



ISSN NO. 2320-5407

Journal Homepage: -www.journalijar.com

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH (IJAR)

Article DOI:10.21474/IJAR01/8172
DOI URL: <http://dx.doi.org/10.21474/IJAR01/8172>



INTERNATIONAL JOURNAL OF
ADVANCED RESEARCH (IJAR)
ISSN 2320-5407
Journal Homepage: <http://www.journalijar.com>
Journal DOI:10.21474/IJAR01

RESEARCH ARTICLE

ANTIMICROBIAL EFFECTS ALCOHOLIC EXTRACT OF YARROW (ACHILLE AMILLEFOLIUM) AGENTS STAPHYLOCOCCUS AUREUS AND ESCHERICHIA COLI THE MOST COMMON SOURCE OF URINARY TRACT INFECTION.

Sabiha Sharif Salih¹, Adel Kamal Khider² and Mariwan Abdullah Hama Salih³.

1. Medical Laboratory Department Technical College of Health Sulaimani Polytechnic University Sulaimani, Iraq.
2. Department of Biology College of Education Salahadin University Erbil, Iraq. Medical Laboratory
3. Medical Laboratory Department Technical College of Health Sulaimani Polytechnic University Sulaimani, Iraq.

Manuscript Info

Manuscript History

Received: 08 October 2018

Final Accepted: 10 November 2018

Published: December 2018

Keywords:-

Yarrow, Achillea millefolium antimicrobial, Escherichia coli, Resistant, and Staphylococcus aureus.

Abstract

Effect plant extract to eliminate resistance genes in E. coli and S. aureus were isolated from urinary tract infections. All the samples in this research were taken from the pregnant woman with the age of (25-45) in Sulaimani city during May (2015-2016). Out of 200 different urine samples, 50 positive cultures were isolated and diagnosis. In (35) samples, E. coli were the predominant microorganisms and were responsible for (17.5%) of the urinary tract infections and the other 15 samples (7.5%) were S. aureus. All isolates were obtained depending on cultural, morphological identification, in addition to, API20E system. The resistances of the isolates were examined against eleven widely used antimicrobials, which were Cefotaxime (Cef), Ampicillin (Amp), Nitrofurantoin (Nit), Penicillin (Pn), Trimethoprim (Tri), Rifampicin (Rif), Tetracycline (Tet), Gentamycin (Gm), Streptomycin (Str), Ciprofloxacin (Cip) and Amikacin (Ak). The positive cultures were classified according to their resistance to the above antimicrobials. They were grouped into (8) E. coli antibiograms and (5) S. aureus antibiogram. All the isolates were resistant to (Pi). Three isolated for E. coli (E3, E18, and E35) and two isolated for S. aureus (S39 and S45), resisted to all the tested antibiotics while the other samples varied in their resistance. The ethanolic extracts in Minimum Inhibitory Concentration [MIC] for Achillea millefolium was (3000) µg/ml. Sub-MIC of plant extract was used as eliminated antibiotic resistant genes of isolates and the results were SMIC 2500 µg/ml of ethanolic extract of Achillea millefolium eliminated the genes that are responsible for E. coli (E3, E18, E35) isolates (Ak, Str, Rf, Gm, Tri, Tet, Cip), while reduced the percentage of resistance for these antibiotics Amp, Cef, and Pi which ranged between (100% - 10%), SMIC 3500 µg/ml of ethanolic extract of Achillea millefolium affected on (S39) genes responding on Str, Rf, Amp, Ak, Gem, Nit, Tri, Tet, Cef and Cip reduced the percentage (%100, while not affected by resistant gene in (Pi), although the genes in (S45) Responding on Ak, Str, Rf, Gem, Nit, Tri, and Tet while not affected on resistant gene in (P, Cef, Cip, and Amp).

Corresponding Author:-Sabiha Sharif Salih.

Address:- Medical Laboratory Department Technical College of Health Sulaimani Polytechnic University Sulaimani, Iraq.

