

Journal homepage: http://www.journalijar.com

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH

RESEARCH ARTICLE

Study on Some Pathological aspects of Salmonella enteridis Experimental Infection in Mice

Khalil H. Al-Jeboori¹, Laheeb J. Al-Hashimi¹ and Haitham I. Baqir²

- 1. Dept. of Pathology, College of Veterinary Medicine, Univ. of Baghdad, Iraq.
- 2. Scientific Researcher, Central Public Health Laboratory, Ministry of Health, Iraq.

Manuscript Info

Manuscript History:

Received: 02 September 2014 Final Accepted: 15 October 2014 Published Online: November 2014

.....

Key words

S. enteridisexperimental pathological lesions.

*Corresponding Author

Khalil H. Al-Jeboori

Abstract

Infections with *Salmonella enteridis*consider the main reason of food poisoning in man and infect other animals species and avian. For the study of pathological lesions caused by *S. enteridis* in mice. A local strain of this microbial agent were obtained from the central public health laboratory. Reidentification for this bacterial strain were done using, biochemical test, API – 20, BiomerieuxVitek instrument and slide agglutination test. The LD50 were 1.4*10⁶ CFU, then this LD50 dose were I/P inoculated in thirty three mice and eleven mice inoculated with phosphate buffer saline (pbs) as a control group. Three mice were sacrificed every three days intervals for 40 days. Among the pathological lesions were micro abscesses and granulomas seen in liver along the experimental periods. Amyloid, lymphoid hyperplasia and different pathological inflammatory lesions were recorded in liver, gall bladder, spleen, pancreas, lungs, intestine, kidneys, heart, brain and meninges, lymph nodes and in periton.

.....

Conclusion: *S. enteridis* experimental infection gave different pathological lesions in the different organs.

Copy Right, IJAR, 2014,. All rights reserved

Introduction

Infections with Salmonella enteridisconsider the main reason of food poisoning in man causing Gastroenteritis and infect other anmals and avian species including different pathological disorder (1). In certain cases these microbial agents colonize the avian intestine as in apparent carriers and during fecal excretions, it transmitted to other birds and meat contamination occurred (1, 2). S. enteridisconsidered the intra and extra – cellular invasive pathogen pass through ileum epithelia into peyer'spatches and through blood or lymphatic vessels reach into other organs (3). Murine salmonellosis is a common disease caused by S. enteridisand S. typhimurium, associated with sever enteritis and septicemia in other internal organs (4). For the importance of S. enteridisin human and animals causing different pathological disorders; this study aimed to identify the different pathological lesions associated with experimental infection of mice with this microbial agent.

Materials and Methods

A local strain of *Salmonella enteridis* type D obtained from the central public health laboratory. The strain was re identified again as a *S. enteridis* using biochemical tests, API 20 E together with BiomerieuxVitekinstrument and slide agglutination test. The lethal dose -50 (LD -50) for this microbial agent was $1.4*10^6$ CFU according to (5) methods.

Animals Inoculation:

Forty four mice were taken and thirty three were inoculated I/P with 0.25 ml of S. enteridissuspension (1.4*10⁶ CFU of bacterial suspension) and eleven mice were inoculated I/P with 0.25 ml of phosphate buffer saline (pbs) as a control group. Three mice were sacrificed at three days intervals and one mouse from the control group along the experiment period (40 days). Pieces of pathological lesions were fixed in 10% neutral buffered formalin, processed

routinely, cut at 5 μ m thickness and stained with hematoxylin and eosin (H&E) and examined under light microscope (6).

The Results and Discussion Histopathological Changes: The Liver:

The lesions began as a micro abscesses composed of the multifocal infiltration of the neutrophils in the liver parenchyma (**Fig** – **1**) and adjacent the blood vessels (central vein and in the portal areas), the neutrophils were replaced by mononuclear cells (lymphocytes and macrophages) during the 2^{nd} week to form early granulomatous during 3^{rd} , 4^{th} week of experimental infection. Amyloid infiltration were seen in the sinusoidal walls later on (**Fig** – **2**).

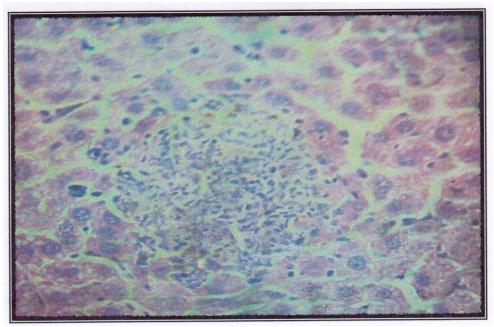


Fig .1: Liver tissue: showed microabscess composed of neutrophils infiltration (HxE) X 400

