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RESEARCH ARTICLE

A study on alteration in Haemato-biochemical parameters in Colibacillosis affected calves

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Abstract

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Mamta Singh Email. mamta2947@gmail.com The objective of the present study was to evaluate haemato-biochemical changes occurred in colibacillosis affected diarrhoeic calves. The study was undertaken in calves at Cattle and Buffalo Farm at I.V.R.I, Izatnagar, during the period from September 2012 to April 2013. A total number of 25 diarrhoeic calves (*Escherichia coli* positive) and five clinically normal healthy calves (as control) of the same herd were taken to investigate the hemato-biochemical profiles. *E. coli* was isolated from the diarrhoeic calves and the animals found positive were used for the study of haemato-biochemical profile. Analysis of blood and serum samples of the diarrhoeic calves revealed significant increase in PCV, total serum protein, serum albumin: globulin (A:G) ratio, serum urea nitrogen, serum albumin, creatinine and potassium, while significant decrease in serum glucose, sodium and chloride were recorded. However, no significant changes were observed in haemoglobin, serum ALT, AST and globulin.

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Introduction

Calf diarrhoea especially occurs in the first week of life (Azizzadeh et al., 2012) and is associated with loss of water and electrolytes that leads to an isotonic or hypotonic dehydration in diarrhoeic calves. Infectious diarrhoea in calves is most commonly associated with enterotoxigenic *Escherichia coli*, *Cryptosporidium parvum*, rotavirus, coronavirus, or some combination of these pathogens (Foster et al., 2009). Among these pathogens, Escherichia coli is the important cause of diarrhoea in neonate. The diarrhoeic calves appeared dull, depressed and lethargic with reduced appetite. The faeces were semisolid to watery with offensive odour, greenish to yellowish white in colour and sometimes even blood stained. Mild to moderate dehydration was found to be a constant feature in diarrhoeic calves. Rectal temperature, pulse and respiration rates were within the normal range (Ramkumar, 2012). The clinicohaemato-biochemical alterations in diarrhoea are complex in nature comprising of serious imbalances of fluid, electrolyte and acid base status threatening the life of the patient.

Materials and Methods

i. Animals

The present study was undertaken in calves at Cattle and Buffalo Farm at I.V.R.I, Izatnagar, in the time period from September 2012 to April 2013. The bovine calves taken for the study belonged to either indigenous Tharparker or Vrindavani (Cross between *Holstein Frisian*, *Brown Swiss* and Haryana) and Murrah Buffulo.

ii. Escherichia coli

E. coli was isolated from the diarrhoeic calves. Those animals found positive for *E. coli* were used for the study of haemato-biochemical profile.

iii.Haematological and Serum Biochemical Profile

Whole blood was collected in EDTA vials for estimation of haemoglobin (Hb), packed cell volume (PCV) and serum for biochemical analysis. The determination of Hb and PCV was done within two hours of collection. However, serum samples were stored at -20°C for further analysis. Haemoglobin concentration (g/dl) in the whole blood was estimated by cyano-methemoglobin method (Vankampen and Zinglstra, 1961). Packed cell volume (PCV %) was determined by capillary micro-haematocrit method (Coles, 1980).

Serum samples stored in deep freeze at -20 ⁰C were used for estimation of the blood urea nitrogen (BUN) by diacetylmonosamine method and serum creatinine by Jaffe's alkaline picrate method (Marsh et al., 1965), serum alanine amino transferase (ALT) and serum aspartate amino transferase (AST) by Reitman and Frankel method (1957), Serum total protein (TP) by modified biuret and albumin by Dumas method (Varley et al., 1980). Serum globulin was estimated by subtracting albumin from total protein and serum albumin: globulin (A:G) ratio was estimated by dividing albumin value with globulin value. Serum glucose and serum chloride were estimated according to O'toludine method (Hultman, 1959) and thiocyanate method (Schoenfeld and Lewellen, 1964) respectively. Also serum sodium and serum potassium were estimated (Trinder, 1951; Miller, 1984).