Introduction:-

Escherichia coli (E. coli) and Staphylococcus aureus (S. aureus) are the most common cause of serious human and animal infections. They are the most common causes of soft tissues and skin infections, bone, surgical wound and joint infections. S. aureus is the most common cause of hospital-acquired bacteremia and respiratory tract infections [1]. E. coli is the most common cause of urinary tract infections (UTIs) [2], enteric infections [3], and the systemic infections including, osteomyelitis, bacteremia, nosocomial pneumonia, and infectious arthritis in humans. E. coli is also the leading cause of neonatal meningitis [4]. There is a wide range of antimicrobials affecting and limiting the growth of E. coli. Some examples of the antimicrobial agents include trimethoprim-sulfamethoxazole, fluoroquinolones, lactams, Aminoglycosides, and, that are normally used to treat community and hospital E. coli infections [5]. However, isolates which are antimicrobial resistant for example, those producing extended-spectrum lactamases and those that are fluoroquinolone-resistant have raised significantly in certain areas that are nosocomial and community-acquired during the 2000's.

E. coli is currently resistant to the several major antimicrobial classes [5]. Penicillin, Cloxacillin, and dicloxacillin are still the antibiotics chosen for the management of serious methicillin-susceptible S. aureus (MSSA) infections, but in patients with penicillin hypersensitivity, first-generation cephalosporin, cephalothin, ceftazidime, lincomycin, cephalexin, clindamycin, and erythromycin have important therapeutic roles in less serious MSSA infections. For serious MRSA infections the patient should be treated with parenteral vancomycin or, if the patient is vancomycin allergic they should be treated with teicoplanin [6].

A major health concern is Antibiotic-resistant Staphylococci, since the bacteria can be easily spread in the environment. Currently we can see that Infections caused by methicillin-resistant S. aureus (MRSA) have increased around the globe during the past twenty years [7, 8]. Multiple drug-resistant S. aureus has been frequently retrieved from foodstuffs [1], water and biofilm formation [11] nasal mucosa of humans [12], clinical cases [13] and livestock [14]. Some reports on S. aureus isolate with partial or complete resistance to vancomycin portend a chemotherapeutic in which effective bactericidal antibiotics against this organism may no longer be easily available [9,10].

The World Health Organization concluded that 80% of the world population used plant extracts or their active decisions were used as medicine conventional therapies [15]. Currently Over 50% of all modern clinical drugs are derivatives that are natural in origin [16]. Iraqi People have traditionally used a number of plants species for treatment of infectious disease and various infection [17]. Furthermore, this research was conducted in order to study and isolate E. coli and S. aureus in urinary tract infection and to test antibiotic-resistant treatment of those two strains with Achillea millefolium (A. millefolium). A. millefolium grows widely all around Europe, Asia, North Africa and North America and it is widespread and frequently used in Italian traditional medicine [18]. Its benefits have been known since ancient times and its spread out over many cultures from Europe to Asia: A. millefolium is recommended for the treatment of many different ailments in Greece, in the region of Thessaloniki [19]. In West Azerbaijan, Iran, they use the Infusion of dried flowers is considered, because it is considered that it is suitable for the treatment of dyspepsia, hemorrhoids, gastritis and dysmenorrhea. Another example is leaves, and flowers are used for the treatment of gastric problems and fever in the Parvati valley, West Himalaya, India.

Materials And Methods:-

200 samples were collected from urine of patients were suffering from UTI in Sulaimani City-Iraq during April (2015-2016). The patients were all pregnant woman between the age of 25-40 years. All isolated were diagnosis depending on Morphological, cultural, and biochemical analysis according to (20) in addition to using the API20E system to analyze the bacteria.

