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DIODE LASER PULPOTOMY ON HUMAN PRIMARY MOLARS A CLINICAL, RADIOLOGICAL AND HISTOLOGICAL STUDY

Thesis submitted to

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In partial fulfillment of the requirements

For the award of the degree of

MASTER OF PHILOSOPHY IN MEDICINE

BY

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Introduction

The incidence of childhood dental caries with widespread geographic distribution has become a worldwide oral health concern and is attributed to the increased intake of carbohydrates and lack of proper oral hygiene practices. The treatment of dental caries in children focuses on preventive intervention with topical application of fluorides and restorations when the lesion is confined to enamel and dentin when untreated due to negligence, ignorance or non-availability of adequate dental care facilities in remote area the carious lesion progresses with the involvement of the pulp dentin complex leading to infection and subsequent extraction resulting premature loss of teeth. This causes an imbalance in the growth of jaw apparatus, arch length discrepancy, masticatory distress, phonation problems, and unpleasant aesthetics. The management of pulpally infected or affected teeth has been tried, to retain the tooth in the arch till its normal exfoliation with different procedures of pulp capping, pulpotomy, pulpectomy, and Lesion Sterilization and Tissue Repair (LSTR).¹

The pulpotomy procedure has been widely practiced with a positive predictive result in comparison to other procedures particularly when the tooth is vital and is defined as "the amputation of vital pulp from the coronal pulp chamber followed by placement of a medicament over the radicular pulp stumps to stimulate repair, fixation or mummification of the remaining vital radicular pulp". The rationale is based on the assumption that inflammation and impaired vascularity caused by the bacterial invasion would be confined to the superficial part of the coronal pulp while maintaining the normal vitality of the radicular pulp^{2,3}. The medicaments and its use for the pulpotomy procedure have been classified into 3 groups (a) paste which makes the radicular pulp tissue completely fixed sterilized and thus obviating infection and internal resorption. **Preservation**-Preservation is fixation of the tissue which is non- inductive and conserves



virtually all of the radicular pulp and the agents used are Glutaraldehyde and Ferric sulfate. **Regeneration**-Regeneration is aimed at keeping the radicular pulp vital, healthy and reparative it completely enclosed within the odontoblast lined dentin chamber while isolating the tissue from noxious restorative material and thereby reducing the chances of internal resorption and induction of reparative dentin formation. The agents are MTA (Mineral Trioxide aggregate), Biodentine, Calcium hydroxide, BMP (Bone Morphogenic Protein), Enriched collagen solution and Collagen calcium phosphate gel, and the recent is by the application of LASER and Electrosurgery. The ideal properties of pulpotomy agents are ought to biocompatible without any adverse biochemical, histological be and immunological effect on the host tissue and on the other hand to have a bactericidal and healing effect on the radicular pulp, to support the regeneration of dentin pulp complex and not interfere with the physiological process of root resorption.^{1,4}

Buckley WK(1904) introduced Formocresol (FC combination of formaldehyde, tricresol, glycerol, and water) as a pulpotomy medicament for primary teeth which has been widely used for decades and considered as "gold standard" due to its bacteriostatic, fixative properties with a success rate of 97%. However, a recent review of the literature and clinical trials suggest formocresol causes potential immunogenic, cytotoxic and mutagenic effects leading to a quest for new biocompatible materials and newer procedures for success of pulpal treatment without any adverse effects^{4,5}.

The term LASER is an acronym for "Light Amplification by Stimulated Emission of Radiation". It was first introduced to medical use in 1959 and has been revolutionized its application for the first time in medical procedures of ophthalmic surgery in 1963. Depending on the application of various tissues, the use of laser



application in dentistry can be categorized as follows: Soft tissue application and hard tissue application^{6,7}. The soft tissue application of laser is wound healing, post herpetic neuralgia, frenectomies, removal of inflamed, hypertrophic tissue, aphthous ulcer, photo-activated dye disinfection, photodynamic therapy for malignancies, aesthetic gingival re-contouring, nerve repair and regeneration, post-surgical pain management, TMJ pain management, and sinusitis⁸⁻¹⁹. The hard tissue applications of laser are photochemical effects, laser fluorescence, cavity preparation, caries and restorative removal etching, and dental hypersensitivity. ²⁰⁻

The most commonly used lasers in dentistry include Holmium Yttrium Aluminium Garnet (Ho: YAG), Neodymium-Yttrium Aluminium Garnet (Nd: YAG), Carbon Dioxide Laser (CO2), Erbium-Yttrium Aluminium Garnet (Er: YAG), Gallium Arsenide (GaAs), Erbium, and Argon lasers. Out of these, diode, Neodymium, Erbium, and CO2 lasers are approved by the Food and Drug Administration (FDA) for use in dentistry. The diode is laser made up of microscopic chips of gallium arsenide or other precious semiconductors emits an infrared light beam that is capable of producing well-localized ablation of soft tissue through conversion of the laser energy to heat that adds the advantage of minimal or no bleeding, faster healing, reduced postoperative infection and minimal use of local anesthetic agent. The bio-stimulative effect of laser produces Adenosine Tri Phosphate (ATP), on mitochondria and create a superficial zone of coagulation with underlying induction and stimulation of dentinogenesis and preservation of pulp vitality. The animal's studies with diode laser pulpotomy revealed the histological changes to be less inflammatory, reduced thermal damage to pulp tissue, accelerated pulpal wound healing, calcific bridge formation and based on clinical and radiological evidence the success of diode laser pulpotomy appears to be promising.²⁴⁻³⁰



Laser in pulpotomy has been clinically tried in pulpally involved human primary teeth with a successful clinical and radiological outcome. However, ideally, all clinical and radiological success outcomes ought to be corroborated by histological study and success. This study is designed to conduct a clinical and radiological evaluation of pulpotomy by diode laser on primary teeth and also includes histological evaluation of LASER pulpotomized teeth which were indicated for serial extraction.

AIMS AND OBJECTIVE

To evaluate the effect of diode laser in pulpotomy of human primary molars in terms of clinical, radiological and histological changes.

- 1. To study the clinical outcome of pain, swelling, mobility, and sinus of diode laser pulpotomy on human primary molars at a different time interval.
- 2. To study the various radiographic changes of furcal radiolucency, internal/ external root resorption and pulp canal obliteration of diode laser pulpotomy of human primary molars at a different time interval.
- 3. To study the histological features of the presence of inflammatory cells infiltrate and the sign of calcific bridge formation of pulp tissue of human primary molar subjected to diode laser pulpotomy.



Material and Methods

The present clinical study entitled "Diode laser pulpotomy on human primary molars: a clinical, radiological and histological study" was carried out in the Department of Pediatric and Preventive dentistry, S.C.B Dental College and Hospital, Cuttack, Odisha during the period from January 2018 to August 2019 with approval from Institutional Ethics Committee. (Annexure I). The participants of the study were selected from children of both genders in the age group of 5-10 years attending the OPD of Pediatric dentistry department having chief complaint of pain in the primary molar teeth. Detailed medical history was obtained and intraoral examination was carried out and the cases which met the inclusion criteria for pulpotomy procedure were selected for the study. The parents/guardian accompanying the participants were described in detail the purpose of this study its methodology and the related risk and benefits, in the local language or another language they can understand. A consent form prepared in English and Odia was distributed to the parents/guardian(Annexure-II) . Only those cases who gave written consent before the start of treatment were enrolled.

