

 <p>ISSN NO. 2320-5407</p>	<p>Journal Homepage: -www.journalijar.com</p> <h2 style="text-align: center;">INTERNATIONAL JOURNAL OF ADVANCED RESEARCH (IJAR)</h2> <p style="text-align: center;">Article DOI:10.21474/IJAR01 DOI URL: http://dx.doi.org/10.21474/IJAR01</p>	 <p>INTERNATIONAL JOURNAL OF ADVANCED RESEARCH (IJAR) ISSN 2320-5407</p> <p>Journal homepage: http://www.journalijar.com Journal DOI:10.21474/IJAR01</p>
---	---	--

RESEARCH ARTICLE

**International Conference on Recent Advances in Biotechnology, Biomolecules and Pharmacy RABBP – 2020
(Organized in Virtual Mode due to COVID-19 Pandemic) during 17th to 19th December 2020 at KL
University, Vijayawada, Andhra Pradesh, India.**

PHYTOCHEMICAL SCREENING OF *MICROSTYLIS WALLICHII* ROOT EXTRACT

**Babitha Sri Majeti, Navya SaiMedikonda, Rohitha Akula, Haritha Bommepali, Rishi Priya Peddamallu and Praveen
Kumar Vemuri**

Department of Biotechnology, Koneru Lakshmaiah Education Foundation, Guntur, Andhra Pradesh, India.

Manuscript Info

Key words:-

Microstylis wallichii, Antioxidant Activity,
Anti-Microbial Activity, Phytochemical
Screening

Abstract

Alternative medicine has become popular these days as it is gaining practices across the globe. Ayurvedic medicine is one of the important forms of alternative medicine that was widely available in India. The present study mainly focuses on the identification of therapeutic properties of *Microstylis wallichii*. The ethanolic extract of *Microstylis wallichii* roots are used for its anti-oxidant and antimicrobial activity. *Microstylis wallichii* dried roots shown good anti-oxidant and anti-microbial properties. The ethanolic extract of *Microstylis wallichii* was checked for anti-microbial activity against pathogenic bacteria such as *E. coli*, *Staphylococcus aureus*, *Pseudomonas*.

Copy Right, IJAR, 2020., All rights reserved.

Introduction:-

The medicinal plants find application in pharmaceutical, cosmetic, agricultural and food industry. The use of the medicinal herbs for curing disease has been documented in history of all civilizations. Man in the pre-historic era was probably not aware about the health hazards associated with irrational therapy [1]. With the onset of research in medicine, it was concluded that plants contain active principles, which are responsible, for curative action of the herbs. Before onset of synthetic era, man was completely dependent on medicinal herbs for prevention and treatment of diseases by using active constituents of the medicinal [2]. With introduction of scientific procedures, the researchers were able to understand about toxic principles present in the green flora [3-5]. The efficacy of some herbal products is beyond doubt, the most recent examples being Silybummarianum (silymarin), *Artemisia annua* (artemesinin) and *Taxus baccata* (taxol) [6-8]. On the other hand, randomized, controlled trials have proved the efficacy of some established remedies, for instance, Ginkgo biloba for tinnitus [9]. In Hypericum some researchers are of the view that hypericin is the active principle of the herb and some believe that hyperforin is responsible for antidepressant action of the herb [10]. Recently research has supported biological activities of some medicinal herbs. Cancer is such a segment where researchers are expecting new molecules from herbs that can provide us with tools for fighting this dreaded disease [11]. The roots and woody portion have been used traditionally for curing various ailments like stomach pains, fever, venereal disease, rheumatism and act as a blood purifier [12] and it also possesses antioxidant, antileprotic effects [13].

Corresponding Author:-Babitha Sri Majeti

Address:-Department of Biotechnology, Koneru Lakshmaiah Education Foundation, Guntur,
Andhra Pradesh, India.

Methodology:-**Plant Material**

Microstyliswallichii root material was selected and made as smoothie for phytochemical screening studies. Root material was dried, powdered and then used for the studies. The procedures recommended in Indian Pharmacopoeia and WHO guidelines were followed to calculate the physico-chemical constants[15].

