

REVIEW ARTICLE

A REVIEW OF THE INNOVATIVE DRYING TECHNOLOGIES FOR BIOPHARMACEUTICALS

Pritha Singha Roy¹, Gaurab Joarder², Saibal Debnath³ and Avisek Pahari⁴

- 1. Assistant Professor, Department of Pharmaceutics, Global College of Pharmaceutical Technology, Krishnanagar, Nadia-741102, West Bengal.
- 2. Undergraduate Student, Global College of Pharmaceutical Technology, Krishnanagar, Nadia-741102, West Bengal.
- 3. Undergraduate Student, Global College of Pharmaceutical Technology, Krishnanagar, Nadia-741102, West Bengal.
- 4. Undergraduate Student, Global College of Pharmaceutical Technology, Krishnanagar, Nadia-741102, West Bengal.

..... Manuscript Info

.....

Abstract

Manuscript History Received: 31 March 2022 Final Accepted: 30 April 2022 Published: May 2022

Key words: -

Process Analytical Technology (PAT), PRINT® Technology, Lynfinity® Technology, Drying Techniques

Reviewing data from the previous twenty to twenty-five years reveals that bio-pharmaceuticals are a sudden, dramatic, and incredibly significant finding in progressively enhancing the quality of life for patients with different kinds of malignancies, auto-immune illnesses, genetic disorders, etc. Drying technologies are a required manufacturing step in the pharmaceutical industry/production unit, and an understanding of drying technologies and how to use them is now an absolute must. With the increased demand for biopharmaceuticals, it is essential to reduce production costs without sacrificing product safety, quality, or effectiveness. The predominant commercial method for generating solid biopharmaceuticals is batch freeze-drying. However, freeze-drying is expensive compared to other procedures and is not ideal for lengthy working hours, in addition to requiring a large initial capital investment and significant energy consumption, resulting in high total expenses. This article discusses innovative drying methods for parenteral biopharmaceuticals. Spin-freeze drying, spray drying, and Lynfinity® Technology enable continuous manufacturing, while PRINT® Technology and MicroglassificationTM manage dry particle characteristics. As a consequence, certain drying processes may need less validation. Process Analytical Technology (PAT) and offline dramatisation may give extra information on CPPs and CQAs during biopharmaceutical manufacture. These processing approaches might boost biopharmaceutical product production while reducing expenses.

.....

Copy Right, IJAR, 2022, All rights reserved.

Introduction:-

It is possible to make complex treatments known as biopharmaceuticals by employing living cells or organisms in combination with cutting-edge engineering. These include complex biomolecules like antibodies, antibody-drug conjugates (ADCs), reassembled proteins (Nanobodies), enzymes, hormones (hormones), vaccinations (gene therapy), etc. Temperature, moisture, lengthy storage, denaturants, organic solvents, shear, and oxygen all have a

Corresponding Author:- Pritha Singha Roy

Address:- Assistant Professor, Department of Pharmaceutics, Global College of Pharmaceutical Technology, Krishnanagar, Nadia-741102, West Bengal.

