

RESEARCH ARTICLE

PHYLOGENETIC ANALYSIS AND SPECIES IDENTIFICATION OF 11 GORGONIAN CORALS (OCTOCORALLIA: ALCYONACEA) IN THE NORTH CENTRAL COAST OF VIETNAM BASED ON *MSH1*mtDNA AND 28S rDNA MARKERS

Nguyen Chi Mai^{1,2}, NinhThi Ngoc^{1,2}, Nguyen Xuan Cuong^{1,2}, Nguyen Hoai Nam¹, Nguyen Tuong Van³, Le Quang Trung⁴, Pham Thi Hoe¹, Nguyen Quang Hung⁴ and Tran My Linh^{1,2}

- 1. Institute of Marine Biochemistry, Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet, CauGiay, Hanoi, Vietnam.
- 2. Graduate University of Science and Technology, VAST, 18 Hoang Quoc Viet, CauGiay, Hanoi, Vietnam.
- 3. Institute of Biotechnology, VAST, 18 Hoang Quoc Viet, CauGiay, Hanoi, Vietnam.
- 4. VNTEST Institute for Quality Testing and Inspection, 7/161/28 Nguyen Xien, Thanh Xuan, Hanoi, Vietnam.

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Manuscript Info

Abstract

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..... Vietnam contains diverse marine ecosystems with the high biodiversity of marine organisms, including gorgonian corals of Alcyonacea order. In order to support traditional classification of these corals, in this study mitochondrial barcoding markers msh1 and nuclear 28S rDNA were developed for analysis of 11 specimens collected in 2015 and 2016 from different islands and bays along the North Central coast of Vietnam. Phylogenetic analyses based on msh1 and 28S sequence polymorphism showed that all specimens belonged to Anthozoa class, Octocorallia sub-class and Alcyonacea order. At lower taxa levels, they were divided into 4 sub-orders, 7 families and 7 genera according to 7 distinct clades with bootstrap values from 99-100%. The identifications of 7 out of 11 specimens including Sinularia brassica (2 specimens) and Sinularialeptoclados, Dichotellagemmacea, Annella reticulata, S. conferta and S. nanolobata were in concordance between morphological and molecular methods. The other 4 specimens were only identified at genus levels of Astrogorgia sp., Melithaea sp. Scleronephthya sp. and Muricella sp. by either msh1-morphology or msh1-28S markers. These results highlight the importance of molecular markers to elucidate patterns of biodiversity and species identification of soft coral.

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Introduction:-

Vietnam containsdiverse and rich resources of coastal ecosystems harbouringthe high biodiversity of marine organisms, especially gorgonian corals of Alcyonacea order. Until 1952, Dawydoffhad first reported about of soft coral species in Vietnam and classified them into 6 families including Alcyonidae, Fasciculariidae, Xaniidae, Telestaceidae, Tubiporidae and Nephtheidae. Tixier-Durivault (1970) conducted the comprehensive study on soft coral samples in the Museum of Oceanography (Nha Trang, Vietnam) and reported 94 soft coral species belonging to 15 genera and 5 families. Among them, 18 species of *Sinularia* genus were recorded as new species. From 1990s to

Corresponding Author:- Tran My Linh

Address:- Institute of Marine Biochemistry, Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet, CauGiay, Hanoi, Vietnam.

2010s, investigationson biodiversity and distribution of soft coral communities in some islands such as Phu Quoc, Con Co, Truong Sa, Ly Son, Cat Ba, Cu Lao Cham and coastal areas such as Nha Trang Bay, Ha Long Bay were carried out. The results revealed that soft coral species in Vietnam belong to 22 genera and 9 families (Vo et al., 1997; Dautova and Savinkin, 2009; Ben and Dautova, 2010; Dautova et al., 2010; Thao and Ngai, 2013; Dautova and Savinkin, 2013; Ben et al., 2018; Ben and Quang 2020). Molecular DNAstudieson soft coral speciesin Vietnam are still very poor. Recently, Lien et al. (2015) analyzed phylogenetic relationship of Sinularia, Sarcophyton and Lobophytum genera in Nha Trang Bay (South Central coast of Vietnam) using 696bp sequences of msh1 genes and 866bp fragment of *irg-cox1* gene. There has not been, however, research on this approach of soft corals in the North Central coast of Vietnam. Accurate identification of soft coral species is fundamental for genetics, physiology, ecology, particularly applied pharmacology. Molecular identification has been used in biodiversity and conservation, but it also has great potential for applications in taxonomy. Recently, based on nucleotide sequences, several coral species-specific molecular markersfromribosomal subunits 16S, 18S, 28S and genes coding for proteins on mitochondrial DNA (mtDNA) have been found (France et al., 1996; McFadden et al., 2006a, 2006b; Shearer et al., 2002; Herrera et al., 2009). Among molecular markers on mtDNA, msh1 gen encoded for MSH1 protein, has been used widely in molecular taxonomy of soft coral. It is known that the genetic diversity of msh1 gene is as twice as other coding genes on mtDNA. Moreover, msh1 gene is present in mtDNA of almost all soft corals published in the present GenBank (van der Ham et al., 2009). The diversity of the *msh1* gene also occurs among species in a genus and has been used to phylogenetic analysis of soft coral species in the Asia-Pacific (Thoma et al., 2009).