INCLUSION CRITERIA

Children in the age group of 5-10 years from both the sexes who had carious and painful primary molars and met the following clinical and radiographic features were included⁵⁵.

(a) Clinical criteria

- i. Carious exposed vital primary molars,
- ii. Absence of tenderness on percussion
- iii. Absence of clinical signs of tooth mobility,
- iv. Absence of sinus/fistula,



- v. Absence of soft tissue swelling/ cellulitis,
- vi. First primary mandibular molars fulfilling the above criteria but indicated for serial extraction for the interceptive orthodontic procedure.

(b) Radiological criteria

- i. Absence of furcal or periapical radiolucency
- ii. Absence of loss of lamina dura
- iii. Absence of widening of the periodontal ligament
- iv. Physiological root resorption less than one third

A total number of 60 primary molars (Maxillary- 6, Mandibular- 54) from both the arches of 60 children of both the genders (Male -37, Female-23) were included in the study. Low-level Laser Pulpotomy (LLLP) was performed in all the selected teeth.

ARMAMENTARIUM

For clinical examination and diagnosis

Gloves

Kidney tray

Mouth mirror (GDC, India)

Explorer (GDC, India)

Tweezers (GDC, India)

For radiographic examination

X-ray machine unit (70 kvP, 7 mA,230V, 50 Hz, New Life Radiology, Italy RVG (ASTRAD IMAGINE)

For clinical procedure



A rubber dam (GDC, India) Air rotor handpiece (NSK, Germany) Benzocaine gel UPS 20% ProGel-B (Septodent) Lignocaine 2% with 1: 80,000adrenaline (2% LIGNOX, Warren, Indoco India) Disposable syringe Spoon excavator (GDC, India) No 2 round bur (Shofu, India) Normal saline (Haseeb Pharmaceuticals Pvt. Ltd, Nagpur, India) Zinc oxide eugenol cement (IRM, Dentsply, USA) Restorative Glass Ionomer cement (type 9, GC Corporation Tokyo, Japan) Glass Ionomer Luting cement (type1, GC Corporation Tokyo, Japan) Mixing pad Stainless steel crown (3M UNITEK, Germany) Sterile cotton Agate spatula Restoration set (GDC India) Vernier caliper Straight handpiece (NSK Germany) **Diode Laser**

iLase (940 nm±10nm ,3.0W Max CW/5.0 W Peak Power(Pulse Mode, BIOLASE ,Inc, USA)

CLINICAL PROCEDURE

A standardized protocol for pulpotomy procedure were carried out in all the selected teeth. Administration of local anesthesia using 2% lignocaine with 1:



80,000adrenaline (2% LIGNOX, Warren, Indoco) isolation with a rubber dam (GDC, INDIA) and access to the pulp chamber was obtained with a no 2 round bur (SHOFU INC.). The coronal pulp was removed using a sharp sterile spoon excavator (GDC) and bleeding was controlled by the application of cotton pallet shocked with normal saline with pressure for 5 minutes. The procedure was repeated until the bleeding was controlled. Then the pulp was ablated to the level of root canal orifice using the 940-nm diode laser (ILase, Biolase) set at 1 W power with a contact pulse mode for 10 sec through a 400-micrometer optical fiber tip. Adequate eye protection was taken by wearing laser protective eyeglasses by the patient, the operator and the assistant during the application of laser. A thin layer of (approx. 2mm) resin reinforced zinc oxide eugenol cement (IRM, Dentsply) base was placed in the pulp chamber followed by type 9GIC(GC gold Label Type 9 GC Corporation, Tokyo, Japan). After one month clinical and radiological evaluation was carried out and a semi-permanent restoration of Stainless Steel crown (3M UNITEK) was cemented in 50 teeth.

The 10 number of pulpotomized mandibular first molars indicated for serial extraction was extracted (5 teeth each on 7th day and 30th day) subjected to histological examination.

Follow up and evaluation.

The clinical and radiological assessment of all teeth treated were carried out at1, 3,6 and 12 months with the following criteria

Clinical Criteria for evaluation

- 1. Presence/absence of pain
- 2. Presence/absence of mobility



- 3. Presence/absence of intraoral swelling
- 4. Presence/absence of sinus

Radiological evaluation criteria

- 1. Presence/absent of radiolucency in the furcal/ Periapical area
- 2. Presence/absence of internal or external root resorption
- 3. Presence /absence of pulp canal obliteration

HISTOLOGICAL STUDY

The extracted teeth were immersed immediately in 10% neutral buffered formalin solution and allowed to remain in it for 24 hours to facilitate fixation. The teeth were then decalcified using 10% formic acid. The specimens were immersed in the solution for five days and the demineralization was checked using a scalpel. The decalcified specimens were washed in running water for removal of the formic acid and send for routine histological tissue processing. The specimens were embedded in molten paraffin wax. Serial sections of 5 µm were cut using soft tissue microtome in a buccolingual plane. The tissue sections were mounted on gelatin-coated slides and subjected to Haematoxylin and Eosin staining (H&E staining). The slides were examined and captured using the Lawrence & Mayo trinocular microscope equipped with a camera (5MP), the image was captured at 40X, 100Xand 400X magnification for histological study and calibrated according to the criteria⁵⁶ described as follows at the department of Oral Pathology and Microbiology, SCB Dental College and Hospital, Cuttack

Inflammatory cell response

Grade 1. Absent or very few inflammatory cells.

Grade 2. A mild or average number of less than 10 inflammatory cells.



Grade 3. Severe inflammatory lesion appearing as an abscess or dense infiltrate involving one-third or more of the coronal pulp.

Grade 4. Completely necrotic pulp.

Dentine bridge formation

Grade 1. Presence of a dentine bridge directly adjacent to some portion of the medicament interface.

Grade 2. Presence of a dentine bridge distant from the medicament interface.

Grade 3. No evidence of any dentine bridge formation in any sections.

The presence or absence of clinical signs of pain, tenderness, swelling, sinus, and mobility was evaluated at 1, 3, 6 and 12 months for the success of the treatment procedure. The presence or absence of radiological sign of periapical or furcal radiolucency, internal or external root resorption and pulp canal obliteration were evaluated for the success of the treatment. The presence or absent of histological sign of Inflammatory cell infiltrate and sign of calcific bridge formation were evaluated for the success of the treatment



Observation

The study sample comprised of 60 children aged between 5-10 years with a mean age of 7.46 ± 1.26 years from both the sexes (Male -37, Female-23). The Diode laser pulpotomy procedure was carried out in all the 60 primary molars from both maxillary and mandibular arches. The Clinical and Radiographic evaluation was carried out in 50 teeth at the interval of 1, 3, 6, and 12 months. 10 number of treated teeth indicated for serial extraction were extracted after the 7th day (N=5) and 30th day (N=5). The extracted teeth were subjected for histological evaluation for inflammatory response and dentinal bridge formation

TABLE-1:

VARIABLE	CATECODY	SA	SAMPLE		
	CATEGORY	No.	%		
AGE	<7	22	36.7		
(In Years)	>7	38	63.3		
GENDER	MALE	37	62		
	FEMALE	23	38		

DISTRIBUTION OF SAMPLE ACCORDING TO AGE AND GENDER

Table-1 depicts the distribution of the sample according to age and gender. According to age 22 (36.7%) were below 7 years of age and 38(63.3%) more than 7 years of age, and according to sex 37(62%) were male and 23(38%) female.