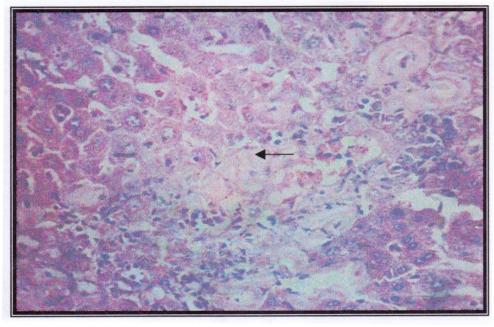


Fig.2: Liver: tissue infiltrated with amyloid .(HxE) X 400

The Gall bladder and Bile ducts:

The similar neutrophils infiltration were seen in the lumen of gall bladder and bile duct (**Fig** - 3), these cellular infiltrates were replaced by mononuclear cells at 2^{nd} week and at 3^{rd} week the mononuclear cells together with fibroblasts were seen in the gall bladder wall with sloughing of their epithelial lining causing chronic cholecystitis.

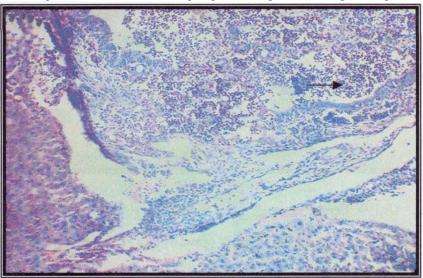


Fig-3: Gall bladder tissue: showed filling their lumen with plus exualade (HxE) X200

The Spleen:

There is extensive lymphoid hyperplasia of white pulp and sever congestion of the red pulp and accentric arterioles of the white pulp. Amyloid infiltrations were seen in the white pulp during the 3^{rd} and 4^{th} weeks postinoculation(**Fig** – **4**).

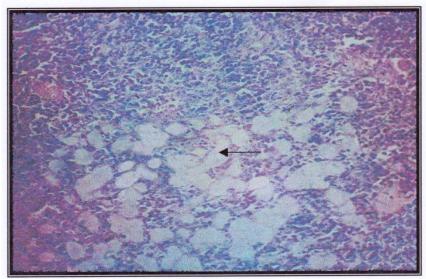


Fig – 4 :S.plenic tissue : showed lymphoid hyperplasia of white pulp and amyloid infiltration ($Hx\ E$) x 200 The Pancreas:

There is infiltration of neutrophils in the peripancreatic tissue and in the pancreas causing distraction of some pancreatic acini($\mathbf{Fig} - \mathbf{5}$). The neutrophils were replaced by mononuclear cells (lymphocytes and macrophages) during the 2^{nd} week postinoculation and during the 3^{rd} week postinoculation the mononuclear cells and fibroblasts were replaced the infected pancreatic tissue.

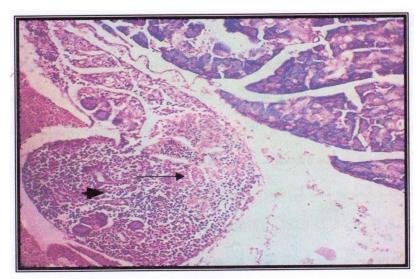


Fig-5 : pancreatic tissue: showed infiltration of neutrophils replacing their acini (HxE) x200 The Lungs:

There is peribronchiallymphoid tissue hyperplasia and interstitial pneumonic lesion began as infiltration of neutrophils and congestion of alveolar capillaries the neutrophils were replaced by lymphocytes and macrophages at 2^{nd} week together withfibroblasts at the 3^{rd} week(**Fig** – **6**).

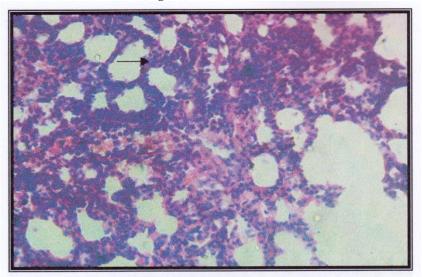


Fig -6: lung tissue: showed interstitial pneumonia (Hx E) x 200

The Heart:

There is infiltration of neutrophils in adipose tissue surrounded the heart and in the myocardium (Fig - 7), the neutrophils were replaced by mononuclear cells and fibroblasts causing focal epicarditis and myocarditis.

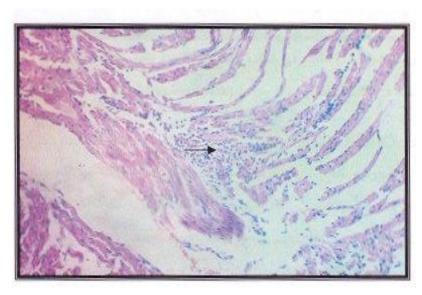


Fig-7: Heart: showed neutrophils inflltration in myocardium (HxE) x 200

The Lymph nodes:

There is extensive lymphoid hyperplasia of cortical region with the secondary lymphoid follicles with germinal centers formation ($\mathbf{Fig} - \mathbf{8}$). Also reticuloendothelialcell hyperplasia of themedulary sinuses and neutrophils infiltration in the capsule.