In the present study, phylogenetic analysis and species identification of 11 soft coral specimens collected in the North Central region of Vietnam were investigated based on the 639bp on their *msh1*mtDNA and the 704bp fragment on 28S rDNA gene. The congruence between morphological analysis and molecular markers in species identification could offer an alternative taxonomic method for further biodiversity studies of soft corals in Vietnam.

Materials and Methods:-

Materials

Soft coral specimens were collected in different islands and bays of the North Central region of Vietnam in 2015-2016using SCUBA diving equipment. A map of sampling location and specimen name are shown in Fig 1. The specimens were washed several times with fresh water to remove sand, algae and other surface organisms and then 5 times with distilled water.One portion of each sample was stored in 70% ethanol for morphological analysis and the smaller portion was kept at -80°C for DNA extraction. All vouchers were deposited at Institute for Marine Biochemistry, Vietnam Academy of Science and Technology (VAST).Morphological characters of specimens were identified by Prof. Do Cong Thung (Institute of Marine Resources and Environment, VAST) right after sampling and cleaning.

PCR and DNA Sequencing:

Genomic DNA of each specimen was isolated using DNeasy® Tissue Kit (Qiagen) according to the manufacturer's instruction. The concentration of DNA was quantified using a NanoDrop 1000 instrument (Thermo Scientific, USA) and by electrophoresis in 0.8% agarose.

The 639 bp DNA fragment of *msh1* gene and704 bp - DNA fragment of 28S rDNA gene wereamplified using genomic DNA of soft coral specimens as templates and the specific primer pairsMSHF (5'-ATGAACCAGATACCTATGC-3') and MSHR (5'-AGTGTTTCTCCCATAACTTC-3');28SF (5'- CGTTGAAAGGGAAGCGAATG -3') and28SR (5'-AGGGAACCAGCTACTAGATG-3'). PCR components were $5 \mu l 10X$ PCR Buffer, 1 mM dNTPs, 2 mM primers, 50 ng of genomic DNA, 1 unit of Taq Polymerase and 2.5 mM of MgCl₂ and H₂O up to total 50 μ l. The PCR cycle were 94°C for 3 min, followed by 30 cycles of 1 min at 94°C, 30s at an annealing temperature of52°C for MSHF/R primers or 55°C for 28SF/R primers, DNA synthesis for 1 min at 72°C, and then a final extension of 10 min at 72°C. Amplicon products were verified by 1.5% agarose gel electrophoresis in TAE buffer. PCR products were purified using QIAquick PCR purification kit (Qiagen, Germany), cloned into pTZ57R/T Vector (Thermo, USA)and sequenced by1st Base (Singapore). Nucleotide sequences of studied soft coral specimens registered to GenBank as accession numbers were listed in Table 1.

DNA Analysis:

Phylogenetic relationship of the 11 soft coral samples (Table 1) was analyzed based on referred sequences from Gene Bank using ClustalW method of Mega 3.1 (Kumar et al., 2004). Phylogeny reconstruction was done with tree

inferences using Neighbor Joining (NJ) method from Mega 3.1 with Bootstrap test of 5000 replicates. For species determination, homology level after aligning DNA sequences of the studied samples and references in the same clades on NJ trees obtained from phylogeny reconstruction analysis was estimated using Multiple Sequence Alignment method of DNAMAN 4.15 (LynnonBioSoft). Specific point mutations (SNPs) within *msh1* and 28S sequences were also indicated using Align of ClustalW and Sequence Data Explorer methods from Mega3.1.



Figure 1:- Collection sites of 11 soft coral specimens in the North Central of Vietnam.