1. Plant Extraction: The alcohol extract of A. millefolium was prepared according to the instructions [21]. By using Soxhlet extraction technique fifty gram of dried powder was produced. The plant extract was concentrated to dryness in the vacuum oven at 50°C. a small (1.0g) portion of the sample was sterilized and diluted in 100 mL of sterile distilled water.
2. Determination of MIC: The dilution method of bacteria isolate inoculum Standardize using standard curve prepared previously as recommended by [22] was used to test The antimicrobial activity of ethanol extracts of A. millefolium. By further diluting the bacterial suspensions we obtained the 1×10^5 CUF inoculum. Using spectrophotometric and by the account of viable cells on nutrient agar [29] [23].

3. Determination of antibiotic susceptibility: By using dilution method in agar plate [23] The antimicrobial resistance phenotypes of all isolated bacteria were determined. The antibiotics purchased from sigma company (Germany) were: Cefotaxime (Cef), Ampicillin (Amp), Gentamycin (Gm), Trimethoprim (Tri), Penicillin (Pn), Tetracycline (Tet), Streptomycin (Str), Nitrofurantoin (Nit), Ciprofloxacin (Cip), Amikacin (Ak), and Rifampicin (Rif). These antibiotics were added to the medium at different concentrations after cooling and sterilization. Then, the medium was poured and mixed into Petri-dishes. After that, it was inoculated using streaking method. Resistance was recorded after incubation at 37°C for 24h.

Result:-

50 positive cultures were isolated from the 200 samples taken for this research. The 50 positive cultures concluded that *E. coli* was a predominant organism and were responsible for 35 (17.5%) of the cases and the rest (15 samples 7.5%) was *S. aureus*. All the isolated samples were examined for their resistance to eleven widely used antibiotics in medicine. The *E. coli* bacteria that were isolated were grouped into 8 biotypes shown in (Table 1) with their corresponding resistant pattern and the isolated *S. aureus* bacteria were grouped into 5 biotypes shown in (Table 2) with their corresponding resistant pattern. Among 35 isolated of *E. coli*, 3 samples (E3, E8, and E35) showed a resistance to all eleven antibiotics. Among 15 isolated of *S. aureus*, 2 samples (S39, and S45) showed a resistance to all eleven antibiotics. The alcohol extract of *A. millefolium* was used on the resistant samples of *E. coli* (E3, E8, and E35) and *S. aureus* (S39, and S45). Sub-MIC 2500 µg/ml was used with Ak, Str, Rf, Gm, Tri, Tet, Cip, and Amp against the resistant genes of *E. coli* (E3, E8, and E35) shown in (Table 3). Sub-MIC 3500 µg/ml was used with Ak, Str, Rf, Gm, Nit, Tri, and Tet, P, Amp, Cip, and Cef against the resistant genes of *S. Aureus* (S39, and S45) shown in (Table 4).

The solution of Sub-MIC (2500 µg/ml) in combination with Ak, Str, Rf, Gm, Tri, Nit, Tet, and Cip, was effective against the resistant genes in *E. coli* (E3, E8, and E35) and decreased resistance 100%, while the plant extract in combination with Amp was only 10% effective against the resistant genes shown in (Table 3).

The solution of Sub-MIC (3500 µg/ml) in combination with the antibiotics Ak, Str, Rf, Gm, Nit, Tri, and Tet decreased genes resistance in *S. aureus* (S39, and S45) 100%, while no effects on the resistant genes for P, Amp, Cip, and Cefin (S45) although not reduced genes resistant in (S39, S45) for Cef which was shown in (Table 4).

Table 1:-Antibiogram *E. Coli* isolated from UTI.

Samples	Ak	P	Amx	Cip	Str	Cef	RF	GM	Tri	Nit	Tet	Amp
1,2,4,6,16	S	R	R	S	S	S	S	S	S	S	S	R
5,7,8,17,32	S	R	S	S	S	S	S	S	S	S	S	R
3,18,35	R	R	R	R	R	R	R	R	R	R	R	R
9,10,20,33	R	R	R	R	R	R	S	S	R	S	S	R
11,12,13,14,15	S	R	S	R	S	S	S	R	R	R	R	R
19,,21,24,25,27	S	R	R	R	R	R	R	R	R	S	S	R
28,31,34	S	R	S	R	R	R	S	S	R	R	S	R
22,23,26,29,30,	S	R	S	R	R	R	S	R	R	S	S	R

Table 2:-Antibiogram *S. aureus* isolated from UTI.

