritis. *Natural Product Research*, 35(23), 5350–5353. <https://doi.org/10.1080/14786419.2020.1758096>
4. Cirmi, S., Randazzo, B., Russo, C., Musumeci, L., Maugeri, A., Montalbano, G., ... & Levanti, M. (2021). Anti-inflammatory effect of a flavonoid-rich extract of orange juice in adult zebrafish subjected to *Vibrio anguillarum*-induced enteritis. *Natural Product Research*, 35(23), 5350-5353.
5. Cui, C., Zhang, L., & Wang, Y. (2016). Curcumin suppresses inflammation and oxidative stress in zebrafish exposed to bacterial endotoxin. *Fish & Shellfish Immunology*, 55, 1–8. <https://doi.org/10.1016/j.fsi.2016.05.005>,
6. Ding, Y., Liu, X., Bu, L., Li, H., & Zhang, S. (2012). Antimicrobial-immunomodulatory activities of zebrafish phosvitin-derived peptide Pt5. *Peptides*, 37(2), 309–313. <https://doi.org/10.1016/j.peptides.2012.07.014>
7. Ehsannia, S., Ahari, H., Kakoolaki, S., Anvar, S. A., & Yousefi, S. (2022). Effects of probiotics on Zebrafish model infected with *Aeromonas hydrophila*: spatial distribution, antimicrobial, and histopathological investigation. *BMC Microbiology*, 22(1). <https://doi.org/10.1186/s12866-022-02491-4>
8. Halliwell, B. and Gutteridge, J.M., (2015). *Free radicals in biology and medicine*. Oxford university press.
9. Holmes, A. H., Moore, L. S., Sundsfjord, A., Steinbakk, M., Regmi, S., Karkey, A., ... & Piddock, L. J. (2016). Understanding the mechanisms and drivers of antimicrobial resistance. *The Lancet*, 387(10014), 176-187.
10. Hong, Y. H., Tseng, C. C., Setyoningrum, D., Yang, Z. P., Maftuch, N., & Hu, S. Y. (2019). Rice Husk Silica Enhances Innate Immune in Zebrafish (*Danio rerio*) and Improves Resistance to *Aeromonas hydrophila* and *Streptococcus iniae* Infection. *Sustainability*, 11(22), 6504. <https://doi.org/10.3390/su11226504>
11. Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Collins, J. E., Humphray, S., McLaren, K., Matthews, L., McLaren, S., Sealy, I., Caccamo, M., Churcher, C., Scott, C., Barrett, J. C., Koch, R., Rauch, G., White, S., . . . Stemple, D. L. (2013). The zebrafish reference genome sequence and its relationship to the human genome. *Nature*, 496(7446), 498–503. <https://doi.org/10.1038/nature12111>
12. Lawrence, C., (2007). The husbandry of zebrafish (*Danio rerio*): A review. *Aquaculture*, 269(1-4), pp.1-20. <https://doi.org/10.1016/j.aquaculture.2007.04.077>
13. Li, H., Chen, W., & Zhao, Y. (2021). Immunomodulatory effects of silkworm-derived polysaccharides in zebrafish (*Danio rerio*). *Fish & Shellfish Immunology*, 112, 135–143. <https://doi.org/10.1016/j.fsi.2021.02.015> ,
14. Li, J., Xu, P., & Liu, Q. (2022). Sp-LECin modulates NF- $\kappa$ B signalling and enhances antibacterial defence in zebrafish challenged with *Aeromonas hydrophila*. *Developmental & Comparative Immunology*, 131, 104350. <https://doi.org/10.1016/j.dci.2022.104350>.
15. Li, S., Guo, L., Si, X., Dai, Z., Zhou, Z., & Wu, Z. (2022). *Lactobacillus plantarum* WCFS1 alleviates *Aeromonas hydrophila* NJ-1-induced inflammation and muscle loss in zebrafish (*Danio rerio*). *Aquaculture*, 548, 737603. <https://doi.org/10.1016/j.aquaculture.2021.737603>
16. Liu, Y., Wang, Q., & Chen, X. (2020). Chitosan induces immune response and enhances the resistance of zebrafish (*Danio rerio*) to *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 101, 309–317. <https://doi.org/10.1016/j.fsi.2020.03.042>,
17. Liu, H., Zhang, L., & Xu, D. (2023). NK-lysin A modulates NF- $\kappa$ B signalling and enhances antibacterial defence in zebrafish. *Developmental & Comparative Immunology*, 136, 104555. <https://doi.org/10.1016/j.dci.2022.104555>.