Extraction using ethanol and water

Five grams of dried coarse powder of roots were macerated with 100ml of 90% ethanol in a closed flask for 24hrs, shaken frequently for 6 hours and allowed to stand for 18hrs. Filtered immediately taking precautions to prevent loss of ethanol. 25ml of the filtrate was evaporated to dryness in a tarred flat bottomed shallow dish. The residue was dried at 105°C and weighed. The percentage of ethanol soluble extraction was calculated with reference to air dried drug. Five grams of coarse powder was weighed and dissolved in 100ml of water in a stoppered flask, heated at 80°C, shaken well and allowed to stand for 10min. It was cooled; 2gms of kieselghur was added and filtered. 5ml of the filtrate was transferred to a tarred evaporating dish and the solvent was evaporated on a water bath. The percentage of water-soluble extraction was calculated with reference to air dried drug.

Determination of crude fiber content

About Two grams of the drug was accurately weighed and extracted with ether. Then 200ml of 1.25% sulphuric acid was added and boiled for 30min under reflux. It was filtered and washed with boiling water until free of acid. The entire residue was rinsed back into flask with 200ml of boiling 1.25% sodium hydroxide solution and again boiled under reflux for 30min. The liquid was quickly filtered and the residue was washed with boiling water until neutral, dried at 110°C to constant weight. It was then ignited to 30min at 60°C, cooled and weighed. The percentage of crude fiber content was calculated with reference to the air-dried drug.

Determination of loss on drying

Glass stoppered shallow bottle was weighed that had been dried in the same conditions to be employed in the determination. About one gram of the sample was transferred to the bottle and distributed evenly by gently side wise shaking to a depth not exceeding 10mm. Place the loaded bottle in a drying chamber (the stopper was removed and left in the chamber). The sample was dried to a constant weight and allowed to cool. The bottle along with the content was weighed. The process was repeated until the successive weights differed not more than 0.5mg (drying to constant weight). The percentage loss of weight was calculated with reference to the air-dried drug.

Determination of foaming Index

1gm of the coarsely powdered drug was weighed and transferred to 500ml conical flask containing 100ml of boiling water. The flask was maintained at moderate boiling at 80-90°C for about 30min. It was cooled, filtered into a volumetric flask and sufficient water was added through the filter to make up the volume to 100ml. Ten stoppered test tubes were cleaned (height 16cm, diameter 1- 6cm) and marked from 1 to 10. 1, 2, 3ml up to 10ml of the filtrate was measured and transferred to each tube and adjusted the volume of the liquid with water to 10ml. Then the tubes were stoppered and shaken lengthwise for 15sec uniformly, allowed standing for 15min the length of the foam was measured in each tube. If the height of the foam in each tube is more than 1cm, the foaming index is more than 1000. In this case, 10ml of the first decoction of the root material is measured and transferred to a 100ml volumetric flask (V2) and the volume is made to 100ml and followed the same procedure.

Fluorescence Analysis

The fluorescence analysis of the drug powder as well as various extracts was carried out by using the method of Chase and Pratt. The behavior of the powder with different chemical reagents was also carried out.

Results and Discussion:-**Ash analysis and moisture contents**

Plant species with medicinal properties have been playing a fundamental role in the efforts for drug discovery all over the world. 80% populations in developing countries are dependent on plants for their primary health care, and despite the significant progress in the field of synthetic organic chemistry of the twentieth century, more than 25% of prescribed medicines in developed countries are derived directly or indirectly from plant sources.

Table 1:- Phytochemical studies.

S.No	Secondary metabolites	Methanol	Ethyl acetate	Ethanollic	Aqueous
1	Steroids	+	-	+	-
2	Triterpenes	+	+	+	-
3	Saponins	-	-	+	++
4	Tri terpinoidalsaponins	+	-	++	-
5	Alkaloids	+	+	+	+
6	Carbohydrates	-	-	+	-
7	Flavonoids	+	+	++	+
8	Tannins	+	+	+	+
9	Glycosides	-	-	+	-
10	Polyphenols	+	+	++	+

Extraction values

Extractions of crude powder drug with different solvents gives different extraction values. Extraction value is one of the useful methods for evaluation of crude drugs, and provides guidance about the most suitable solvent to be used for extraction, and also helps in detecting various types of adulteration and exhausted materials i.e., Water and alcohol soluble extraction values can be used for the detection of adulterants, defective processing and poor quality of the drug while petroleum ether soluble extraction value indicates lipid contents present in the crude drugs. The fluorescence analysis of powder with various reagents and extracts are given in the Table 3 and 4.