Samples	Ak	P	Am	Ci	Str	Ce	Rf	GM	Tri	Nit	Tet	Amp
39,45	R	R	R	R	R	R	R	R	R	R	R	R
36,37,40	S	R	R	R	S	S	S	R	R	S	S	S
38,41,44,46	S	R	R	S	S	R	S	R	S	R	S	S
42,43,47	R	R	S	S	R	R	S	R	S	S	S	R
48,49,50	S	R	R	R	S	R	S	R	R	S	R	R

Table 3:-Effect of *A.millefolium* on resistant genes in *E. coli*.

Samples	Plant extract	Ak	P	Amp	Cip	Str	Cef	Rf	Gm	Tri	Nit	Tet
E coli 3	SMIC 2500	S	R	R	S	S	R	S	S	S	S	S
E18	SMIC 2500	S	R	R	S	S	R	S	S	S	S	S
E35	SMIC 2500	S	R	S	S	S	R	S	S	S	S	S

Table 4:-Effect of *A.millefolium* on resistant genes in *S. aureus*.

Samples	Plant extract	Ak	P	Amp	Cip	Str	Cef	Rf	Gm	Tri	Nit	Tet
S39	SMIC 3500	S	S	S	S	S	R	S	S	S	S	S
S45	SMIC 3500	S	R	R	R	S	R	S	S	S	S	S

Discussion:-

The evolution and spreading of antimicrobial resistance bacteria is generally due to, consumption, mistreatment, self-medication by patients, excessive prescription in which, it is anticipated that 70-80% of prescriptions for antimicrobials are probably advised intentionally by the health professionals like doctors, nurses and pharmacists. In spite of the fact that most acute viral diarrheal and nasopharyngitis events are not viral in origin, yet, antibiotics are used at random to treat them. Reasons for over definition are often in absence of certainty, pharmaceutical company pressure or patient pressure to get treatment. Furthermore; poverty and lacking access to antibiotics constitute a major factor in the development of antibiotic-resistance bacteria in addition to improper diagnosis and wrong treatments which, in many instances the laboratory diagnostic compels the physician to define antibiotics empirically, thus, increasing the likelihood of the patient receiving a wrong antibiotic. The accessibility of antibiotics over-the-counter and sales promotion schemes by the pharmaceutical companies also leads to pressure on the doctors to promote wholesales to the patients, thus, increasing the likelihood of over usage and increasing the probability of emerging antimicrobial resistant bacteria. Another major issue is changing drugs which is contributing to the development of resistance because different drugs contain either the wrong ingredient or lesser amount of the active ingredient.

The treatment of infectious diseases with antimicrobial agents carrying onto present problems in the modern day. Medical studies showed a significant increase in the occurrence of side effects and the resistance that pathogenic microorganism against several antibiotics [28]. However, in the modern day attention has been paid to plant and their biologically active compounds which are isolated and used in herbal medicine such as flavonoids, alkaloids, tannins, Phenolic and glycosides Compounds [29]. The activating of the plant on resistant genes because Plants, is a source of medicinal compounds have continued to play a major role in the maintenance of human health since ancient times. So that, plants with potential antimicrobial activity has to be tested against the right microbial model to confirm its activity [29]. Thus, the aim of this research was to study two Pathogenic bacteria (*E. coli* and *S. aureus*) from the urine of pregnant women who were suffering from Urinary Tract Infections (UTI) and study the effects of *A.millefolium* extract on their antibiotic-resistant gene. In this research, 200 samples were taken from a pregnant woman at age 25 to 40 years old at Sulaimani City. From those 200 samples, 50 samples were positive cultures. The most common bacteria isolated in these patients was *E. coli* (35 samples, 17.5%) and the rest (15 samples 7.5 %) was

S. aureus. This finding is similar to other reports, which indicated that UTI isolated in patients is caused by the commonest pathogen *E. coli* that a gram-negative bacterium [25, 21]. This was because of the fact that strains of *E. coli* are causing the urinary tract and possess a variety of virulence characteristics which facilitate their intestinal carriage and persistence in the vagina and then ascension and invasion of the normal urinary tract [26].