TABLE -2:

DISTRIBUTION OF SAMPLE ACCORDING TO ARCH AND TOOTH TYPE

ARCH	ТООТН	SAMPLE	
		No.	%
MAXILLARY	1 st primary molar	4	6.7
(N=6)	2 nd primary molar	2	3.3
MANDIBULAR	1 st primary molar	28	46.7
(N=54)	2 nd primary molar	26	43.3

Table-2 depicts the distribution of the sample according to arch and tooth type (Primary 1st or 2nd molar). In the maxillary arch, the first primary molar comprises of 4 (6.7%) teeth and the 2nd primary molar 2(3.3%) teeth. In the mandibular arch, 1st primary molar comprises of 28(46.7%) teeth and 2nd primary molar26 (43.3%) teeth.

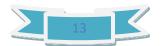


TABLE -3:

ASSESSMENT OF CLINICAL FEATURES AT VARIOUS STUDY INTERVALS

CLINICAL	1M	ONTH	3 MONTH		6 MONTH		12 MONTH	
FEATURE	Present	Absent	Present	Absent	Present	Absent	Present	Absent
	No of							
	tooth (%)							
PAIN	-	50(100)	-	50(100)	2(4)	48(98)	3(6)	47(94)
MOBILITY	-	50(100)	-	50 (100)	-	50 (100)	-	50 (100)
SWELLING	-	50 (100)	-	50 (100)	-	50 (100)	-	50 (100)
SINUS	-	50 (100)	-	50(100)	-	50(100)	-	50(100)

Table-3 depicts the clinical feature of pain, mobility, swelling, and sinus at study intervals of 1, 3, 6 and 12 months. As regards to pain it was absent in 1 and 3 months (100%) and at the end of 6 months 2 (4%) teeth presented with of pain and 48 (98%) without pain and at 12 month 3 (6%) teeth presented with pain and 47(94%) without pain. Mobility, swelling, and sinus were not present in any teeth at 1, 3, 6 and 12 months follow up with 100 % success.

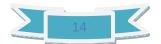


TABLE-4:

ASSESSMENT OF RADIOLOGICAL FEATURES AT VARIOUS STUDY INTERVAL

RADIOLOGICAL FEATURE	1 M	1 MONTH		3 MONTH		6 MONTH		12 MONTH	
	Present	Absent	Present	Absent	Present	Absent	Present	Absent	
	No of	No of	No of	No of					
	tooth (%)	tooth	tooth (%)	tooth					
						(%)		(%)	
FURCAL	-	50(100)	-	50(100)	2(4)	48(96)	3(6)	47(94)	
RADIOLUCENCY									
INTERNAL/EXTERNAL	-	50(100)	-	50 (100)	-	50 (100)	1(2)	49(98)	
ROOT RESORPTION									
PULP CANAL	-	50 (100)	-	50 (100)	-	50 (100)	1(2)	49(98)	
OBLITERATION									

Table-4 depicts the radiographic assessment at various study intervals of 1, 3, 6 and 12 months. As regards to the periapical radiolucency, it was absent in 1 and 3 months (100%) and at the end of 6 months, 2 (4%) teeth presented with furcal radiolucency and 48 (96%) without furcal radiolucency and at 12 months 3 (6%) teeth presented with furcal radiolucency and 47(94%) without furcal radiolucency. As regards to internal/ external root resorption and pulp canal obliteration, it was absent in 1,3and 6 months (100%) and at the end of 12 months, 1 (2%) tooth was presented with internal/external root resorption and 1 (2%) tooth with pulp canal obliteration.



TABLE 5:

CLINICAL AND RADIOLOGICAL ASSESSMENT AT VARIOUS STUDY INTERVAL

	CLINICAL SUCCESS		RADIOLOGICAL SUCCESS		
FOLLOW UP PERIOD	No. of tooth	%	No, of tooth	%	
1 MONTH	50	100	50	100	
3 MONTH	50	100	50	100	
6 MONTH	48	96	48	96	
12 MONTH	47	94	45	90	

Table 5 depicts clinical and radiographic success of the study at 1, 3, 6 and 12 months follow up.

The clinical success was considered when none of the clinical evaluation criteria were present i.e. pain, swelling, mobility and sinus. The clinical success at the interval of 1, 3, 6 and 12 month was 100%, 100%, 96% and 94% respectively.

The radiographic success was considered when none of the radiographic criteria were present i.e. periapical radiolucency, internal /external root resorption and pulp canal obliteration.

The radiological success was 100%, 100%, 96% and 90% in 1, 3, 6 and 12 month follow up respectively.



HISTOLOGICAL EVALUATION

TABLE 6:

INFLAMMATORY CELL RESPONSE SCORE AT THE 7TH DAY AND THE 30THDAY:

DURATION	INFLAMMATORY CELL RESPONSE SCORES						
	GRADE 1	GRADE 2	GRADE 3	GRADE 4	Total		
	No.of tooth	No.of tooth	No.of tooth	No.of tooth			
7 TH DAY	3	2	-	-	5		
30 TH DAY	5	-	-	-	5		

Grade 1. Absent or very few inflammatory cells.

Grade 2. Mild or average number less than 10 inflammatory cells.

Grade 3. Severe inflammatory lesion appearing as an abscess or dense infiltrate involving one-third or more of the coronal pulp.

Grade 4. Completely necrotic pulp.

Table 6 depicts inflammatory cells' response of pulp tissue to diode laser application after 7th day and 30th day. On the 7th day out of 5 pulpotomized teeth 3 teeth showed very few inflammatory cells and 2 teeth showed mild inflammatory cell, no teeth showed severe inflammatory cell or necrotic pulp. At the 30th day, all the teeth showed no inflammatory cells.



TABLE 7

DENTIN BRIDGE FORMATION SCORES ATTHE 7TH DAY ANDTHE 30TH DAY:

DURATION	DENTIN BRIDGE FORMATION SCORES					
	GRADE 1	GRADE 2	GRADE 3	Total		
	No. of	No. of No. of No. of tooth N		No. of		
	tooth	tooth		tooth		
7 DAY	-	-	5	5		
30 DAY	-	-	5	5		

Grade 1. Presence of a dentine bridge directly adjacent to some portion of the medicament interface.

Grade 2. Presence of a dentine bridge distant from the medicament interface.

Grade 3. No evidence of any dentine bridge formation in any sections.