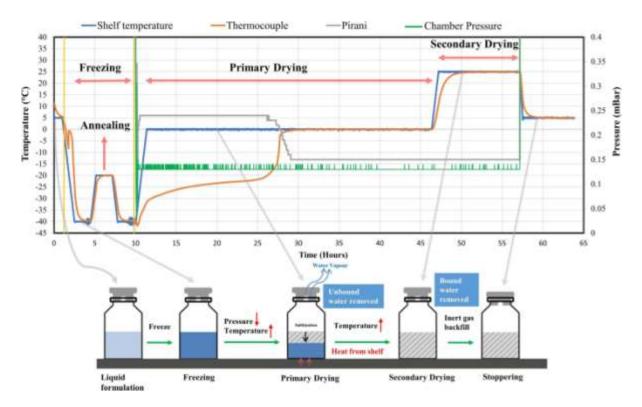
deleterious effect on the resilience of protein-based biopharmaceuticals. In biopharmaceutical products, protein aggregation is one of the most difficult circumstances to control, and it may occur at any stage of the production process or even during administration. [Declerck, P.J., 2012] A number of fill-finish techniques are used to synthesise parenteral biopharmaceuticals, which may be either liquid or stable. Freeze-dried biopharmaceuticals are widely available in European markets. When it comes to producing highly powerful biopharmaceuticals, freezedrying is the gold standard. [Gervasi, V.et al., 2018] Freeze drying has a number of benefits that have been discovered. Heat-sensitive and unstable biopharmaceuticals may be preserved for longer periods of time by freezedrying, which also cuts down on waste. After processing, the low residual moisture content (RMC) and moderate temperatures minimise chemical breakdown, maintaining the product in a solid condition. Traditional freeze-drying, on the other hand, has a number of drawbacks, including long processing times, expensive capital costs, and reduced system efficiency due to excessive electricity demand. [Martagan, T.et al., 2020] A static method will also increase the likelihood of a method failure, which will cause the whole batch to be rejected [Tang, X., Pikal, M.J., 2004]. Because a batch system can only handle small amounts of unit doses, a large number of vials is required. No matter how meticulously a product is manufactured, human participation throughout the process may raise the danger of contamination (cGMP). Thirdly, it was discovered that the dryer's design, field closure, load condition, and product composition all affect heat and mass transfer and scaling up [Rambhatla, S., Pikal, M.J., 2003]. Due to radiation from the cabinet walls, vials at the cabinet's perimeter dry faster than vials in the cabinet's centre, resulting in a problem of vial heterogeneity. Free-flowing powders produced by various drying processes may be reconstituted much faster than effective high-interest freeze-dried protein cakes. No longer well known in the biopharmaceutical business, non-stop drying technology provides numerous characteristics that might be studied to overcome the problems associated with traditional freeze-drying. Readers have referred to the aforementioned points of view for particular numbers on alternative non-stop freeze-drying processes [Pisano, R., 2019]. Several new drying methods are examined in this study, including spin-freeze drying, freeze drying of suspended vials without stopping, active freeze drying (spray freezing), dynamic freeze drying (dynamic freeze drying), Lynfinity® drying (spray drying), PRINT drving (printing) and MicroglassificationTM drving for biopharmaceuticals. Product COAs and procedure CPPs may be evaluated in detail using biopharmaceutical characterization approaches and PAT in combination with drying techniques [Al-Hussein, A., Gieseler, H., 2012]. This ensures that goods are safe and effective before they are provided to patients. Procedure components are crucial in ensuring biopharmaceutical balance, and the justification for using excipients specific to the drying technique was covered in section 4. A variety of drying technologies may minimise the complexity of validation procedures and allow for a successful scale-up using a quality-by-design (QbD) approach, in addition to the aforementioned difficulties. A table with a variety of alternate drying procedures may be found here. Even though some of these methods are advertised as bulk drying, a powder/product filling machine can be used to make single unit doses [Wang, B., McCoy, T.R., 2015].

Biopharmaceutical Drying Technologies:-

Single Dose Drying Technologies:-

Conventional Batch Freeze-drying: -

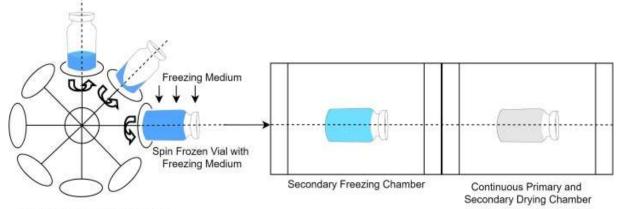
In freeze-drying, water molecules are removed from the solution in order to freeze it. A condenser and a vacuum pump are the most common parts of a batch freeze-dryer. This procedure is divided into three stages: freezing, drying, and drying again [Wang, W., Roberts, C.J., 2018]. Vacuum-sealed vials are placed in the drying room, where they are partly dried. Products are frozen to temperatures between -400° and -600° C. for crystalline additives below the eutectic point (Tm) or for amorphous compounds below the frozen product's glass transition temperature (Tg'). Non-cGMP freeze-drying cycles may use thermocouples and other Wi-Fi temperature sensors to monitor the average product temperature [Broadwin, S.M., 1965]. In addition to crystallising bulking chemicals and improving product consistency, annealing may be used to execute an additional step. Primary drying is a three-step process that begins with the freezing of liquid water and ends with the formation of water vapour. Water vapour may go from the vials to the condenser when the vials are partially stoppered. Sublimation may be achieved with a longer shelf life and lower pressure in the chamber, but the product temperature must remain below its Tg' or Tm. The desorption step is the last stage of secondary drying. This is done to ensure that the final moisture content is at the desired level [Corver, J.A.W.M., 2012].