Results and Discussion

The haemato-biochemical changes recorded in *E. coli* affected diarrhoeic calves (25) and control calves (5) are depicted in table 1. No significant change in haemoglobin values were observed in diarrhoeic calves but highly significant increase was recorded in the value of PCV in all the cases of calf diarrhoea (43.8±0.48%) as compared to the healthy control calves (34.08±0.4). Similarly, present findings are in accordance with those as obtained by Tenant et al.(1968) where they observed a higher value of PCV in scouring calves as compared to normal value (39.03±0.88%). Grove-White and White (1999) also observed a significantly increased value of PCV (48.4 ±10.3 %) in diarrhoeic calves as compared to the non-diarrhoeic calves (33.6 ±4.3 %). Increase in PCV value in diarrhoeic calves was apparently due to hemo-concentration associated with dehydration and hypovolemia.

A significant increase in Total Serum Protein $(7.74\pm0.08g/dl)$ and serum albumin $(3.71\pm0.03g/dl)$ in diarrhoeic calves were observed apparently due to associated dehydration. Tenant et al., (1972), Constable et al., (1996) and Walker et al., (1998) observed the significant increase in both PCV and TSP, which indicates hypovolemia, hemo-concentration and reduced glomerular filtration rate. A significant increase in A: G ratio (0.92 ± 0.06) was observed in diarrhoeic calves suggesting either increase in albumin, or due to dehydration (Kaneko, 1989), or decrease in globulins. No significant changes in ALT and AST activities were observed, which indicates the absence of marked hepatic damage. These results were in consistent with the findings of Lewis et al., (1975).

The marked reduction in the value of serum glucose in diarrhoeic calves $(44.67\pm1.80 \text{ mg/dl})$ as compared to healthy control calves $(75.13\pm1.57 \text{ mg/dl})$ was observed, which is in close agreement with Lewis et al., (1975) who observed hypoglycaemia with glucose concentration below 40 mg/dl of plasma in acute severe diarrhea. Grove-White and White (1999) observed a reduced glucose level of 4.87(1.9) mmol/L in diarrhoeic calves as compared to the non-diarrhoeic control calves having glucose level 5.8(0.88) mol/L. Morris et al. (1985) suggested that severe hypoglycaemia may occur as a result of reduced rate of conversion of lactic acid to glucose. Other

factors involved in causing hypoglycaemia in diarrhoeic calves may include anorexia, decreased intestinal absorption of glucose (Bywater et al., 1969; Bywater, 1977), a low level of glucose reserves at this age (Shelly, 1969) and alternation in tissue metabolism caused by decreased blood flow and oxygenation associated with hypovolemic shock (Tenant et al., 1972) which has been invariably present in hypoglycaemic diarrhoeic calves.

A significant decrease in the level of serum sodium in diarrhoeic calves (109.23 \pm 2.68 mmol/L) was recorded against the normal healthy calves (141.94 \pm 1.24 mmol/L). These findings are in agreement with the findings of Sridhar et al., (1988), Maach et al., (1992), Aly et al., (1996) and Grove-White and White, (1999). Hyponatremia occurs as a result of excessive secretion of the Na⁺ ions by intestinal villus cells which are lost through the intestinal tract particularly in enterotoxigenic *E. coli* induced diarrhoea (Radostits et al., 2007). Most of the diarrhoea causing microorganisms disrupt the intestinal function and dehydrate the body either by increasing the chloride-secreting activity of the crypt cell or impairing the absorption of sodium by the villus cells or both (Hirschhorn et al., 1991). Then the fluid that is normally returned to the blood across the intestinal wall is lost in the watery stool.

The present data also revealed a significant increase in serum potassium (5.11±0.13 mmol/L) in diarrhoeic calves. Similar findings were also reported by Sridhar et al., (1988) and Grove-White. (1999). Hyperkalaemia in experimentally induced diarrhoea was also observed by Roy et al., (1984), Groutides and Michell (1990), Michellet et al., (1992), Constable et al. (1996) and Walker et al. (1998). Hyperkalaemia may be due to increased potassium retention by kidney and also due to cellular damage (Fisher, 1971).