Table-7depicts dentine bridge formation in response to diode laser application after the 7th day and the 30th day. After 7thday and 30th day no teeth showed any sign of formation of dentin bridge.



Discussion

Modern pediatric endodontic procedure and technique for pulpal treatment of primary teeth has always remained a challenge. The use of formocresol, glutaraldehyde, and other medicaments has been successfully tried with variable results and concern regarding their biocompatibility. A search for alternative methods has already been advocated including the application of LASER (Light Amplification by Stimulated Emission of Radiation), as the LASER beam does not come into contact with the tissue in this process, the cutting process does not mechanically affects or damage the irradiated tissue and has the potential to make the area aseptic⁴⁹.

A relatively newer method that has emerged is the use of DIODE Laser a Low-level Laser Therapy (LLLT) energy has been advocated for use in pulpal treatment (Pulpotomy) of primary molars and reported to be able to overcome the histologic deficits thereby accelerating the wound healing of the pulp and the expression of the lectins and collagens and also enhances the formation of calcified nodules in human dental pulp fibroblasts, alkaline phosphatase activity, and the production of collagen and osteocalcin⁵⁷. Evidence suggests it also accelerates the reparative dentin bridge formations, accelerated wound healing, regeneration, relief of pain, and enhancement of local immunity. It also promotes cell proliferation, the formation of granulation tissue and accelerates collagen synthesis by fostering the formation of type I and type III procollagen specific pools of mRNA, increases adenosine triphosphate (ATP) synthesis within the mitochondria and activates the lymphocytes⁵⁸⁻⁶⁰. The LLLT can stimulate the odontoblastic cells by activating the cell signalling molecules, which helps in modulating the activity of the transforming growth factor-beta ligands, ultimately leading to mineralization and formation of tertiary dentin due to the presence of Smads, a class of proteins^{46,61}.



In the present study iLase (940 nm \pm 10nm, 3.0W Max CW/5.0 W Peak Power (Pulse Mode, BIOLASE, Inc, USA) diode laser has been selected since it is proved to be efficient by the researcher for its application in pulp therapy of primary dentition without any adverse effect^{26,35,62,63}.

The clinical parameters assessed in the present study included pain, mobility swelling, and sinus at different time interval of 1, 3, 6 and 12 months and the presence of pain was seen in only 2 teeth in 6 months and 3 teeth in 12 months and no teeth showed any sign of mobility, swelling and sinus in all the follow-up time interval with a clinical success rate of 98 % and 94% at 6 and 12 months respectively. The radiological parameter assessed in the present study included furcal radiolucency, internal/external root resorption and pulp canal obliteration at different time intervals of 1, 3,6and 12 months. There was a presence of furcal radiolucency only in 2 teeth and 3 teeth at the end of 6 and 12 months with a success rate of 96% and 94% respectively at 6 and 12 months. Regarding internal/external root resorption and pulp canal obliteration, it was present only in one tooth in each with overall success of 94 % at the end of 12 month follow up and in conformity with the earlier report published indicating the fact that the use of diode laser in pulpotomy procedure of primary teeth and considered to be safe and successful^{49,51,63}.

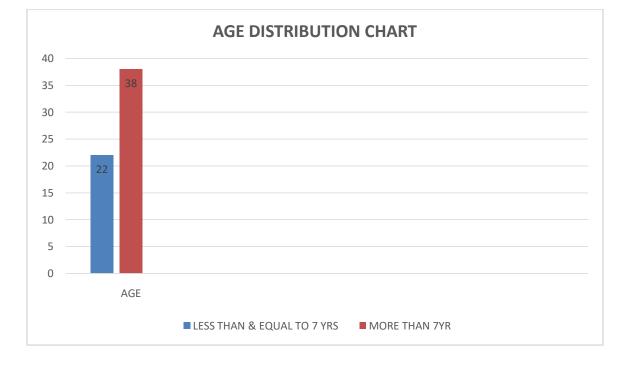
The comparison of different medicaments for pulpotomy of primary molars when compared with LLLT at different time interval as indicated in the following facts, formocresol pulpotomy showed a clinical and radiological success rates ranging between 70 to 97%^{64,65} and when compared with LASER the clinical success and radiological success was 100% and 90% respectively³¹ in spite of clinical success rate of formocresol the mutagenic and toxic effects have been reported cytotoxic limiting its use in pulpotomy⁶⁶⁻⁶⁹. The clinical and radiological



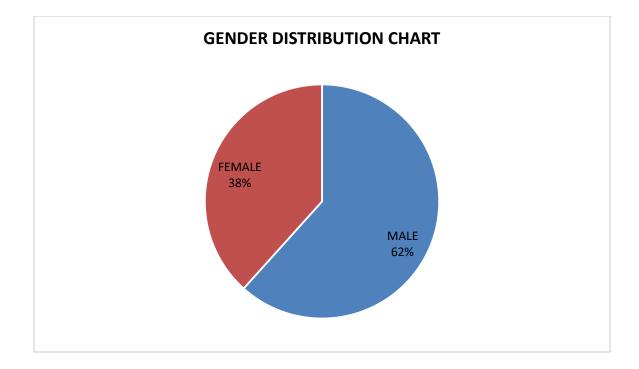
success of Ferric sulfate pulpotomy from the different studies ranges from 78% - 100% and 43% - 97%⁷⁰⁻⁷³ when compared with laser clinical success of 93% and radiological success of 78% was observed⁷⁴. Electrosurgical pulpotomy showed the clinical success ranged from 87.5% - 100% while radiographic success rate ranged from 84% - 96.8%⁷⁵⁻⁷⁶when compared with laser it showed 100% clinical and 80% radiological success³. Many research studies on laser pulpotomy show a clinical success of ranged 85.71% – 100% and radiological success of ranged 67% - 94.1%.)^{49,51,63}. MTA pulpotomy show with a high success rate of 80% to 100%⁷⁷⁻⁷⁹ On the other hand, calcium hydroxide pulpotomy showed significantly worse result and the failure rate ranged from 20- 47% ⁸⁰⁻⁸⁴.

Histological studies of pulpotomized animals teeth with diode laser revealed of least amount of inflammatory cell infiltrate and maximal healing after 7 days follow up^{37,39} and pulpotomy with Er: YAG laser showed no inflammation or resorption in any cases after one week follow up but after 60 days follow up a layer of odontoblastic cell formed³⁵. In a studied for 30th day and 45th-day post pulpotomy, histological changes showed Laser pulpotomized teeth with lesser inflammatory cell infiltration and more macrophage infiltration, and as regards to calcific bridge formation LASER found to be able to initiate dentin bridge at the interface between the pulp and the material^{85,86}. Nd: YAG LASER Pulpotomy on human primary teeth showed after 7th and 60th day follow up less inflammatory cell but no dentine bridge formation⁴⁷.Hard tissue barrier formation and continuous odontoblastic layer was observed in a study when pulpotomy was carried out with LASER with calcium hydroxide sealing suggesting successful response with the support that LLLT reduced trauma and inflammation in the pulp tissue, helping the healing process⁸⁷.