A significantly higher temperature is used to free water molecules attached to the product's surface while the temperature of the product remains below the solid-kingdom glass transition point during the secondary drying stage (Tg). A silicone stopper and an aluminium lid seal are used to reseal all vials after secondary drying. [Becker, W., 1957] All vials are scrutinised before they are released and shipped.

Spin-freeze-drying of Unit doses: -

Spin-freeze-drying is the name given to this new generation of continuous freeze-drying because of the rotation of vials containing the semi-liquid product of interest along their perpendicular axis. First, the vials containing the liquid product are spun at a high speed, usually between 2500 and 3000 rpm, for a predefined amount of time along their longitudinal axis [De Meyeret al., 2015]. Flowing through the rotating vials is a sterile cryogenic gas such as nitrogen or carbon dioxide gas. During the process of freezing and heating the dispersion layer in the vial, the frozen product spreads over the interior walls, creating a large surface area. In order to accomplish crystallisation and the proper form of the excipients, certain modifications are made to the cooling process settings in a temperature-controlled chamber [Vanbillemontet al., 2020]. Alternatively, the outer floor of the vials may be ringed or covered in order to improve heat dispersion. To remove the water from the vials at the end of the day, they are taken to the secondary drying chamber. For drying, between 30 minutes and two hours are needed. [Lammenset al., 2018]



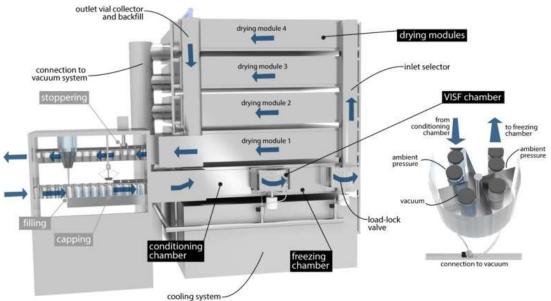
Continuous Spin Freezing of vials

Initiation reduces processing time by ten to forty times, depending on the vial and product system. The spin-frozen vials' sublimation dynamics were influenced by the manner of freezing, but the batch-frozen vials were not. The sublimation rates of spin-frozen vials may be predicted by comparing the sublimation rates at different shelf temperatures and chamber pressures. According to one study, spin-frozen vial sublimation costs were most affected by shelf temperature when the vial was properly in touch with the shelf. [Troutet al., 2018] As with frozen vials, chamber tension will have less of an impact when stored at room temperature. It was thought that this happened because the inside of spinning vials had a much larger surface area for the product than traditional frozen vials [Pisano R., 2019].

Prototypes for spin-freeze drying that mimic GMP have been developed by Rhea Vita and the University of Ghent. Spin freeze drying generation has proven to be a capable competitor with batch freeze-death in terms of method-related stresses and PAT, although the feasibility of implementing > five parallel lines to generate a higher throughput along with the associated costs in a cGMP environment would be fascinating to investigate [Capozziet al., 2019].

Continuous Freeze-drying of suspended vials: -

Non-stop freeze-drying of suspended vials is a new idea that was recently proposed in the PCT patent. Molecules in this approach are linked together in order to maintain a constant flow through the vials, which is an activity unit in itself [Van Der Wel, P.G., 2012]. For the most part, vials are filled and partly sealed before being put in the freezedryer; this is the major reason for their frequent use. This is followed by the freezing process that uses the vacuuminduced surface freezing method (VISF) to begin freezing the vials [Luy, B., Stamato, H., 2020]. A faster sublimation rate means less drying time in the end, saving time over the old method. The vials sustain the passage of heat to the radiating surfaces throughout all phases of freezing. This is a problem with the standard freeze-drying process because warm air isn't spread out in the same way everywhere.

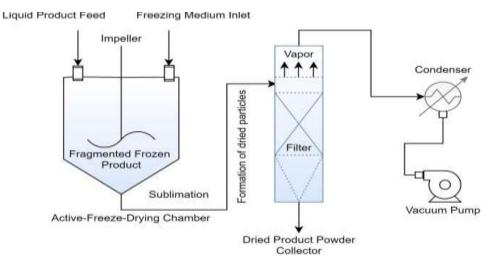


The continuous freeze-drying process took just six hours, compared to a 51-hour batch freeze-drying cycle. Nonprevent freeze-drying requires a scale that is six to eight times smaller than the typical batch freeze-dryer. In the end, non-prevent freeze-drying is more beneficial in terms of slow drying time and PAT for all vials. However, executing this non-prevent approach in a cGMP environment with concomitant scale-up and validation produces better outcomes [Duerkopet al., 2018].