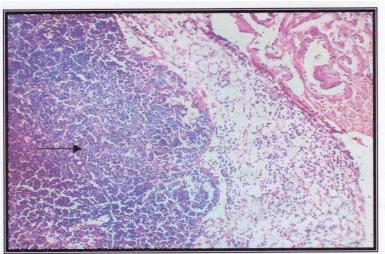


Fig-8: lymphoid tissue: showed extensive , lymphoid hyperplasia and infiltration of neutrophils in the capsule (HxE) x 200

The Kidney:

There is neutrophils, lymphocytes and macrophages infiltration at the renal interstitial tissue (Fig - 9). The similar cellular infiltrations were seen in the renal pelvis and in the renal capsule and inperiureteric area. Amyloid infiltrations were seen in the glomeruli (Fig - 10).

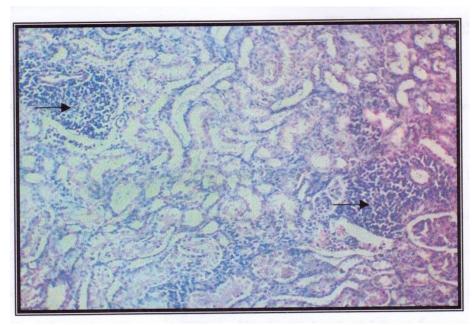


Fig-9 : Renal tissue: showed infiltration of neutrophils and mononuclear cells in the renal interstitial tissue (HxE) x 200

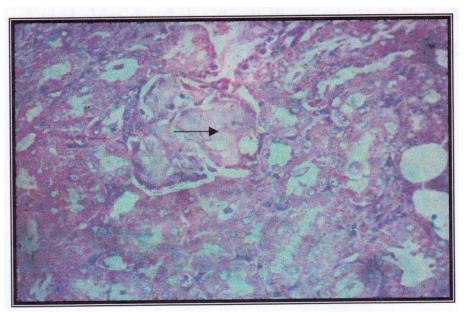


Fig-10: Renal tissue: showed $\,$ infiltration of amyloid in the glomeruli $\,$ ($\,$ HxE) x400 $\,$

The Intestine:

There is infiltration of neutrophils lymphocytes and macrophages at the intestinal mucosa. Also, these cellular infiltrations together with mucin were seen in intestinal lumen. In chronic cases sloughing of intestinal epithelia and intestinal wall fibrous replacing the intestinal mucous glands (Fig - 11).

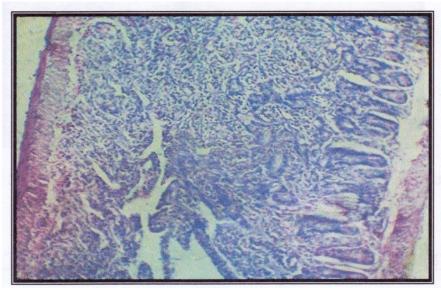


Fig-11: Intestinal tissue: showed infiltration of mononuclear cells and fibroblasts in the intestinal wall and sloughing of their mucosal epithelia (HxE) x 200

The Brain and Meninges:

There is perivascular leukocytic cuffing (Fig-12) in the brain and meninges, focal gliosis and neural degeneration.

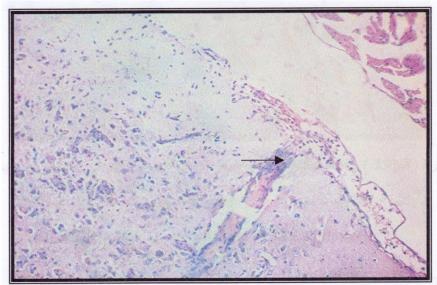


Fig -12: Brain : showed congestion of their blood . vessels and perivascular cuffing $\,$ ($\,$ HxE) x 200. The Peritoneum:

There is infiltration of neutrophils which replaced by lymphocytes and macrophages at the 2^{nd} week and these inflammatory cells together with fibroblasts were seen in peritoneal tissue. Also thrombosis (**Fig** – **13**) of peritoneal blood vessels.

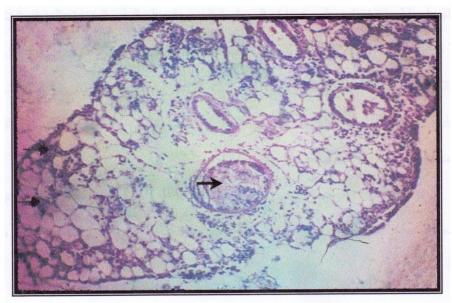


Fig-13 :periton: showed infiltration of neutrophils and thrombosis of their blood vessels (HxE) x 200.