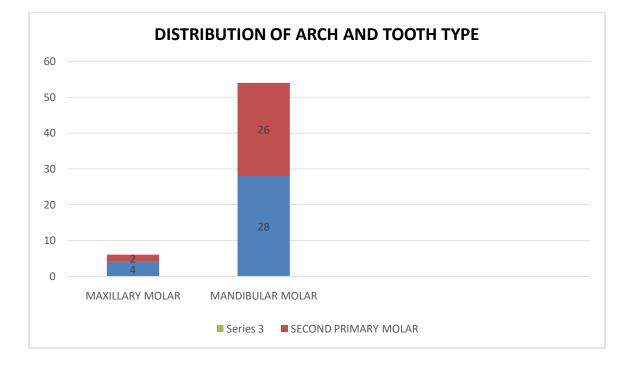


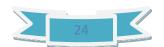


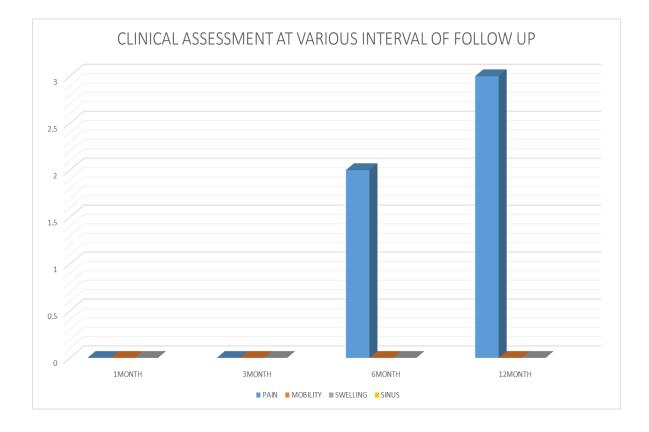




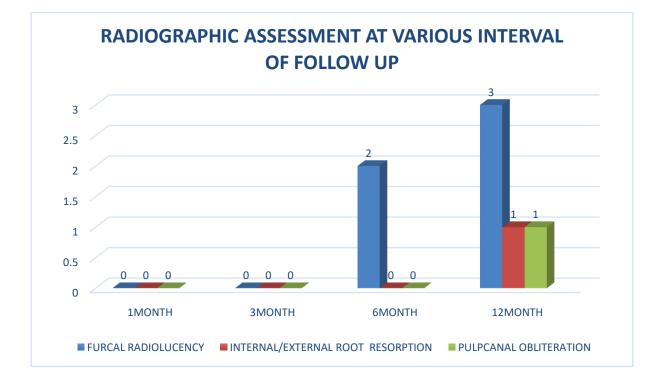


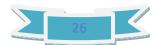


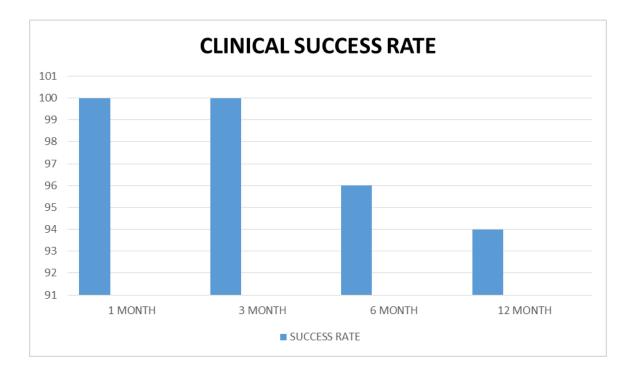




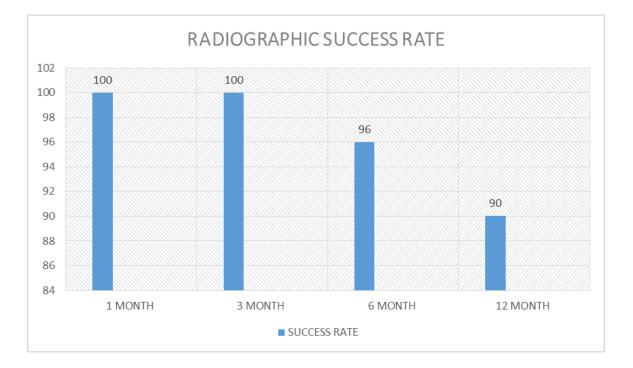








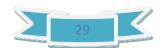








ARMAMENTARIUM





DIODE LASER









LASER GLASS PROTECTION





PRE OPERATIVE XRAY SHOWING CARIOUS 74



ACCESS OPENING DONE



RUBBER DAM PLACED



REMOVAL OF CORONAL PULP

CLINICAL PROCEDURE Contd..



MOIST COTTON PLACED



LASER EXPOSED



POST LASER EXPOSURE



IRM RESTORATION DONE



GIC RESTORATION DONE

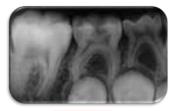


STAINLESS STEEL CROWN PLACED



CLINICAL PROCEDURE contd..

RADIOLOGICAL EVALUTION



PRE OP



1 MONTH



3 MONTH



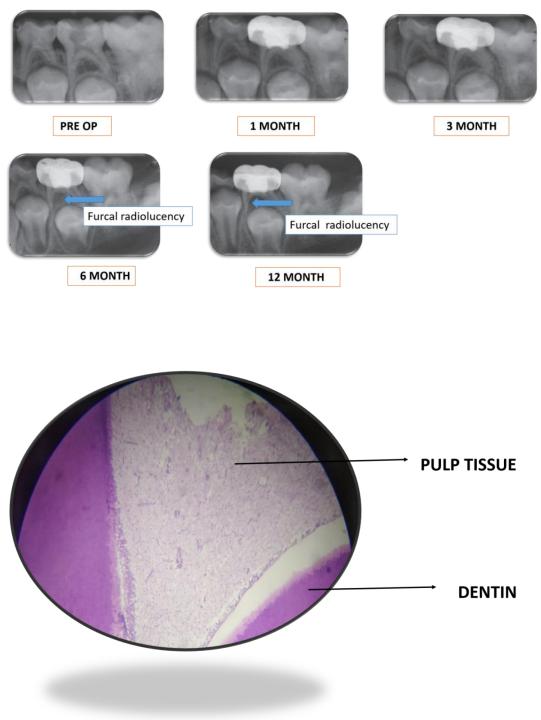
6 MONTH



12 MONTH



RADIOLOGICAL EVALUTION



Pulp tissue showing no inflammatory cells





Pulp tissue showing few inflammatory cells



Conclusion

Following conclusion can be drawn from the result of the study

- 1. The overall clinical success rate of diode laser pulpotomy was 94% at the end of 12 months follow up with absent of swelling, sinus, and mobility
- 2. The overall radiographic success rate of diode laser pulpotomy was 90% at the end of 12 months follow up with absent of sign of furcal radiolucency, internal/external root resorption and pulp canal obliteration
- 3. The histological evaluation showed no inflammatory cells respond in diode laser pulpotomy at 7th day and 30th day follow up indicating proper healing of the pulp tissue
- 4. Calcific bridge formation was not seen due to shorter study duration
- 5. The diode laser is very effective in pediatric dentistry and often felt comfortable and acceptable by the children and their families with a magical effect.