Bulk Drying Technologies: -Active-freeze-drying: -

With little handling, it is possible to dry heat-sensitive bulk materials, from solutions, suspensions, and pastes to wet solids, with little handling using the active-freeze-drying process. [Struschkaet al., 2016] The vacuum dryer, impeller, collecting filter, product collector, condenser, and vacuum pump make up the gliding technique.

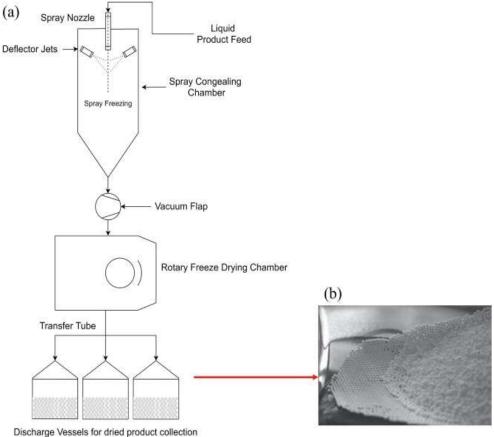
A thermostatically regulated heating and cooling jacket surrounds the chamber. During the sublimation of frozen granules, the granules are spread out in conjunction with the stirring action in order to achieve the desired distribution of the granules. The dry layer is constantly split up into bits due to the swirling motion caused by mixing, thereby decreasing the barrier to vapour flow. When the temperature of the product is equal to the temperature of the chamber wall, the drying process is complete [Sebasti a oet al., 2019].



When tested on nanocrystals at a small scale, the energetic-freeze-drying method worked well for ketoconazole-type medications. Scanning electron microscopy may also be used to reveal large parts (SEM). On the other hand, particle size analysis indicated that there were nanocrystal aggregates ranging in size from 1 to 100 m along the edges of the vials [Lowe D.et al., 2018]. The nanocrystal suspensions are physically stable even after two weeks of storage after active freeze-drying, and this is despite the freeze-drying principle being used to make bulk powder in a batch procedure. It's possible that constant churning in an insulating vessel will raise the ambient temperature somewhat. As a result, it is critical as the product's temperature must reflect it during sublimation. For tiny molecules and stable biopharmaceuticals, active-freeze-drying technology may be more suited. Nevertheless, the practicality of this approach has to be researched for commercially synthesised parenteral biopharmaceuticals. [DeMarcoet al., 2015]

Spray-freezing and Dynamic Freeze-drying Technology: -

Two major processes are involved in this procedure: spray freezing and dynamic bulk freeze drying. Spray freezing is the first step. The floating price, nozzle frequency, viscosity, and orifice diameter all determine the droplet length. According to some estimates, there are 1,000 to 5,000 drops every second. [IMA Life, 2019] The freeze chamber is a cylindrical tank with two sides. It takes time for the droplets to come into contact with N2 (l). Sterile N2 (g) is administered internally at a temperature ranging between -80° C and -1000° C and 50° C. Each 1.5-meter cube of ice typically contains between three hundred and one thousand micrometer-diameter ice spheres of homogenous size. Sprayed freezing, the dynamic freeze-drying technique, uses a revolving freeze-dryer that is placed within an enclosed vacuum chamber and rotates on its longitudinal axis while the temperature rises to freeze the product. The conductive heat switch for the product has been completed and is now ready for use. The exits at both ends of the drum make a big difference in how much water vapour can move through the drum [Siowet al., 2018].

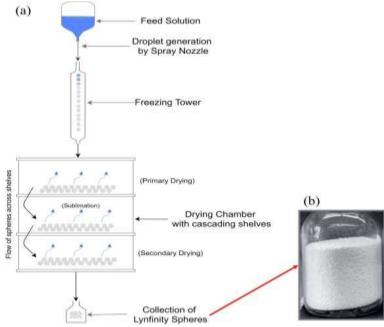


All of these components work together to enhance mass transfer: cryogenic fuel, rotating drum surface, IR radiators, and a massive floor of frozen microspheres. It is also changed by rapid reconstitution and minimum protein aggregation at 25 °C, when loose-flowing powder is modified. Debris loss due to excessive sublimation is minimised when a considerable amount of solid material is present. For feed items with low levels of stable content, producing larger particle sizes, such as 2 to 3 mm, may reduce the risk of particle loss. A 100-liter bulk with a stable content of 20% can be spray-frozen in 10 to 20 hours; however, dynamic freeze-drying with a yield of > 97 percent takes 24 hours [Celik M.et al., 2010].