Results and Discussion:-

Phylogenetic analysis

After sequencing and extracting, partial *msh1*mtDNA of 11 specimens and 28S rDNA of 10 specimens respectively ranged from 707-719bp and 739-753bp, of which fragments of 627-639bp *msh1* and 695-709bp 28S of the specimens were used for analysis due to the available length of related referred sequences from GenBank. Phylogenetic analyses based on *msh1*mtDNA sequence polymorphism of 11 specimens and referred taxa (Fig2) showed that all samples of this research belonged toAnthozoa class, Octocorallia Sub-class and Alcyonacea order. At lower taxa levels, they were divided into 4 sub-orders, 7 families and 7 genera according to 7 distinct clades (1 to 7) with bootstrap values from 99-100%. Phylogenetic tree of 28S rDNA (Fig3) were similar to that of *msh1*mtDNA, but only with 6 genera (without clade 4) because sequence of 28S fragment ofNL-07 was not available. Of 11 specimens, 5 were clustered into clades 2a and 2b with referred taxa of *Sinularia* genus. The results of phylogenetic analyses indicated the high taxonomical diversity of Alcyonaceancorals in sampling regions. Clade order between *msh1* and 28Swas not the same possibly because of difference in evolution rate between mtDNA and nuclear rDNA. Basically, mitochondrial genes evolve 50–100 times slower than nuclear genes in anthozoans (Hellberg, 2006; Chen et al., 2009). Some publications (McFadden et al., 2014, Prada et al., 2014, van Oppen et al., 2001) revealed that lack of concordance among different molecular markers in phylogenetic analyses is not uncommon in corals.



Figure 2:- Phylogenetic Neighbor Joining tree reconstructed based on DNA polymorphisms of 627-639bp *msh1* fragments of 11 soft coral specimens and related referred taxa.Numbers in open circles: clade number corresponding to taxa in particular families and sub-orders on the right of the tree. Numbers after referred taxa: accession no. from GenBank. Only bootstrap values more than 70% were represented next to the nodes of the tree.

On phylogenetic trees instructed by *msh1*and28S markers, taxa of 6 genera, including *Annella*, *Astrogorgia*, *Dichotella*, *Melithaea*, *Muricella* and *Scleronephthya* were monophyletic, while those of *Sinularia*were found polyphyletic with 2 separate clades 2a and 2b (Fig 2 and 3). The polyphyletic sub-clades of 2a and 2bmay be resulted from the hybridization among taxa in this genus, which was mentioned in some previous studies. Chemical markers of five new terpenoids isolated from *Sinularia maxima* and *S. polydactyla*elucidated the hybridization between these two soft coral species (Kamel et al., 2009). More recently, DNA barcode markers, *mtMutS* and 28S rDNA, proved the hybridization occurring within *Sinularia* genus (Quattrini et al., 2019). Analyses of *Sinularia*hybridization for clade 2b (Fig 3) based on 695-719bp 28S rDNA sequences of 10 soft coral specimens and related reference sequences resulted in seven SNPs along the aligned sequences. These SNPs could, therefore, add more evidence of hybridization of soft corals, which was one of the important motivations for evolution of *Sinularia* genus and Anthozoa class (Willis et al., 2006).

Species determination:

Analysis of homology levels

Alignments between sequences of msh1mtDNA and 28S rDNA fragments of 11 specimens and reference taxa in each clade on Fig 2 and Fig 3 showed the high homology level (99.7-100.0%) for both markers (Table 1). Five specimenswere classified as Sinularia brassica (HV-03, NL-06), S.leptoclados (SLE-05), Dichotellagenmacea (NL-03) and Annella reticulata (VM-02) based on msh1 marker with 99.7-100% of identity and 28S marker with 99.9-100% of identity), which was also in agreement with previously morphological identification. In case of unclassified by morphological criteria due to samples damaged, NL-08 and NL-05 specimens were still diagnostic at the genus level as Astrogorgiasp. and Melithaea sp., respectively. Msh1 markers with identity to referred taxa from 99.8-100% were in accordance with morphological analysis to identify specimens of SN-04 and SCO-05 to beSinulariananolobata and Sinulariaconferta, while 28S rDNA sequences of these two species were not available from GenBank. Though 28S fragments of HM-09 and Scleronephthyacorymbosawas aligned with 100% of identity, this specimen was still decided to be *Scleronephthya sp.* because morphological diagnosis classified this specimen only at genus level and msh1 sequence of S. corymbosawas unavailable in GenBank. Similarly, NL-07 was unclassified at species level in Muricella genus due to morphological identification to be Muricella sp. and identity of *msh1* sequence of this specimen and *Muricella sp.* referred in GenBankto be 100%. In short, among 11 specimens, seven of HV-03 andNL-06(Sinularia brassica), SLE-05(S.leptoclados), NL-03 (Dichotellagenmacea), VM-02 (Annella reticulata), SN-04(S. conferta) and SCO-05(S. nanolobata) were classified at species level, of which the first five wereby three markers (*msh1*, 28S and morphology) and the other two by *msh1* and morphology. The other four of NL-08, NL-05, HM-09 and NL-07 were only identified at genus level to be Astrogorgiasp., Melithaea sp., Scleronephthya sp. and Muricella sp. by either msh1-morphology or msh1-28S markers.