All the isolates were tested for antibiotic-resistant as it is shown in (Table 1) and (Table 2). It was concluded that three strains of *E. coli* (E3, E18, and E35) were totally resistant to all used antibiotics, and two strains of *S. aureus* (S39, and S45) were also totally resistant to the antibiotics. The alcoholic extract of *A. millefolium* was used to reduce antibiotic-resistant, which was shown in the (Table 3) and (Table 4). Antibacterial effect of *A. millefolium* was evaluated in vitro against the three pathogenic bacteria species of *E. coli* (E3, E18, and E35) were more responsive to the *A. millefolium* than *S. aureus* at the applied concentrations (SMIC 2500 µg/ml) and reduced resistant genes 100% in combination with Ak, Str, Rf, Gm, Tri, Nit, Tet and Cip, but did not reduce any genes resistant for P and Cef shown in T3. Also, The Ethanol plant Extract, Which Was Used For Reducing Resistant Genes In *S. aureus* At The Applied Concentrations (Smic 3500 µg/ml) affected the resistant genes in S39 and S45, which was shown in (Table 4). SMIC Plant extracts converted genes resistant for Ak, Str, Rf, Gm, Nit, Tri and Tet in S39 and S45 to sensitive but there was no effect on genes resistant to Pi, Amp and Cip in (S45) and it did not reduce any genes resistant for Cef. The result of this study agreed with the recent investigation, which has also pointed out to the notable effect of extracts of aerial parts of *A. millefolium* as it showed a broad spectrum antimicrobial activities against *S. aureus* and *E. coli* [24, 3]. It could be concluded that the alcoholic extract of *A. millefolium* has considered be able to reduce the resistive genes of both *S. aureus* or *E. coli*.