Continuous Aseptic Spray-freeze-drying Technology by IMA Life: -

IMA Lifestyles invented the Lynfinity[®] Spray-freeze after patent changes. A temperature-controlled droplet zone forces the product feed to produce uniform droplets under the effect of frequency vibrations, which starts the spraying system's development. Low temperatures are maintained in the cooling fluid. Frozen spheres gather in the bottom of the chamber. Cascade shelves sway gently to transfer frozen spheres through the drying chamber at a predetermined pace. The series chamber is used to gather a huge number of dried spheres [Roser B., 1991].

In comparison to traditional freeze-drying, this method is expected to provide benefits such as bulk processing with less handling of trays, as well as higher productivity and decreased downtime. As a second benefit, it facilitates the movement of heat and mass between products and travs. For three reasons, it might be a continuous, high-volume approach in a sterile environment. No matter how well a spray-freeze-dried product is, particle aggregation was shown to cause a small increase in turbidity over a freeze-dried equivalent. Increased surfactant awareness resulted in an improvement in turbidity [Wales T.E., Engen J.R., 2006].



Continuous bulk processing can allow for the use of additional PATs for real-time process and product monitoring, but care must be taken to ensure product yield, stability of spray-freeze-dried parenteral biopharmaceuticals, footprint of an industrial-scale system, and suitable powder filling options in cGMP environments [Vhora I.et al., 2019].

Spray-drying:-

Spray-drying is a technique with several applications in the biopharmaceutical industry. In the first place, it saves money by doing away with the need for a large number of unit operations. As a second benefit, the one-step method makes it possible to produce powders with great flowability in a continuous manner. The evaporation time might be as short as a few milliseconds or as long as a few seconds. For section 3 pivotal research, SPX Glid Inc. has FDA-inspected aseptic spray dryers and synthesised clinical chemicals using aseptic spray drying. The authors looked at how well the Anhydro MS-35 Spray-dryer worked to make mAbs from dry powder. [Wenzel S.et al.,2007]

The liquid supply is drawn toward the spray nozzle at a set pace using a peristaltic pump [Bowen M.et al., 2013]. The wet-bulb temperature is equal to or lower than that of the sprayed droplets in the chamber. The temperature of gasoline production is measured in the bottom drying chamber. Fuel waft prices help drive waste into the cyclone where it may be separated according to density. Protein balance may be altered if the gas leaving the cyclone collector has a temperature greater than or equal to Tg and if the particles remain in the cyclone collector for a longer period of time than Tg. Dry powder biopharmaceuticals can be administered parenterally, inhaled, or nebulized for use in industry and medicine [Albasarah Y.Y.et al., 2010]. Lysozyme's enzymatic activity was shown to be heavily influenced by the temperature of the input. CPPs and spray-drying protein formulation additions are now being proven by the authors. The lysozyme activity was reduced by ultrasonic nozzle-induced vibrations and mechanical stress, in addition to the high outlet temperatures [GangulyA.et al., 2018]. In addition to temperature, shear and interfacial denaturation during atomization and spraying may have a major impact on proteins, which are susceptible to such stresses. When developing and selecting excipients for spray-drying proteins, consideration must be given to the effect that spraying circumstances have on the protein prior to dehydration [Garcia A.et al, 2012]. A two-fluid nozzle's nozzle tip typically has a diameter as it emerges from the liquid feed. An atomizing gas travelling with the flow rate of gasoline forms a resulting speed valve in the integration area. A fluid spray nozzle's shear charge was predicted [Euliss L.E.et al.]. For the past several decades, spray-drying has been studied for a wide range of goods as one of the most p+rominent commercial drying techniques. We need to look into the CPPs for spraydrying and the molecular processes of how excipients interact with parenteral biopharmaceuticals while spraydrying [DeSimone J.M., 2016].