A reduced value of serum chloride was observed in diarrhoeic calves $(5.11\pm0.13 \text{ mmol/L})$ as compared with the normal healthy calves $(102.06\pm1.84 \text{ mmol/L})$ which is in conformity with data as reported by many workers (Hartmann et al., 1983; Maach et al., 1992; Aly et al., 1996). However, the present findings are contradictory to the reports of some previous studies in which hyperchloraemia have been observed in diarrhoeic calves (Magdolna et al., 1976; Sridhar et al., 1988). Hyperchloraemia was reported to occur as a result of prolonged increased loss of Cl⁻ ions in the intestinal tract during diarrhoea (McSherry and Grinyer, 1954; Radiosits et al., 2007), and failure of gastric H⁺ and Cl⁻ ions to be reabsorbed by the villi of small intestine (Radostits et al., 2007).

A moderate increase in serum urea nitrogen in diarrhoeic calves (27.4±0.60 mg/dl) was recorded as compared to control (19.87±0.95mg/dl) and increased serum creatinine values were also recorded in diarrhoeic calves $(1.68\pm0.03 \text{ mg/dl})$ against the healthy control calves $(0.69\pm0.05 \text{ mg/dl})$. An increase in both the parameters was in consistent with the report by many researchers (Groutide and Michell 1990a; Michellet al., 1992; Constable et al., 1996; Walker et al., 1998; Grove-White, 1999). Tenant et al., (1972) observed a significant increase in BUN of 50.1±18.5mg/dl in calves with acute enteric infection while Grove-White and White (1999) recorded a significant increase in urea level as 13.9±7.1 mmol/L in diarrhoeic calves against the value of non-diarrhoeic control calves, (3.3+0.8 mmol/L). Similarly, a significant increase in creatinine level of 221 ±129.5 mmol/L was also observed in diarrhoeic calves as compared to non-diarrhoeic control calves, (101±13.7 mmol/L). The significant increase in serum urea nitrogen and creatinine in the present study was attributable to dehydration as a result of diarrhoea. The hypovolemia due to dehydration results in concentration of the plasma solutes with proportionate increase in both the parameters. However, such cases of pre-renal azotaemia need to be differentiated from other conditions including decreased peripheral vascular perfusion. Further, in case of hypovolemia, impaired excretion of urea and creatinine may occur, secondary due to reduced renal blood flow and GFR (glomerular filtration rate). Tenant et al. (1972) demonstrated varying degrees of renal insufficiency in calves with severe diarrhoeal diseases. They believed that renal insufficiency was caused primarily by decreased blood flow (pre-renal azotemia) as commonly observed by many workers in diarrhoeic calves (Groutides and Michell, 1990a and Bouda et al., (1987). Haematobiochemical changes recorded in Colibacillosis affected diarrhoeic calves in the present study may be useful for treatment of calf diarrhea as it is the most common cause in neonatal calves.

Parameters	Healthy control calves (n =5)	Diarrhoeic calves (n= 25)
Hb(g%)	12.76±0.08	12.64±0.18
PCV (%)	34.08±0.4	43.8±0.48**
Total protein (g/dl)	7.16±0.07	7.74±0.08**
Albumin (g/dl)	3.23±0.07	3.71±0.03**
Globulin (g/dl)	3.93±0.19	4.03±0.11
A:G ratio	0.82±0.07	0.92±0.06*
Glucose (mg/dl)	75.13±1.57	42.48±1.96**
Creatinine (mg/dl)	0.69±0.05	1.68±0.03**
Urea nitrogen (mg/dl)	19.87±0.95	27.4±0.60**
Sodium (mmol/L)	141.94±1.24	109.23±2.68**
Potassium (mmol/L)	4.32±0.08	5.11±0.13**
Chloride (mmol/L)	102.06±1.84	95.11±2.19**
ALT (U/L)	38.88±1.22	38.92±1.159
AST(U/L)	16.47±1.01	16.34±0.97

Table No.1: Haemato- biochemical changes in healthy and diarrhoeic calves

* Significant at p<0.05, ** Significant at p<0.01

Conclusion

Analysis of blood and serum samples of the diarrhoeic calves revealed significant increases in PCV, TSP, serum A: G ratio, serum urea nitrogen, serum albumin, creatinine and potassium and significant decreases in serum glucose, sodium and chloride. As regards haemoglobin, serum ALT, AST and globulin, no significant alteration were observed between the diarrhoeic calves and healthy control calves.