Figure3:- Phylogenetic Neighbor Joining tree reconstructed based on DNA polymorphisms of 695-709bp 28S fragments of 10 soft coral specimens and related referred taxa.Numbers in open circles: clade number corresponding to taxa in particular families and sub-order on the right of the tree. Numbers after referred taxa: accession no. fromGenBank. Only bootstrap values more than 70% were represented next to the nodes of the tree.

	Homology	sh1mtDNA*	Homology levels of 609-709bp 28S rDNA**					Morphological			
C*	Reference taxa		Taxa in thisstudy			Reference taxa		Taxa in thisstudy			classification
C	Taxa	Acc. No.	Taxa	Acc. No.	%	Taxa	Acc. No.	Taxa	Acc. No.	%	for species level***
C1	Scleronephthya sp.	DQ3028 25	HM- 09	MW0778 99	100. 0	Scleronephthya corymbosa	JX1243 50	HM- 09	MW0778 89	100. 0	Scleronephthya sp. (Studer, 1887)
C2	Sinularia brassica	HG9700 90	HV- 03	MW0779 00	100. 0	Sinularia brassica	KF915 491	HV- 03	MW0778 90	100. 0	Sinularia brassica (May, 1898)
а	Sinularia brassica	MH5166 94	NL-06	MW0779 03	99.7	Sinularia brassica	MF817 931	NL-06	MW0778 93	100. 0	Sinularia brassica (May, 1898)
	Sinulariaconfer ta	FJ62138 9	SCO- 05	MW0779 07	100. 0	Sinularia sp.	KC542 826	SCO- 05	MW0778 97	99.7	Sinulariaconfert a (Dana, 1846)
C2 b	Sinularialeptoc lados	KC5428 57	SLE- 05	MW0779 06	99.8	Sinularialeptocl ados	MF817 912	SLE- 05	MW0778 96	99.9	Sinularialeptocl ados (Ehrenberg, 1834)
	Sinulariananol obata	FJ62145 1	SN-04	MW0779 08	99.8	Sinularia sp.	KF915 519	SN-04	MW0778 98	99.9	Sinulariananolo bata (Verseveldt, 1977)
C3	Astrogorgiasp.	KF9155 66	NL-08	MW0779 04	100. 0	Astrogorgiasp.	KF915 326	NL-08	MW0778 94	99.8	NA
C4	Muricellasp.	KF8561 97	NL-07	MW0779 09	100. 0	Muricellasp.	NA	NL-07	NA	NA	Muricellasp. (Verrill, 1868)
C5	Dichotellagem macea	KF8036 69	NL-03	MW0779 01	100. 0	Dichotellagemm acea	JX2037 01	NL-03	MW0778 91	100. 0	Dichotellagemm acea (Milne Edwards &Haime, 1857)
C6	Annella reticulata	KP7139 84	VM- 02	MW0779 05	100. 0	Annella reticulata	KF915 314	VM- 02	MW0778 95	100. 0	Annellareticulat a (Ellis &Solander, 1786)
C7	Melithaea sp.	JX20379 9	NL-05	MW0779 02	99.8	Melithaea sp.	KC845 912	NL-05	MW0778 92	100. 0	NA

 Table 1:- Combination of molecular markers and morphological charactersfor species identification of soft coral specimens in this study.