References:-

1. HH Abulreesh, Organji SR, "The prevalence of multidrug-resistant staphylococci in food and the environment of Makkah, Saudi Arabia," *Res J Microbiol.*, 6(6):510-523, 2011.
2. B. Foxman, "The epidemiology of urinary tract infection," *Nat Rev Urol.*, 7:653-60.644, 2010.
3. F. Candan; M. Unlu; B. Tepe; D. Daferera; M. Polissiou; A. Sökmen; Akpulat, H.A., "Antioxidant and antimicrobial activity of the essential oil and methanol extracts of achillea millefolium subsp. Millefolium,"
4. KS. Kim, "Current concepts on the pathogenesis of Escherichia coli meningitis: implications for therapy and prevention," *Curr Opin Infect Dis.*, 25:273-8, 2012.
5. JD. Pitout, "Extraintestinal pathogenic Escherichia coli: an update on antimicrobial resistance, laboratory diagnosis, and treatment," *Expert Rev Anti Infect Ther.*, 10:1165-76, 2012.
6. C. Rayner, WJ. Munckhof, "Antibiotics currently used in the treatment of infections caused by Staphylococcus aureus," *Intern Med J.*, 35 Suppl2:S3-16, 2005.
7. S. Deresinski, "Methicillin-resistant Staphylococcus aureus: an evolutionary, epidemiologic and therapeutic odyssey," *Clin Infect Dis.*, 40:562-573, 2005.
8. G Ippolito, S Leone, FN Lauria, E Nicastrì, RP Wenzel, "Methicillin-resistant Staphylococcus aureus," *The superbug. Int. J. Infect. Dis.*, 14 (Suppl. 4): S7-S11, 2010.
9. KHiramatsu, H Hanaki, T Ino, K Yabuta, T Oguri, FC Tenover, "Methicillin-resistant Staphylococcus aureus clinical strain with reduced vancomycin susceptibility," *J Antimicrob Chemother*, 40(1):135-6, 1997.
10. Centers for Disease Control and Prevention (CDC), "Staphylococcus aureus resistant to vancomycin-United States," *MMWR Morb Mortal Wkly Rep.* 51(26):565-7, 2002.
11. M Lancellotti, MP de Oliveira MP, FA de Avila. "Research on Staphylococcus sp. in biofilm formation in water pipes and sensibility to antibiotics," *Braz J Oral Sci.* 6: 1283-1288, 2007.
12. M Acco, FS Ferreira, JAPhenriques, EC Tondo, "Identification of multiple strains of Staphylococcus aureus colonizing nasal mucosa of food handlers," *Food Microbiol.* 20:489-493, 2003.
13. S Stefani, A Goglio, "Methicillin-resistant Staphylococcus aureus: related infections and antibiotic resistant," *Int. J. Infect. Dis.* 14 (Suppl. 4): S19-S22, 2010.
14. M Wulf, A Voss, "MRSA in livestock animals-an epidemic waiting to happen?" *Clin. Microbiol. Infect.* 14: 519-521, 2008.
15. B. Anonymous, *J. Acad. Indus. Res.* 1(11): 50-53, 1993. (28), 13-14. Infections among hospitalized patients,"
16. J.T. Baker; R.P. Borris; B. Carte, "Natural product drug discovery and development new perceptives on international collaboration," *J. Nat. Prod.*, (58), 1325-1325, 1995.
17. A.K. Najla; F. Awaz, "Isolation and antibacterial evaluation of plant extracts from some medicinal plants in Kurdistan region," *J. Duhok Univ.* 12, (1) (Special Issue), 250-255. 2009.

18. A Pieroni, CLQuave, "Traditional pharma copoeias and medicines among Albanians and Italians in southern Italy: a comparison," *JEthnopharmacol*, 101: 258–270, 2005.
19. S Kokkini, V Kleftoyanni, E Hanlidou, R Karousou, "The herbalmarket of Thessaloniki (N Greece) and its relation to the ethnobotanicaltradition," *J Ethnopharmacol* 91: 281299, 2004.
20. w.h. Andrews and t. hammock, "bacteriological analytical manual," fda, us food and drug administration 2000.
21. J.B.Harbone, T.J. Mabray and H. Mabray, "Physical and Function of Flavonoids", AcademicPress,
22. R. Cruickshank, J.P. Duguid, B.P., "Marmiental Microbiology and R.H.A. Swain, Medical Microbiology". 12th Edn., Churchill Livingstone, London, 2: 236,432-434,1975.
23. R.m.atlas,a.e. brown and l.c. parks, "laboratory manual experimental microbiology," mosby-year book inc., st Louis, 1995.
24. Hasson Rasha N., "Antibacterial Activity of Water and Alcoholic Crude Extract of Flower Achillea millefolium," 2011
25. M.A. Mahmood, Department of Basic Sciences, College of Dentistry, Baghdad University, Baghdad-Iraq, 2011
26. T. Annabelle, M.D. Dytan, A. Jennifer and M.D. Chua. "Surveillance of pathogens 15 and resistance patterns in urinary tract infections," *Phil. J. Microbiol. Infect. Dis.* 28(1): 11-14, 1999.
27. C.M Kunin, "Resistance to antimicrobial drugs a world-wide calamity," *Annals of Internal. Medi.*, (118), 557-561, 1993.
28. C. Evans; A. Bansot; A. Samuel, "Efficacy of some nape medical plants against Salmonella: an in vitro study," *J.Ethnopharmacol.*, (80), 12-24, 2002.
29. R. Nair; T. Kaalariye; S. Chanda, "Antimicrobial activity of some selected Indian medicinal flora," *Turk J. Biol.*, (29), 41- 47, 2005.