18. Luukinen, H., Hammarén, M. M., Vanha-Aho, L. M., Svorjova, A., Kantanen, L., Järvinen, S., Luukinen, B. V., Dufour, E., Rämetsä, M., Hytönen, V. P., & Parikka, M. (2017a). Priming of innate antimycobacterial immunity by heat-killed *Listeria monocytogenes* induces sterilizing response in the adult zebrafish tuberculosis model. *Disease Models & Mechanisms*. <https://doi.org/10.1242/dmm.031658>
19. Madigan, M.T., Bender, K.S., Buckley, D.H., Sattley, W.M. and Stahl, D.A., (2018). *Microbial growth and its control*. Brock Biology of Microorganisms, 15th edn. Pearson Education, New York, pp.173-208.

20. Mahmoudi, F., & Baradaran, B. (2024). Microbial secondary metabolites as immunomodulators. *J MicrobBiochemTechnol*, Vol.16(3), 1000615.
21. Martinez, R. A., Gómez, R., & Silva, J. (2022). Immunomodulatory effects of Black Soldier Fly Larvae protein hydrolysates in zebrafish (*Danio rerio*). *Fish & Shellfish Immunology*, 125, 289–297. <https://doi.org/10.1016/j.fsi.2022.05.017> ,
22. Nayak, S., Khozin-Goldberg, I., Cohen, G., & Zilberg, D. (2018). Dietary Supplementation With  $\omega$ 6 LC-PUFA-Rich Algae Modulates Zebrafish Immune Function and Improves Resistance to Streptococcal Infection. *Frontiers in Immunology*, 9. <https://doi.org/10.3389/fimmu.2018.01960>
23. O'Neill, J. (2016). Tackling drug-resistant infections globally: final report and recommendations.
24. Patwardhan, B. and Gautam, M. (2005). Botanical immunodrugs: scope and opportunities. *Drug discovery today*, 10(7), pp.495-502. [https://doi.org/10.1016/S1359-6446\(04\)03357-4](https://doi.org/10.1016/S1359-6446(04)03357-4)
25. Pan, C. Y., Wang, Y. D., & Chen, J. Y. (2012a). Immunomodulatory effects of dietary *Bacillus coagulans* in grouper (*Epinephelus coioides*) and zebrafish (*Danio rerio*) infected with *Vibrio vulnificus*. *Aquaculture International*, 21(5), 1155–1168. <https://doi.org/10.1007/s10499-012-9619-0>
26. Pirofski, L., & Casadevall, A. (2006). Immunomodulators as an antimicrobial tool. *Current Opinion in Microbiology*, 9(5), 489–495. <https://doi.org/10.1016/j.mib.2006.08.004>
27. Rahman, M. M., Lee, K. Y., & Kim, Y. S. (2020). Immunostimulatory and anti-inflammatory effects of ginger extract in zebrafish (*Danio rerio*). *Fish & Shellfish Immunology*, 98, 100–109. <https://doi.org/10.1016/j.fsi.2020.01.014>,
28. Rashidian, G., Moghaddam, M. M., Mirnejad, R., & Azad, Z. M. (2021). Supplementation of zebrafish (*Danio rerio*) diet using a short antimicrobial peptide: Evaluation of growth performance, immunomodulatory function, antioxidant activity, and disease resistance. *Fish & Shellfish Immunology*, 119, 42–50. <https://doi.org/10.1016/j.fsi.2021.09.035>
29. Rolig, A. S., Parthasarathy, R., Burns, A. R., Bohannon, B. J., & Guillemin, K. (2018). The secreted factor AimA from *Aeromonas* modulates host immune responses and promotes tolerance in zebrafish. *Cell Host & Microbe*, 24(2), 179–191.e7. <https://doi.org/10.1016/j.chom.2018.06.001>, Burns, A. R., Rolig, A. S., & Guillemin, K. (2020). Immunomodulation by AimA protein shapes host-microbe interactions in zebrafish. *Trends in Immunology*, 41(7), 544–555. <https://doi.org/10.1016/j.it.2020.04.004>
30. Singh, P., Kumar, S., & Patel, A. (2023). Impact of Black Soldier Fly Larvae lipid extracts on immune response and gut microbiota in zebrafish. *Journal of Insect Science*, 23(2), 45. <https://doi.org/10.1093/jisesa/iead021>.