*) C1-C7: clade 1-clade 7 in Fig. 2 and 3; **) H: homology level (%); ***) By Prof. Do Cong Thung

Single nucleotide polymorphism analysis

Separate alignment of *msh1* fragments (Fig 2) and 28S fragments (Fig3) of soft coral taxa respectively resulted in 174 and 188 single nucleotide polymorphisms (Fig4 and 5). These diagnostic SNPs distinguished each of 7 specimens to be classified at species level and each of 4 being identified at genus level (Table 1). Number of SNPs along *msh1* fragments of 11 taxa ranged from 0-50 (Fig 4) while 28S fragments of 10 taxa varied from 1-85 along them (Fig 5), revealing lower substitution rates of *msh1*mtDNA than those of 28S rDNA in corals as estimated by Hellberg (2006).Moreover, nucleotide substitution rate of the same coral taxon also varied depending on particular genomes. In this study, *Melithaea sp.* hasthe highest number of 50SNPs on *msh1* fragment, while the taxon is the second position with18 SNPs on 28S fragment. In contrast, *D. genmacea* had the highest number of 84 SNPs on *msh1* (0-9) and 28S fragments of 38 SNPs on *msh1* fragments of the species (Fig4 and 5). The lowest number of SNPs on *msh1* (0-9) and 28S fragments of *Sinularia* taxa in clade 2b again implied their hybridization. Difference in nucleotide substitution rate between *msh1* and 28S rDNA could interpret the difference in order of 7 clades between *msh1* and 28S phylogenetic trees of 11 specimen and related referred sequences (Fig 2 and 3).

! Title=msh1;									
NSites=151			1111	11111111111	1111111111	2222222222	22222222222	222233	
1	1112335	5566666777	8889990000	0111122244	4666777899	0012233344	5555666778	889900	
i	1280254064	7801257014	0450573678	9456823624	8078047828	1718934757	2348245160	231836	
#HM-09	TTTAAGGTCG	TGTACGAAAA	CCTAATGGAG	TAGATAG	CGCGCTTGGT	ACCAAATCCC	AAGCTGGGGG	TACTCT	
1.Scleronephthya sp (HM-09)		G			G				
2.Astrogorgia sp (NL-08)						T			
3.Annella reticulata (VM-02)	A.TA		CGC	CG.	A	GC			
4.Sinularia brassica (HV-03)	T				C			T.	
5.Sinularia brassica (NL-06)	T				C			т.	
6.Melithaea sp (NL-05)	A	.AG-C	AGA	CAC	TC	.GTA.	GG.T.AAAA.	C.AC	
7.Sinularia nanolobata (SN-04)					.A				
8.Sinularia leptoclados (SLE-05)A	c			ТТ.		c	.G	
9.Sinularia conferta (SCO-05)									
10.Dichotella gemmacea (NL-03)	GCC	GT-T.T.	.TCC	.CCCC.GG		TG. T	AA	C	
11.Muricella sp(NL-07)	CG	c			A-A	CG	c		
[33333333333	3333444444	4444444444	444444555	555555555555555555555555555555555555555	555555555555555555555555555555555555555	566666666	66666	Σ
[0112333477	8899001122	3334445666	7778999000	1113345566	6677777899	9000011122	23333	
[9081024938	5649693569	0234586256	1470159267	1465973802	8912349123	7456912313	82346	
#HM-09	TAGACCGTTG	TCTATTAGAT	ATAAGAGTAA	TCCTGCGGGG	TATACGTACG	TAATTTAAAA	GGATGACTCC	CATGT	
1.Scleronephthya sp (HM-09)				.c	CAC.	т	G.AC		11
2.Astrogorgia sp (NL-08)			G		A				5
3.Annella reticulata (VM-02)		CG	AG	T		T	T	т	20
4.Sinularia brassica (HV-03)		.TA		A.			-A-G	C	11
5.Sinularia brassica (NL-06)								C .	11
6.Melithaea sp (NL-05)		. T A		TA.			-A-G		
	AGA.TAAA.A	A	.A	TA. CCA	C.A	CACTG	-A-G	.GCA.	50
7.Sinularia nanolobata (SN-04)	AGA.TAAA.A	C.C.	.A	A. CCA	C.A	CACTG	-A-G	.GCA.	50 1
7.Sinularia nanolobata (SN-04) 8.Sinularia leptoclados (SLE-05	AGA.TAAA.A	C.C.	.A	A. CCA A	C.A 	CACTG 	-A-G	.GCA.	50 1 9
7.Sinularia nanolobata (SN-04) 8.Sinularia leptoclados (SLE-05 9.Sinularia conferta (SCO-05)	AGA.TAAA.A	C.C.	.A	A. CCA A	C.A	CACTG 	-A-G A 	.GCA.	50 1 9 0
 7. Sinularia nanolobata (SN-04) 8. Sinularia leptoclados (SLE-05) 9. Sinularia conferta (SCO-05) 10. Dichotella gemmacea (NL-03) 	AGA.TAAA.A	C.C.	.A	A. CCA A A AA	C.A .G GAA	CACTG G 	-A-G A 	.GCA.	50 1 9 0 38
 Sinularia nanolobata (SN-04) Sinularia leptoclados (SLE-05 Sinularia conferta (SCO-05) Dichotella gemmacea (NL-03) Muricella sp KF856197 (NL-07) 	AGA. TAAA.A)	C.C.	.A	A. CCA A A A AA	C.A 	CACTG 	-A-G A T. C.	.GCA.	50 1 9 0 38 18