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Competing interests

The authors declare that they have no competing interests.

References

- 1. Aly, A.O., Abd-El-Wahed, Z.H., Kohilo, K. and El-Shiekh, A.R. (1986). Some studies on clinical, haematological and biochemical changes in diarrhoeic neonatal buffalo calves with reference to hygienic conditions. Assuit Vet.Med. J. **35**: 91-104.
- Azizzadeh, M., Shooroki, H. F., Kamalabadi, A. S. and Stevenson, M. A., (2012). Factors affecting calf mortality in Iranian Holstein dairy herds. Prev. Vet. Med. 104:335–340.
- 3. Bouda, J., Passch-Martinen, L. Candanosa, E.A., Lopez-Romahn, C. and Quiroz-Rocha, G.F. (1987). Study of the clinical and biochemical parameters before and during oral rehydration in calves with diarrhoea. Veterinaria Mexico, **28**: 87-91.
- 4. Bywater, R.J. (1977). Evaluation of an oral glucose-glycine- electrolyte formulation and amoxicillin for the treatment of diarrhoea in calves. Am. J. Vet. Res. **38**: 1983-1987.
- 5. Bywater, R.J. and Penhale, W.J. (1969). Depressed lactase activity in the intestinal mucous membrane of calves after neonatal diarrhoea. Res. Vet. Sci. **10**(7):1035-1038.
- Coles E. H. (1980). Veterinary Clinical Pathology, 3 rd edition. W. R. Saunders, Company, London.Colorado: Veterinary Services, NAHMS, Animals and plant inspection Service, US Dept. of Agriculture, 37.
- 7. Constable, P.D. Goher, H.M., Morin, D.E. and Thurmon, J.C. (1996). Use of hypertonic saline dextran solution to resuscitate hypovolemic calves with diarrhoea. Am. J. Vet. Res. **57:** 97-104.
- 8. Fisher, E.W. and Dela Fuente, G.H. (1971). Water and electrolyte studies in new born calves with particular reference to the effect of diarrhoea, Res. Vet. Sci. 13:315-322.
- 9. Foster D.M. and Smith G.W., (2009). Pathophysiology of Diarrhoea in Calves. Veterinary Clinics of North America: Food Animal Practice, **25**:(1) 13–36
- 10. Groutides, C.P. and Michell, A.R. (1990). Changes in plasma composition in calves survining or dying from diarrhea. Br. Vet. J. **146**:205-210.
- 11. Grove- White, D.H. and White, D.G. (1999). Abdominal distension in collapsed diarrhoeic calves: biochemical findings and treatment. Vet. Rec. **144**: 639-642.
- 12. Hartman H., Mayer, H., & Steinbach, G, Rossow, N. and Lesche, R. (1983). Influence of diarrhea on electrolyte content and osmolarity of the blood of claves. Mona. Fur. Veterinarmedizin, **38**: 292-296.
- Hirschhom., N and Greenough III, W.B (1991). Progress in oral rehydration therapy. Scientific American. 264:50-56.
- Horvay- Magodlna, S., Romvary, A. and Misley, A. (1976). Combined treatment of coli enteritis of calves. Magyar AllotorvosokLapja. 32: 381-384
- 15. Hultman, E. (1959). Rapid specific method for determination of aldosaccharides in body fluids. Nature. **183**: 108-109.
- Kaneko. J.J., (1989). Clinico-Biochemistry of Domestic Animals. 4th Ed. Department of Clinical pathology school of Vet. Med. University of California, Davis, California.
- 17. Lewis, L.P., Phillip, R.W. and Elliot, C.D. (1975). Changes in plasma glucose and lactate concentration and enzyme activities in the neonatal calf with diarrhoea. Am. J. Vet. Res. **36**: 413-16.
- 18. Maach, L., Grunder, H.D. and Boujya, A. (1992). Clinical and haematological investigations in new born Holstein Friesian calves with diarrhea in Morocco. Deutsche TierarztlicheWochenschrfi.**99:** 133-140.
- 19. Marsh, W.H., Fingerhut and H. Miller, (1965). Non protein nitrogen, urea, urates, creatinine and creatinine. In practical clinical biochemistry. 5th (edn), William Heinemann, Medical book ltd, London, PP: 460.
- 20. McSherry, B.J. and Grinyer, I. (1954). Disturbance in acid-base balance and electrolyte in calf diarrhea and their treatment. A report of eighteen cases. Am. J. Vet. Res. **12:**535-541.
- Michell, A.R., Brooks, H.W., White, D.G. and Wagstaff, A.J. (1992). The comparative effectiveness of three commercial oral solutions in correcting fluid, electrolyte and acid –base disturbances caused by calf diarrhea. Br .Vet .J. 148: 507-522.
- 22. Miller, W.G. (1984). Eletrolytes In: Clinical Chemistry Theory, Analysis, and Correlation, Kaplan, L.A and Pesce A.J (Eds.), CV Mosby, Toronto pp. 1044-1079
- Morris, J.A. Weils, G.A.H., Scott, A.C. and Sojka, W.J. (1985). A comparison of methods for demonstrating colonization in the small intestine of piglets by enterotoxigenicE. coli. Br. Vet, J. 141: 484-489.
- 24. Radostits, O.M., Gay, C.C., Hincheliff, K.W. and Costable, P.D. (2007). Veterinary Medicine. A text book of the diseases of cattle, sheep, pigs, goats and horses, 10th Edition, New York, W.B. Saunders Company Ltd.77, 847-896