Bibliography

- 1. Fuks AB, E Edelman -Europepmc.org Pulp therapy in the primary dentition. Current opinion in dentistry, 1991.
- 2. Textbook of Pediatric Dentistry: Second Edition, Ramond L Braham & Merle E Morris.
- Yadav P, Indushekar Kr, Saraf Bg, Sheoran N, Sardana D.(2014).Comparative evaluation of Ferric sulfate,Electrosurgical and Diode Laser on human primary molar pulpotomy: an "in vivo" study. www.jstage.jst.go.jp/browse/islsm.
- Ranly DM. Pulpotomy therapy in primary teeth: new modalities for old rationales. Pediatric Dentistry: November/Decembe1r9 94- Volume 16, Number6
- Sweet CA. Procedure for treatment of exposed and pulpless deciduous teeth. J Am Dent Association 1930; 17:1150-1153.
- 6. Einstein A. Zur Quantentheorie der Strahlung. Physiol Z. 1917; 18:121–8.
- Gross AJ, Hermann TR. History of lasers. World J Urol. 2007; 25:217–20. [PubMed: 17564717]
- Kitsmaniuk ZD, Demochko VB, Popovich VI. The use of low energy lasers for preventing and treating postoperative and radiation-induced complications in patients with head and neck tumors. Vopr Onkol. 1992; 38:980–6. [PubMed: 1300810]
- Iijima K, Shimoyama N, Shimoyama M, Yamamoto T, Shimizu T, Mizuguchi T. Effect of repeated irradiation of low-power He- Ne laser in pain relief from postherpetic neuralgia. Clin J Pain. 1989; 5:271–4. [PubMed: 2562460]
- Olivi G, Genovese MD, Caprioglio C. Evidence-based dentistry on laser pediatric dentistry. Eur J Paediatr Dent. 2009; 10:29–40. [PubMed: 19364243]
- 11. Yeh S, Jain K, Andreana S. Using a diode laser to uncover dental implants in secondstage surgery. Gen Dent. 2005; 53:414–7. [PubMed: 16366049]
- Hargate G. A randomized double-blind study comparing the effect of 1072-nm light against placebo for the treatment of herpes labialis. Clin Exp Dermatol. 2006; 31:638–41.
 [PubMed: 16780494]
- Dobson J, Wilson M. Sensitization of oral bacteria in biofilms to killing by light from a low-power laser. Arch Oral Biol. 1992; 37:883–7. [PubMed: 1334649]
- Dougherty TJ. An update on photodynamic therapy applications. J Clin Laser Med Surg. 2002; 20:3–7.[PubMed: 11902352.



- Sarver DM, Yanosky M. Principles of cosmetic dentistry in orthodontics: Part 2. Soft tissue laser technology and cosmetic gingival contouring. Am J Orthod Dentofac Orthop. 2005; 127:85–90. [PubMed:15643420].
- Mester AF, Snow JB, Shaman P. Photochemical effects of laser irradiation on the neuritic outgrowth of olfactory neuroepithelial explants. Otolaryngol Head Neck Surg. 1991; 105:449–56. [PubMed: 1945434]
- Kawakami T, Ibaraki Y, Haraguchi K, Odachi H, Kawamura H, Kubota M, et al. The effectiveness of GaAlAs semiconductor laser treatment to decrease pain after irradiation. Higashi Nippon Shigaku Zasshi. 1989; 8:57–62. [PubMed: 2519920]
- Kruchinina I, Feniksova LV, Rybalkin SV, Pekli FF. The therapeutic effect of heliumneon laser on the microcirculation of nasal mucosa in children with acute and chronic maxillary sinusitis as measured by conjunctival biomicroscopy. Vestn Otorinolaringol. 1991; 3:26–30. [PubMed: 1862594]
- Fleming MG, Maillet WA. Photopolymerization of composite resin using the argon laser. J Can Dent Assoc. 1999; 65:447–50. [PubMed: 10518340]
- With PJ, Nord A. Caries experience in orthodontically treated individuals. Angle Orthod. 1977; 47:59–64. [PubMed: 264779]
- Burkes EJ, Hoke J, Gomes E, Wolbarsht M. Wet versus dry enamel ablation by Er: YAG laser. J Prosthet Dent. 1992; 67:847–51. [PubMed: 1403876]
- 22. Dostalova T, Jelinkova H, Kucerova H, Krejsa O, Hamal K, and Kubelka J, et al. Noncontact Er: YAG laser ablation: Clinical evaluation. J Clin Laser Med Surg. 1998; 16:273–82. [PubMed: 9893509]
- 23. Niranjani et al (2015).Clinical Evaluation of Success of Primary Teeth Pulpotomy Using Mineral Trioxide Aggregate[®], Laser and Biodentine[™]- an In Vivo Study, Journal of Clinical and Diagnostic Research. 2015 Apr, Vol-9(4): ZC35-ZC37
- 24. Yodaiken RE. The uncertain consequences of formaldehyde toxicity. Journal of the American Dental Association 1981;246: 1677–1678
- 25. Gontijo I, Navarro RS, Haypek P, Ciamponi AL, Haddad AE. The applications of diode and Er: YAG lasers in labial frenectomy in infant patients. J Dent Child.2005;72:10–5.
- 26. Saltzman B, Sigal M, Clokie C, Rukavina J, Titley K & Kulkarni VG: Assessment of a novel alternative to conventional formocresol-zinc oxide eugenol pulpotomy for the



treatment of pulpally involved human primary teeth: diode laser-mineral trioxide aggregate pulpotomy. International Journal of Paediatric Dentistry 2005 15: 437–447.

- Verma S, Maheswari S, Singh R, Choudhary P. (2012) Laser in Dentistry: An innovative tool in modern dental practice. National Journal of maxillofacial surgery2012 Juldec3(2):124-132.
- 28. Maturo P, Perugia C, Docimo R (2013). The versatility of an 810 nm diode laser in pediatric dentistry. International Journal Of clinical dentistry vol. 6, no. 2.
- 29. McNally KM, Gillings BR, Dawes JM. Dye-assisted diode laser ablation of carious enamel and dentine. Australian Dental Journal 1999; 44: 169–175
- Jeffrey IWM, Lawson B, Saunders EM. Dentinal temperature transients caused by exposure to CO2 laser irradiation and possible pulpal damage. Journal of Dentistry 1990; 18: 31–36.
- 31. Ansari G, Morovati SP, Asgary S. Evaluation of four pulpotomy techniques in primary molars: A randomized controlled trial. Iran Endod J. 2018;13(1):7-12.
- 32. Pratima B, Chandan GD, Nidhi T, Nitish I, Sankriti M, Nagaveni S. Postoperative assessment of diode laser zinc oxide eugenol and mineral trioxide aggregate pulpotomy procedures in children: A comparative clinical study. J Indian SocPedodPrev Dent 2018;36:308-14.
- 33. Kuo YH, Jr Rung L, Huang WH, Chiang ML. Clinical outcomes for primary molars treated by different types of pulpotomy: A retrospective cohort study. Journal of the Formosan Medical Association (2018) 117, 24-33.
- Junquera MA et al Clinical, Radiographic and Histological Evaluation of Primary Teeth Pulpotomy Using MTA and Ferric Sulfate. Brazilian Dental Journal (2018) 29(2): 159-165.
- 35. Konyala HR, Mareddy AR, Puppala N, Reddy NV, Mallela MK, Susheela KP. Clinical, Radiological, and Histological Assessment of Magnetic Nanoparticles as Pulpotomy Medicament in Primary Molars. Int J Clin Pediatr Dent 2018;11(4):283-287.
- 36. Joshi P.A comparative evaluation between formocresol and diode laser-assisted pulpotomy in primary molars– an in vivo study. ejpmr, 2017,4(5), 569-575.
- 37. Prabhakar AR, Yavagal C, Shagale AM. Evaluation of a Novel Pulpotomy Technique with a Low-level Laser Therapy and Its Comparison with Mineral Trioxide Aggregate and Pulpotec: An Animal Trial. Int J Laser Dent 2015;5(3):58-62.