The infection with *S. enteridis*associated with neutrophils infiltration due to the *S. enteridis*endotoxins which attract the neutrophils in all the infected organs, similarly reported by (7). During the 2nd and 3rd weeks, these neutrophils infiltration transform into early and chronic granuloma which seen in liver similarly reported by (8, 9, 10, 11, 12). During the experimental infection by *S. typhimurium S. typhi* oby *S. paratyphi* A, B. lymphoid hyperplasia in the lymph nodes and in the spleen were reported by (9) following experimental infection with these microbial agents due recurrent immunological stimulation by microbial endotoxins. Amyloid infiltration in different organs such as liver, spleen,kidney were also reported in other animal species following experimental salmonella infection (13) similar lesions were reported in animals used for antisera preparation (14). Other lesion such as encephalitis and meningitis were reported in children due to *S. enteritis* (15). Other important lesions such cholecystitis, cholangitis, enteritis, pancreatitis, pneumonia, myocarditis and interstitial nephritis were seen in the mice in this study were also reported by other markers (10, 11,12) indicated that these bacterial agents were invasive bacteria infect the all organs and cause the various lesions similarly reported for other salmonella species during the natural and experimental infection in the different animals species (8, 13).

The References:

- 1. Barrow, P.A; Huggins, M.B.; Lovel, M.A. and Simpson, J.M. (1987). Observations on the pathogenesis of experimental *Salmonella typhimurium* infection in chickens Res. Vet. Sci. 42:194 199.
- **2. Barrow, P.A; Hassan, J.O. and Berchieri, A. Jr. (1990).** Reduction in fecal excretion of *Salmonella typhimurium*strain F98 in chickens vaccinated with live and killed *S. typhimurium*organisms. Epidemiol. Infect. 104(3) 413 426.
- **3. Shahin, A.** (2005). Investigation of the humoral and cellular immune responses of chickens to *Salmonella typhimurium* live vaccine. Ph. D. Thesis. Faculty of Veterinary Medicine. Ludwig Maximillans University, Munich.
- **4. Rubin, R; H. and Weinstein, L. (1977).** Salmonellosis, microbiological, pathological and clinical features. Stratton intercontinental Medical Book Corp. New York.
- 5. Dixon, W.J. (1980). Efficient analysis of experimental observations. Ann. Res. Pharmacol. Toxicol. 20:441 462.
- **6. Luna, L.G.** (1968). Manual of histological staining methods of the Armed Forces Institute of pathology, 3rd. ed. McGraw Hill Book Company, USA.
- Islam, L; Nabi, A; Ahmed, K. and Sultan. N. (2002). Endotoxins of enteric pathogens are chemotactic factor of human neutrophils. J. Biochem. Mol. Biol. 35(5)482 – 487.
- 8. Nakoneczna, I. and Hsu, H. S. (1980). The comparative histopathology of primary and secondary lesions in murine Salmonellosis Br. J. Exp. Pathol. 61:76 84.
- 9. Nakoneczna, I. and Hsu, H. S. (1983). Histopathological study of protective immunity against murine salmonellosis induced by killed vaccine. Infect. Immun. 39(1) 423 430.
- **10. Al-Jeboori, K. H.** (1997). Bacteriological, Immunological and pathological parameters associated with *Salmonella typhi* infection in man, mice and Guinea pigs. Ph. D. Thesis college of Veterinary Medicine, University of Baghdad, Iraq.

- **11. Al-Jeboori, K. H.** (2000). A study on the bacterial dissemination and experimental pathology of *Salmonella paratyphi* B infection in white mice Iraq. J. Vet. Med. 24(2) 212 229.
- **12. Al-Jeboori, K. H.** (2007). A study on the bacterial dissemination and experimental pathology of *Salmonella paratyphi* A infection in white mice. The Iraqi J. Med. Sci. 5(3) 23 30.
- **13. Al-Shebibi, S. and Al-Jeboori, K. H. (2001).** A study on bacterial dissemination and experimental intraperitoneal infection of two weeks old chickens with *Salmonella typhimurium*. Iraqi J. Vet. Med. 25(2) 145 157.
- **14. Jones, T. C; Hunt, R, D. and King, N.W. (1997).** Veterinary pathology 6th. Ed. Lippincott Williams and Walkins, Baltimore, Maryland, USA.
- 15. Clark, I. A; Awburn, M.M; Whitten, R. O; Harper, C.G; Liomba, N.G; Molyneux, M. E. and Tylor, T. E. (2003). Tissue distribution of migration inhibitory factor and inducible nitric oxide synthase. in malaria falciparum and sepsis in Africon children. Malarial Journal 2:6.