- 25. Ramkumar, P. K. (2012). Evaluation of therapeutic potential of plant extracts against e.coli diarrhoea in calves. MVSc thesis, IVRI Deemed University, Izatnagar, India
- Reitman, S. and Frankel, S. (1957). Calorimetric determination of serum glutamic oxaloacetictransminase and serum glutamic pyruvic transaminase. Am. J. Clin. Pathol. 28: 56-63.
- 27. Roy, S., Sinha, R.P. and Prasad, R.S. (1984). Evaluation of oral fluids for treatment of diarrhea in kids. Indian Vet. J. **61**: 946-952.
- 28. Schoenfeld, R.G. and Lewellen, C.J. (1964). A colorimetric method for determination of Serum chloride. Clin. Chem. **10**(6): 553.
- 29. Shelly W. S., Mebus, C. A., Underdahl, N.R., Rhodes. M.B., and Twiehaus, M.J. (1969). Future studies on neonatal calf diarrhea virus .Proc. Annu. Meet .US. Anim. Health Assoc., **73**:97-99
- Sridhar, Pachauri, S.P. and Kumar, B.E. and Nightengale, G.T. (1988). Clinico-pathological alterations in calf scour. Indian Vet. J.65: 771-774.
- 31. Tenant. B and Reina-Guerra, H.M. (1968). Hypoglycaemia in neonatal calves associated with acute diarrhea. Cornell Vet. **58**:136-146.
- 32. Tenant, B. Harrold, D. and Reina-Guerra, H.M. (1972). Physiologic and metabolic factors in the pathogenesis of neonatal entero infections in calves. J. Am. Vet. Med. Assoc. **161**: 993-1007.
- 33. Trinder, P. (1951). A rapid method for the determination of sodium in serum. Anal. Chem. 76: 596-599.
- 34. Vankampen, E.J. and Zinglstra, W.G. (1961). Colorimetric determination of haemoglobin. Clinica Chemica Acta. 6: 3588.
- Varley, H., A.H. Grawlock and Bell, M. (1980). Practical biochemistry. Vol.I 5thedn, William Heinmann, Medical book ltd, London: 458-484.
- Walker, P.G., Constable, P.D., Morin, D.E., Foreman, J.H., Drackley, J.K. and Thurmon, J.C. (1998). Comparison of hypertonic saline-dextran solution and lactated Ringer's solution for resuscitating severely dehydrated calves with diarrhoea, J. Amer, Vet. Med. Assoc., 213: 113-22.