Print® Technology: -

Particle reproduction in non-wetting templates is often referred to as "PRINT" technology. The semiconductor microelectronics industry makes use of this technique, which is descended from lithographic procedures. It is possible to create monodisperse, individually formed micro-and nano-debris of hydrogels, polymers, APIs, and other substances using micro-moulding-based particle layout and engineering [Garcia A.et al., 2012]. Preclinical and medical statements for pharmaceutical inhalation powders using liquid organisation may now be created with nonpreventable particles using a roll-to-roll technology that has been integrated with cGMP processes [Morton, S.W. et al.,2013]. It is done by pouring perfluoropolyether into a closed silicon drawer template that holds selected forms of m, such as 5 m and 200 nm-sized shapes. Sandwiched between the cavities of the PFPE mould and a high ground potential polyethylene film were aqueous protein samples, including insulin, albumin, and albumin mixtures containing siRNA or paclitaxel [Allison S. Det al., 1999]. When these approaches are used together, nanoparticle stability is improved, drug release is delayed, and the physicochemical properties of the drug are improved. As the spraying progressed, this changed to debris loss. A water-washing step is necessary to remove excess cationic polyelectrolytes. Electron microscopy pictures of polyelectrolyte-coated particles [Duncan, P.B., Needham, D., 2006] reveal the form and integrity of the recovered particles. When combined with a stronger immune response, PRINT®-produced vaccine debris boosts antigen-binding by a factor of 200. When these nanoparticles come into contact with certain biomolecules in their surroundings, their surface area, length, and shape are all changed to aid in the interaction. In a section 1 clinical study [Aniketet al., 2015], GSK's PRINT® generated dry-powder ribavirin system has shown superior physicochemical properties with environmentally friendly and easily accessible API delivery to the lungs in a section 1 clinical study [Nguyenet al., 2020]. A few inhaled proteins, gene therapy products, and vaccines have shown promising results with this technology. However, the stability of large parenteral masses and enzymes through PRINT® may be an interesting research area. This technology has the potential for non-preventive manufacturing in large cGMP facilities.

Microglassification: -

Excipients, such as saccharides and polyalcohols, are often utilised to substitute water in protein restoration formulations. MicroglassificationTM is a cutting-edge technology for recovering proteins by drying protein microdroplets in an immiscible drying fluid in line with previously recognised micropipette manipulation procedures [Kammari, R., Topp, E.M., 2020]. Using a micropipette, a drop of the protein solution is released into the organ chamber, where it sticks to the tip of the instrument. MicroglassifiedTM beads were created by extracting a droplet of water from a natural chamber. Fluorescence spectra of native and rehydrated microglassified BSA showed that the tertiary structure of the protein was retained. It was shown that when MicroglassifiedTM BSA was preserved better, the saturation point of water in alcohol was greater [ElKassas K.et al., 2021]. These enzymes were tested in a second experiment using this same method. The results were similar to those of the first experiment. Even after months or years of storage, enzyme activity and secondary structure composition remained relatively stable. Because of water activity [Bjelo'sevi'c M.et al., 2020], MicroglassifiedTM and freeze-dried lysozyme absorb the same amount of water. As a result, the authors hypothesised that the moisture content of both kinds would be identical. The MicroglassificationTM of an elastin-like polypeptide with precise control over its length and form for chemotherapy has also been achieved. Bio-sensors and optical tools have been implanted using this method in the last several years. The application of MicroglassificationTM on commercial enzymes, antibodies, and vaccines, as well as the creation of biopharmaceutical parenteral formulations with high awareness, might be used to improve the utility of the technique yet further. [Kolhe P.et al., 2009]

Biopharmaceutical Characterization: -

To ensure that biopharmaceutical products are safe and effective, the ICH Q5E recommends doing a product comparison job before making any modifications to the manufacturer's production process. For biopharmaceutical goods, it is possible to employ analytical and characterization procedures to collect CQAs in both stable and liquid forms in order to assess any decrease in CQAs after drying. Even though recurrent analysis will provide more information on the influence of CPPs on the development of CQAs, not all methodologies are used [Pinto J.T.et al., 2021]. Mass control release assays include a few chromatographic and spectroscopic techniques; however, they do not give high-selection information on protein stability and interactions with excipients, despite their widespread use. An earlier study using HDX-MS in the context of U.S. protein degradation research looked at the conformational stability of proteins in liquids, freeze-dried solutions, and protein adsorption on solid surfaces. NMR in conjunction with Magnetic Resonance Imaging has been investigated [Ji S.et al., 2016] for the fast reconstitution of freeze-dried items. Fluorescence spectroscopy has been shown to be useful in the reconstitution of freeze-dried BSA. Improvements in CD spectroscopy have also allowed scientists to investigate problematic decreased vacuum