- Uloopi K, Vinay C, Ratnaditya A, Gopal AS, Mrudula K, Rao RC. Clinical evaluation of low-level diode laser application for primary teeth pulpotomy. J ClinDiagnRes : JCDR. 2016;10(1): Zc67-70.
- 39. Prabhakar AR, Shagale AM, Yavagal C. Histological Evaluation of a Novel Pulpotomy Technique with Low-level Laser Therapy and Its Comparison with Formocresol: A Randomized Animal Trial. Int J Laser Dent 2016;6(1):1-5.
- 40. Marques NCT, Neto NL, de Oliveira Rodini C, Fernandes AP, Sakai VT, Machado MA, et al. Low-level laser therapy as an alternative for pulpotomy in human primary teeth. Lasers Med Sci 2015;30:1815-22.
- 41. Gupta G, Rana V, Srivastava N, Chandna P. Laser Pulpotomy—An Effective Alternative to Conventional Techniques: A 12 Months Clinicoradiographic Study. Int J Clin Pediatr Dent 2015;8(1):18-21.
- 42. Neto LN, Moretti SA, Sakai TV, Machado MM. Clinical and radiographic outcomes of the use of capping materials in vital pulp therapy of human primary teeth. Brazilian Dental Sciences 2015; 18(1): 75-80.
- 43. Fernandes AP, Lourenco Neto N, Teixeira Marques NC, SilveiraMoretti AB, Sakai VT, Cruvinel Silva T, et al. Clinical and radiographic outcomes of the use of low-level laser therapy in the vital pulp of primary teeth. Int J Paediatr Dent 2015;25: 144-50.
- 44. El-Meligy O, Abdalla M, El-Baraway S, El-Tekya M, Dean JA. Histological evaluation of electrosurgery and formocresol pulpotomy techniques in primary teeth in dogs. J Clin Pediatr Dent 2001 Fall;26(1):81-85.
- 45. Cannon M, Wagner C, Thobaben JZ, Jurado R, Solt D. Early Response of Mechanically Exposed Dental Pulps of Swine to Antibacterial-Hemostatic Agents or Diode Laser Irradiation. J Clin Pediatr Dent 35(3): 271–276, 2011.
- 46. Sivadas S, Rao A, Natarajan S, Shenoy R, Srikrishna SB. Pulpal response to ferric sulfate and diode laser when used as pulpotomy agent: An in vivo study. J Clin Diagn Res: JCDR. 2017;11(6):87-91.
- 47. Golpayegani MV, Ansari G, Tadayon N, Shams S, Mir M (2009). Low-Level Laser Therapy for Pulpotomy Treatment of Primary Molars. J Dent Tehran Univ Med Sci, 6:168-74.



- 48. De Souza EB, Cai S, Simionato MR, Lage-Marques JL. High-power diode laser in the disinfection in depth of the root canal dentin. Oral Surg Oral Med Oral Pathol Oral RadiolEndod 2008;106:68-72.
- 49. Odabas ME, Bodur H, Baris E, Demir C (2007).Clinical, Radiographic, and Histopathologic Evaluation of Nd: YAG Laser Pulpotomy on Human Primary Teeth. J Endod, 33:415–21.
- 50. Sheila C et al. Clinical and radiographic evaluation of diode laser (810nm) Pulpotomy and formocresol pulpotomy-an invivo study. Annals and Essences of Dentistry 10.5368/aedj.2017.9.3.1.1.
- Liu JF (2006). Effects of Nd: YAG laser pulpotomy on human primary molars. J Endod, 32:404-7.
- 52. Huth KC, Paschos E, Hajek-Al-Khatar N, Hollweck R, Crispin A, Hickel R, Folwaczny M (2005). Effectiveness of 4 Pulpotomy Techniques— Randomized Controlled Trial. J Dent Res, 84:1144-48.
- 53. Durmus B, Tanboga I. In vivo evaluation of the treatment outcome of pulpotomy in primary molars using a diode laser, formocresol, and ferric sulphate. Photo med Laser Surg. 2014;32(5):289-95.
- 54. Dean JA, Mack RB, Fulkerson BT, Sanders BJ (2002). Comparison of electrosurgical and formocresol pulpotomy procedures in children. Int J Pediatr Dent, 12:177-82.
- 55. Guideline on pulp therapy for primary and immature permanent teeth. Reference Manual 2014;37(6):244-252.
- 56. Cox CF, Su[¨] bay RK, Suzuki S, Suzuki SH, Ostro E. Biocompatibility of various dental materials: pulp healing with a surface seal. Int. J Periodontics Restorative Dent 1996; 16:241–251.
- 57. Sun G, Tuner J. Low-level laser therapy in dentistry. DentClin North Am 2004 Oct;48(4):1061-1076.
- 58. Toomarian L, Fekrazad R, Sharifi D, Baghaei M, Rahimi H, Eslami B. Histopathological evaluation of pulpotomy with Er, Cr: YSGG laser vs formocresol. Lasers Med Sci. 2008;23(4):443-50.
- 59. Jukic S, Anic I, Koba K, Najzar-Fleger D, Matsumoto K. The effect of pulpotomy usingCO2 and Nd: YAG lasers on dental pulp tissue. IntEndod J 1997;30:175–80.