Figure 4:- Analysis of SNPs on 627-639pb fragments of *msh1*mtDNA of 11 soft coral taxa.Numbers on the top: position of SNPs on *msh1* DNA sequences of particular taxon; A, C, T, G, mutation points; (-): either nucleotide or Indel; same nucleotide as that of HM-09 on the top; A, C, T, G, species specific mutation points.

! Title 288/									
NSites=150			1111	1111111111	1111111111	1111112222	22222222222	22222	
(2333334444	4556667889	9999990012	3344445556	6777777000	0099990011	3333444444	55556	
(5246090125	9064564351	2346890707	6035603671	7024569035	7001371260	0239234579	02343	
#HM-09	TGTGGCAGCG	CATCGCACGC	TGTCGACCCG	GCGCGCACGA	GCGCT-CTCG	TGTAGCCTGC	AGCGAGG	TCCTA	
1.Scleronephthya sp(HM=09)		CA C	T		$\cdots \cdots $		A		
2.Astrogorgia sp(NL-08)			T		· · · · · T · – · ·				
3.Annella reticulata(VM-02)	· · · · T · · · · · ·		6	λT	CT				
4.Sinularia brassica(HV-03)		T			.Τ λλΤ-Τ.	T	G		
5.5inularia brassica(NL-06)		T		– .A.	T. AAT-T.	T	G		
6.Melithaea sp(NL-05)	CAG-A,	CT	- T	.GG		. T .	T		
7.Sinularia sp(SN-04)	A A	T .C-	Tereset	· · · · · T - · · ·					
8.Sinularia leptoclados(SLE-05)			T T						
9.Sinularia sp(SCO-05)	T			–					
10.Dichotella_gemmacea(NL-03)	. TA T-C	G-ATAG	-ACGAGATAA	T TACT.G	A-CT T	GTATA.AG.A	-CGTTTCCTC	GGGGT	
(222222222222	33333333333	3333333444	444444444	444444555	55555555555	888888888888888888888888888888888888888	88888	Σ.
C C C C C C C C C C C C C C C C C C C	6667777888	1233333444	6667778334	46678888888	99999999000	0011111122	2222235556	66688	
	6781249256	0235609050	2595677041	0461004560	1345709013	4502567901	2456731560	12439	
· · · · · · ·				2401024204					
#ID4-09	CGAGACCCGT	ATGTCGACTG	AGCACCCGGC	CCAACTCTAA	ATGAGTGAGA	GCTCCTTGGG	GGTATGT	CAATT	
#HM-09 1.Scleronephthya_ap(HM-09)	CGAGACCCGT	ATGTCGACTG	AGCACCCGGC	CCAACTCTAA	ATGAGTGAGA	GCTCCTTGGG	GGTATGT	CAATT	11
#HM-09 1.Scleronephthya sp(HM-09) 2.Astrogorgia sp(NL-08)	CGAGACCCGT	λΤGTCGACTG λ	AGCACCCGGC A	CCAACTCTAA	ATGAGTGAGA	GCTCCTTGGG TT C.T	GGTATGT TATATC.C	CAATT . GG	11 21
#IM-09 1.Scleronephthya sp(HM-09) 2.Astrogorgia sp(NL-08) 3.Annella_reticulata(∀M-02)	CGAGACCCGT	ATGTCGACTG AA	AGCACCCGGC A G TT	CCAACTCTAA	ATGAGTGAGA	GCTCCTTGGG TT C.T	GGTATGT TATATC.C	GG	11 21 10