31. Subramaniam, M. D., Venkatesan, D., Iyer, M., Subbarayan, S., Govindasami, V., Roy, A., Narayanasamy, A., Kamalakannan, S., Gopalakrishnan, A. V., Thangarasu, R., Kumar, N. S., & Vellingiri, B. (2020). Biosurfactants and anti-inflammatory activity: A potential new approach towards COVID-19. *Current Opinion in Environmental Science & Health*, 17, 72–81. <https://doi.org/10.1016/j.coesh.2020.09.002>
32. Tejchman, K., Kotfis, K., & Sienko, J. (2021). Biomarkers and Mechanisms of Oxidative Stress—Last 20 Years of Research with an Emphasis on Kidney Damage and Renal Transplantation. *International Journal of Molecular Sciences*, 22(15), 8010. <https://doi.org/10.3390/ijms22158010>
33. Venkata, A. A., Mohamed, R., Bhavimani, G., Madan, M., Dattatreya, A. S., Azeemuddin, M., & N, M. S. (2021a). Evaluation of immunomodulatory activity of Immusante® in zebrafish. *Journal of Applied Biology and Biotechnology*. <https://doi.org/10.7324/jabb.2021.9310>
34. Wang, Y., Ren, Z., Fu, L., & Su, X. (2016). Two highly adhesive lactic acid bacteria strains are protective in zebrafish infected with *Aeromonas hydrophila* by evocation of gut mucosal immunity. *Journal of Applied Microbiology*, 120(2), 441–451. <https://doi.org/10.1111/jam.13002>
35. Wang, X., Li, Y., & Chen, J. (2022). Immunomodulatory and antimicrobial effects of NK-lysin A in zebrafish (*Danio rerio*). *Fish & Shellfish Immunology*, 121, 258–266. <https://doi.org/10.1016/j.fsi.2021.12.034>,
36. Yi, C. C., Liu, C. H., Chuang, K. P., Chang, Y. T., & Hu, S. Y. (2019). A potential probiotic *Chromobacterium aquaticum* with bacteriocin-like activity enhances the expression of indicator genes associated with nutrient metabolism, growth performance and innate immunity against pathogen infections in zebrafish (*Danio rerio*). *Fish & Shellfish Immunology*, 93, 124–134. <https://doi.org/10.1016/j.fsi.2019.07.042>
37. Zhang, Y., Liu, Q., Wang, Y., Chen, B., & Li, X. (2019). Astragalus polysaccharide enhances immune response and disease resistance in zebrafish (*Danio rerio*) via the NF- $\kappa$ B signalling pathway. *Journal of Ethnopharmacology*, 239, 111921. <https://doi.org/10.1016/j.jep.2019.111921>.
38. Zhang, W., Li, X., & Zhou, Y. (2021). Immunoregulatory and antioxidant activities of chitosan in zebrafish: Role of NF- $\kappa$ B and oxidative stress pathways. *International Journal of Biological Macromolecules*, 180, 683–691. <https://doi.org/10.1016/j.ijbiomac.2021.03.089>.

39. Zhang, J., Wang, Q., & Liu, S. (2023). Silkworm polysaccharides regulate immune responses and oxidative stress in zebrafish: Involvement of NF- $\kappa$ B and MAPK pathways. *International Journal of Biological Macromolecules*, 221, 1349–1358. <https://doi.org/10.1016/j.ijbiomac.2023.01.042>.
40. Zou, W., Wu, H., & Sun, W. (2020). Curcumin protects zebrafish embryos from oxidative and immune damage induced by environmental toxins. *Aquatic Toxicology*, 220, 105398. <https://doi.org/10.1016/j.aquatox.2020.105398>.