Table 2:- Fluorescence analysis of powder.

S.No	Reagents	Day light	Short UV	LongUV
1. Powdered root (PR)	Red		Red Dark	Red
2 PR + 1 N HCl	Yellow		Red	Red
3 PR + 1 N NaOH	Red		Red	Pale Red
4 PR + 50% HCl	Yellow		Fluorescent Red	Fluorescent Red
5 PR + 50% H ₂ SO ₄	Dark Red		Dark Red	Dark Red
6 PR +50% HNO ₃	Dark brown		Brown	Reddish brown
7 PR + ethanol	Red Fluorescent	Red		Light Red
8 PR + ethanol + 1 N NaOH	Red		Brown	Red

Conclusion:-

Phytochemicals present in the different extracts of roots of *Microstyliswallichii* was identified as prominent source for anti-oxidant property. Among the extracts ethanolic extract has highest anti-oxidant property when compared to other extracts. In the present study it was found that *Microstyliswallichii* roots ethanolic extract has an excellent antimicrobial activity. The pathogenic bacteria were inhibited in presence of the root extracts of *Microstyliswallichii*. Therefore, the future studies should be aimed to exploit this plant to be used as one of the best medicinal plants is controlling pathogenic bacteria.

References:-

1. Singh AP. Promising phytochemicals from Indian medicinal plants. Ethnobotanical leaflets. 2005;2005(1):18.
2. Chang CL, Lin Y, Bartolome AP, Chen YC, Chiu SC, Yang WC. Herbal therapies for type 2 diabetes mellitus: chemistry, biology, and potential application of selected plants and compounds. Evidence-Based Complementary and Alternative Medicine. 2013 May 13;2013.
3. Nigam S. Tephrosia purpurea: From traditional use to scientific assessment: A Review. Environ Sci Ecol: Curr Res. 2021;2:1020
4. Cunningham SD, Berti WR. Remediation of contaminated soils with green plants: an overview. In Vitro Cellular & Developmental Biology-Plant. 1993 Oct 1;29(4):207-12.
5. Mergen F. A toxic principle in the leaves of Ailanthus. Botanical Gazette. 1959 Sep 1;121(1):32-6. Post-White J, Ladas EJ, Kelly KM. Advances in the use of milk thistle (Silybum marianum). Integrative cancer therapies. 2007 Jun;6(2):104-9.

7. Duke SO, Vaughn KC, Croom EM, Elsohly HN. Artemisinin, a constituent of annual wormwood (*Artemisia annua*), is a selective phytotoxin. *Weed Science*. 1987 Jul;35(4):499-505.
8. Ramawat KG, GOYAL S, KUMAR N, Sonie KC. Natural products of plant origin with therapeutic potentiality and their biotechnological production. *Plant Tissue Culture and Molecular Markers*. 2009:247.
9. Izzo AA, Hoon-Kim S, Radhakrishnan R, Williamson EM. A critical approach to evaluating clinical efficacy, adverse events and drug interactions of herbal remedies. *Phytotherapy Research*. 2016 May;30(5):691-700.
10. Kirakosyan A, Sirvent TM, Gibson DM, Kaufman PB. The production of hypericins and hyperforin by in vitro cultures of St. John's wort (*Hypericum perforatum*). *Biotechnology and applied biochemistry*. 2004 Feb;39(1):71-81.
11. Renukadevi KP, Sultana SS. Determination of antibacterial, antioxidant and cytotoxicity effect of *Indigofera tinctoria* on lung cancer cell line NCI-h69. *Int J Pharmacol*. 2011 Mar;7(3):356-62.
12. Vaidyanathan D, SenthilkumarMS, Basha MG. Studies on ethnomedicinal plants used by malayali tribals in Kolli hills of Eastern ghats, Tamilnadu, India. *Asian J Plant Sci Res*. 2013;3(6):29-45.
13. Kalpana T, Karunakar N, Reddy MS, Prabhakar MC, Krishna DR. Assessment of antioxidant activity of some antileprotic drugs. *Arzneimittelforschung*. 2001 Aug;51(08):633-7.