#HM-09 1.Scleronephthya ap(HM-09) 2.Astrogorgia sp(NL=08) 3.Annella reticulata(VM=02) 4.Sinularia brassica(HV-03)	CGAGACCCGT	ATGTCGACTG A	AGCACCCGGC A G TT 	CCAACT CTAA	ATGAGTGAGA	GCTCC TTGGG 	GGTATGT TATAT-, C, C	GG	11 21 10 15
#IM-09 1.Scleronephthya ap(HM-09) 2.Astrogorgia ap(NL=08) 3.Annella reticulata(VM=02) 4.Sinularia brassica(HV-03) 5.Sinularia brassica(NL-06)	CGAGACCCGT	ATGTCGACTG A	AGCACCCGGC λ	С СААСТСТАА 	Аталаталал 	GCTCCTTGGG TT AT-A AT-A	GGTATGT TATATC.C. λ	GG	11 21 10 15
#IM-09 1. Scleronephthya ap(IM-09) 2. Astrogorgia ap(NL-08) 3. Annella reticulata(VM-02) 4. Sinularia brassica(NV-03) 5. Sinularia brassica(NL-06) 6. Melithea ap(NL-05)	CGAGACCCGT	ATGTCGACTG 	AGCACCCGGC G TT	С СААСТСТАА 	Аталаталал 	GCTCCTTGGG TT =A. AT-A AC-T	GGTATGT TATATC.C A	GG	11 21 10 15 15
#IM-09 1.scleronephthya ap(IM-09) 2.Astrogorgia ap(NL-08) 3.Annella reticulta(VM-02) 4.Sinularia brassica(NL-06) 5.Sinularia brassica(NL-06) 6.Melithaea ap(NL-05) 7.Sinularia ap(SN-04)	CGAGACCCGT	ATGTCGACTG -CA 	AGCACCCGGC G A A A	ССААСТСТАА 		GCTCCTTGGG .TT. AT. AT. AT. 	GGTATGT TATATC.C λ λ	CAATT .GG 	11 21 10 15 15 18
#IM-09 1. Scleronephthya ap(IM-09) 2. Astrogorgia ap(NL-08) 3. Annella reticulata(VM-02)) 5. Ainularia prassica(NL-06) 6. Melithaea ap(NL-05) 7. Sinularia ap(SN-04) 0. Sinularia ap(SN-04)	СGAGACCCGT ТТ.АА Т	ATGTCGACTG -CA T T T	λgcλcccggc ,,,,,,,, .	.T	Аталаталал 	GCTCCTTGGG 	GGTATGT TATATC.C λ λ	CAATT .GG 	11 21 10 15 19 10
#IM-09 1. Scleronephthya ap(IM-09) 2. Astrogorgia ap(NL-08) 3. Annella reticulta(VM-02) 4. Simularia brassica(NV-03) 5. Simularia brassica(NL-06) 6. Melithaea ap(NL-05) 7. Simularia ap(SN-04) 8. Simularia ap(SC0-05) 9. Simularia ap(SC0-05)	CGAGACCCGT	ATGTCGACTG A -CA T T C.	AGCACCCGGC 		Аталаталал 	GCTCCTTGGG .T.T.T. C.T ATA .AT-A GC-T	GG TATGT TATATC.C A A A	CAATT .GG CC	11 21 15 15 18 10 1 2
#IM-09 1. Scleronephthya ap(IM-09) 2. Astrogorgia ap(NL-08) 3. Annella reticulta(VM-02) 4. Sinularia prassics(IV-03) 4. Sinularia prassics(IV-03) 5. Molithaa ap(NL-05) 6. Sinularia sp(SR-04) 6. Sinularia sp(SR-04) 9. Sinularia sp(SC0-05) 10. Dichotella gemmacea(NL-03)	CGAGACCCGT TT.AA 	ATGTCGACTG 	AGCACCCGGC 	ссалстстал 		GCTCCTTGGG . T. T. 	GG ТАТGТ ТАТАТ-, С. С 	CAATT .GG 	11 21 15 15 19 10 285