- 60. Todea C, Kerezsi C, Balabuc C, Calniceanu M, Filip L. Pulpcapping conventional to laser-assisted therapy (I). J OralLaserAppl 2008; 8(2):71-82.
- 61. Mareddy A, Mallikarjun SB, Shetty PV, Vanga V, Rao N, Chandru TP. Histological evaluation of diode laser pulpotomy in dogs. J Oral Laser Appl 2010;10:7-16.
- 62. Alghamdi KM, Kumar A, Moussa NA. Low-level laser therapy: a useful technique for enhancing the proliferation of various cultured cells. Lasers Med Sci. 2012;27(1):237-49.
- 63. Kreisler M, Christoffers AB, Willershausen B, Hoedt B. Effect of low-level GaAlAs laser irradiation on the proliferation rate of human periodontal ligament fibroblasts: an in vitro study. J Clin Periodontol. 2003;30(4):353-58.
- 64. Hicks MJ, Barr ES, Flatiz CM. Formocresol pulpotomies in primary molar: a radiographic study in pediatric dentistry practice. J Pedod 1986;10:331–39.
- 65. Fuks AB, Bimstein E. Clinical evaluation of diluted formocresol pulpotomies in the primary tooth of school children. Pediatr Dent 1981;3:321–24.
- 66. Myers DR, Pashley D, Whitford G, Mckinney RV. Tissue changes induced by the absorption of formocresol from pulpotomy sites in dogs. Pediatr Dent 1983;5:6–8.
- 67. Lewis BB, Chester SB. Formaldehyde in dentistry: a review of mutagenic and carcinogenic potential. J Am Dent Assoc 1981;103:429 –34.
- 68. Fei AL, Udin RD, Johnson R (1991). A clinical study of ferric sulfate as a pulpotomy agent in primary teeth. Pediatr Dent, 13:327-32.
- 69. Casas MJ, Layug MA, Kenny DJ, Johnston DH, Judd PL (2003). Two-year outcomes of primary molar ferric sulfate pulpotomy and root canal therapy. Pediatr Dent, 25:97-102.
- 70. Smith NL, Seale NS, Nunn ME (2000). Ferric sulfate pulpotomy in primary molars: a retrospective study. Pediatr Dent, 22:192-9.
- Ibricevic H, Al-Jame Q (2003). Ferric sulfate and formocresol in pulpotomy of primary molars: a long term follow-up study. Eur J Paediatr Dent, 4:28-32.
- 72. Casas JM, Kenny DJ, Johnston DH, Judd PL (2004). Long-term outcomes of primary molar ferric sulfate pulpotomy and root canal therapy. Pediatr Dent, 26:44-48.
- Papagiannoulis L (2002). Clinical studies on ferric sulfate as a pulpotomy medicament in primary teeth. Eur J Paediatr Dent, 3:126-32.
- 74. Neamatollahi H, Tajik A (2006). Comparison of clinical and radiographic success rates of pulpotomy in primary molars using Formocresol, Ferric Sulfate and Mineral Trioxide Aggregate (MTA). J Dent, 3:7-14.



- 75. Mack RB, Dean JA (1993). Electrosurgical pulpotomy: A retrospective human study. ASDC J Dentist Child, 60:107-14.
- 76. Bahrololoomi Z, Moeintaghavi A, Emtiazi M, Hosseini G (2008). Clinical and radiographic comparison of primary molars after formocresol and electrosurgical pulpotomy: a randomized clinical trial. Indian J Dent Res, 19:219-23.
- 77. FarrokhGisoure E (2011). Comparison of three pulpotomy agents in primary molars: a randomized clinical trial. Iran Endod J, 6:11-14.
- 78. Torabinejad M, Hong CU, McDonald F, Pitt Ford TR. Physical and chemical properties of new root-end filling material. J Endod 1995;21:349-53.
- 79. Maroto M, Barberia E, Planells P, Garcia Godoy F. Dentin bridge formation after mineral trioxide aggregate (MTA) pulpotomies in primary teeth. Am J Dent 2005;18:151-54.
- 80. Pitt Ford TA, Dorn SO, Kariyawasam SP. Effect of various zinc oxide materials as rootend fillings on healing after replantation. IntEndod J 1995;28:273-78.
- 81. FarrokhGisoure E (2011). Comparison of three pulpotomy agents in primary molars: a randomized clinical trial. Iran Endod J, 6:11-14.
- 82. Schröder U (1978). A 2-year follow-up of primary molars, pulpotomized with a gentle technique and capped with calcium hydroxide. Scand J Dent Res 86 273-278.
- 83. Gruythuysen RJ, Weerheijm KL (1997). Calcium hydroxide pulpotomy with a lightcured cavity-sealing material after two years. J Dent Child 64:251-253.
- 84. Waterhouse PJ, Nunn JH, Whitworth JM (2000). An investigation of the relative efficacy of Buckley's Formocresol and calcium hydroxide in primary molar vital pulp therapy. Br Dent J 188:32-36.
- 85. Alghamdi KM, Kumar A, Moussa NA. Low-level laser therapy: a useful technique for enhancing the proliferation of various cultured cells. Lasers Med Sci. 2012;27(1):237-49.
- 86. Jukic S, Anic I, Koba K, Najzar-Fleger D, Matsumoto K. The effect of pulpotomy usingCO2 and Nd: YAG lasers on dental pulp tissue. IntEndod J 1997;30:175–80.
- 87. Oliveira T M, Moretti A BS, Sakai V T, LourençoNeto N, Machado M A A M and Abdo R C C 2013 Clinical, radiographic and histologic analysis of the effects of pulp capping materials used in pulpotomies of human primary teeth Eur. Arch. Paediatr. Dent. 14 65– 71



- 88. Caicedo R, Abbott PV, Alongi DJ, Alarcon MY. Clinical, radiographic and histological analysis of the effects of mineral trioxide aggregate used in direct pulp capping and pulpotomies of primary teeth. Australian Dental Journal 2006;51:(4):297-305.
- 89. Luomanen M, LehtoVp, MeurmanJh (1988) Myofibroblasts in healing laser wounds of rat tongue mucosa. Archives of Oral Biology 28, 287–91.
- 90. Matsui S, Takeuchi H, Tsujimoto Y, Matsushima K. Effects of Smads and BMPs induced by Ga-Al-As Laser irradiation on calcification ability of human dental pulp cells. J Oral Sci 2008;50:75-81.
- 91. Tatey Y, Yoshida K, Yoshibha N, Iwaku M, Okiji T, Orshima H. Odonotoblast responses to GaAlAs laser irradiation in rat molars: An experimental study using heat shock protein-25 Immunohistochemistry. Eur J Oral Sci 2006:114;50-57.
- 92. Jukic S, Anic I, Koba K, Nayzar-Fleger D, Matsumoto K. The effect of pulpotomy using carbon dioxide and Nd: YAG lasers on dental pulp tissue. Int Endodontic J 1997;30:175-180.
- 93. Olivi G, Genovese MD, Caprioglio C (2009) Evidence-based dentistry on laser pediatric dentistry: review and outlook. Eur J Paediatr Dent 10:29–40.
- 94. Martens LC (2011) Laser physics and a review of laser applications in dentistry for children. Eur Arch Paediatr Dent 12:61–67.
- 95. Cannon M, Wagner C, Thobaben JZ, Jurado R, Solt D (2011) Early response of mechanically exposed dental pulps of swine to antibacterial-hemostatic agents or diode laser irradiation. J Clin Pediatr Dent 35:271–276.
- 96. Basso FG, Pansani TN, Turrioni AP, Bagnato VS, Hebling J, de Souza Costa CA (2012) In vitro wound healing improvement by low-level laser therapy application in cultured gingival fibroblasts. Int J Dent 2012:719452. DOI:10.1155/2012/719452.