Figure 5:- Analysis of SNPs on 695-709bp fragments of 28S rDNA of 10 soft coral taxa.Numbers on the top: position of mutation points on 28S sequences of particular taxon; A, C, T, G, mutation points; (-): either nucleotide or Indel; same nucleotide as that of HM-09 on the top; A, C, T, G, species specific mutation points.

Msh1 and 28S markers supporting morphological clasification in determination of soft coral taxa:

Recently, classification of corals still has been based on morphological characters of colony morphology and/or sclerites comparison. In fact, the first approach was unable to determine at species level in some genera for mixture of morphological characters among them, while their sclerite were so varied that many species were not identified if applied skeleton composition comparison (van Ofwegen and Groenenberg, 2007). Similarly, both approaches were unable to identify some octocoral taxa or were incongruent with morphospecie identification due to high variation, especially taxa in such genus as *Sinularia*,within which the hybridization between closely related taxa happened (Benayahu et al., 2012; McFadden et al., 2014; 2017; Quattrini et al., 2019). Molecular data such as mtDNA and rDNA could support to overcome such limitation in coral classification (Reijnen, 2015; Reijnen and Van der Meij, 2017). Besidesnuclear DNA, loci from mtDNA such as *msh1*, *MutS*, *ND2*, *16S* ... were available in GenBank and widely used as markers for identification of numerous octocoral taxa (McFadden et al., 2010). However, in some cases, sequences of mitochondrial loci were not informatively polymorphic enough to identify coral taxa at species level due to their slow rates of evolution (Shearer et al., 2002; Huang et al., 2008). Only 80% of *Sinularia*species were in agreement with morphospecies when identified using *MutS*mtDNA (McFadden et al., 2014). They also concluded that a single DNA marker, e.g.,*mtMutS*was not powerful enough to discriminate most closely related species such

asSinularia spp. Loci from rDNA genome, i.e., 18S, 1TS, 5.8S and 28S were shown to be more powerful than those from mtDNA to resolve relationships of corals below genuslevel (van Ofwegen and Groenenberg, 2007). Therefore, markers from mtDNA could be combined with those from rDNA to discriminate coral taxa at species level, especially closely-related species within most coral genera (Benayahu et al., 2018). In this study, results of determination of 11 soft corals by *msh1* and 28S markers were in agreement to those by morphological classification (Table 1), again indicating the strong support of the two DNA markers to morphospecies of these soft coral species. In spite of powerfulness in discrimination of coral taxa, including closely-related species in a genus, molecular markers including *msh1* and 28S loci showed some disadvantages. Of all specimens, five were identified at species level by all *msh1*, 28Sand morphological markers. However, the other 6 specimens were determined at species level or only genus level with the pair marker of *msh1*- morphospecies, *msh1-28S* due to unavailability of related *msh1*mtDNA and 28S rDNA in GenBank. When related sequences of these two markers are available, these genus-level taxa will be resolved.

Divergence of gorgonian corals:

The molecular analysis in this study indicated the high diversity of gorgonian corals in Alcyonaceaorder along the North Central coast of Vietnam (Fig 1). All 11 specimens were classified to be 10 species (Table1) in 7 genera, 7 families and 4 suborders (Fig2and 3). Of 11 specimens, fivebelonged to Sinularia, indicating the richness of the species in this genus. Vietnam occupied as long as 3000km of tropical coastal length, within which high diversity of marine animal including soft coral has been investigated.Ben et al.(2010) had found a total of 60 taxa including 10 genera and 5 families from 85 collected specimens in Ly Son Island(30 km from the SouthCentral coast of Quang Ngai province), of which 14 species of Sinularia, 9 species of Lobophytum, 6 species of Sarcophyton genera were newly recorded in Vietnam. After that, a phylogenetic investigation grouped 11 soft corals in Nha Trang Bay (KhanhHoa province) grouped into four different clades with different gorgonian coral genera of Sinularia, Sarcophyton, Lobophytum and a mixed betweenSarcophyton and Lobophytum(Lien et al., 2015). More recently, Ben et al. (2020) also found species richness of soft corals along Cu Lao Cham Marine Protected Area (South Central coast of Quang Nam province), in which a total of 45 taxa belonging to 12 genera and 7 familieswere identified. Among them, Sinularia genus was the highest diversity group with 19 species, followed by Sarcophyton with 8 species and Lobophytum with 6 species. However, it is the fact that he above findings have been only spotty trying. Recently, molecular markers, especially DNA barcodes have been proved really powerful to resolve various problematic aspects of marine animals. An overall project on application of these tools to deal with phylogenetic patterns, species determination and diversity research on marine animalsincluding soft coralsalong 3000km coastal length of Vietnam, could be, therefore, programmed in order to effective conservation and sustainable development of these resources in the country.

Conclusion:-

In this study, the developed molecular markers clearly supported the classification of soft corals at species level. The identifications of 9 out of 11 soft coral specimens based on *msh1*mtDNA and 28S rDNA polymorphism of the soft coral specimens and referred taxa werein concordance with the traditional taxonomy. There were 7 specimens to be classified at species level, of which 5 were identified *msh*, 28S and morphological markers and the other two by *msh1* and morphospecies. The other four were only identified at genus level by either *msh1*-morphology or *msh1-28S* marker pairs. All of the taxa belonged toAnthozoa class, Octocorallia sub-class and Alcyonacea order, from which they were divided into 4 sub-orders, 7 families and 7genera according to 7 distinct clades on phylogenetic trees with high confidence intervals from 99-100%.

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