ultraviolet site circumstances. In the fields of protein photostability, photoisomerization, protein-ligand interactions, and RNA characterization, the SRCD is well-known for its tools. Despite the fact that they frequently have their own distinct benefits, characterization approaches may provide insight into biopharmaceutical stability when used in conjunction with one another. When used in conjunction with drying technologies [Rathore, N., Rajan, R.S., 2008], biopharmaceutical companies may find these approaches useful in the production and classification of their medicines.

Formulation Aspects For Drying Technologies: -

Freeze-dried biopharmaceutical products are often prepared using buffers (salts), amino acids, sugar, bulking agents, surfactants, and tonics. Biopharmaceuticals may benefit from Trehalose's high glass transition temperature (> 100 °C), as well as other properties, during spray-drying. Iysozyme may improve the overall stability of Iysozyme if a 2:1 protein-to-sugar ratio is shown to provide an accurate stabilising effect. It has also been shown that the spraydrying of lysozyme with methanol as a co-solvent enhanced the EPF of the spray-dried product above that of the water-based product. The fact that trehalose and sucrose can crystallise in both their frozen and dry states despite the fact that storage at higher temperatures and moisture may cause the protein shape and stability to erode is significant, regardless of their advantages. It is possible, however, that mannitol crystallisation in the absence of amorphous stabilisers will have an adverse effect on protein structure and stability. Solubility and protein balance of protein solutions may be improved by using L-arginine and L-arginine hydrochloride as amino acids. For proteins, arginine may function as the major stabiliser in conjunction with other excipients or by itself. Another benefit of using these amino acids in powder formulations is improved flowability, dispensability, and aerosolization due to the fact that they prevent proteins from being damaged by the high pressures created during atomizationSpray-dried protein formulations often employ leucine, whereas liquid and freeze-dried protein formulations commonly use histidine. At the air-liquid, strong-liquid, and liquid-liquid interfaces, surfactants play an important role in preventing protein aggregation as well. But more research is needed to figure out how stabilisers like freeze-drying and spray-drying work for different drying methods like freeze-drying and spray-drying, which are listed in Table 4.

Scale-Up, Packaging And Validation Aspects For Drying Technologies: -

The most commonly used strategies for moving a pharmaceutical production technology from one site to another are scale-up and period changeover. Fill-finish processes like compounding, filtering, and vial filling, among others, demand extra time for freeze-drying in vials. To make matters worse, in the company's most costly footprint, like the controlled area for aseptic filling, long loading times, unloading durations, and vial inspection times all affect the efficiency of individual operations and cause delays in stock and inventory turnover times. These superfluous components are eliminated when using energy-freeze drying, spray drying, and spray-freeze drying. Major packaging additives, fill-finish line equipment, sterilising techniques, fill quantity, freeze-drying cycles, capping, inspection, and container closing integrity all need further certification and validation. With case drying techniques such as sprinkling or sprinkling and freeze drving, it is simpler to fill containers such as vials, ampoules, syringes, and so on, and the quantity of free-flowing powder is reduced. It is also easier for clinics to administer infusions since several strengths of the drug are available. so that drying processes with well-defined form constraints may be scaled up and used as a platform, and so that business operations can be more resilient. It's tough to explain freezedrying on a small scale since everyone behaves like it's an outstanding drying machine. The real issue is dependent on the vial transfer coefficient, the freezing temperature, and the shelf space available to the sample. There are a number of drying methods that are not as affected by the unpredictable nature of drying in a container, such as energetic-freeze drying, spray drying, and spray freeze drying. Regulatory decisions must be made for the implementation and approval stages of a current company product when it moves from a batch freeze-drying method to an extensive drying process. The opportunity drying method's performance enhancement may be overshadowed by the high cost of biopharmaceutical DS, engineering lines for validation batches, long-term balancing data, and the necessity to propose a revision. Potential drying techniques may also be extremely useful in the development and manufacture of new goods in modern ideas.

Feasibility Of Pat For Drying Technologies: -

The ICH Q8 (R2) ideas for 'Pharmaceutical Development' include PAT, a QbD strategy for creating, analysing, and managing data. Enhancing and optimising freeze-drying cycles is the major goal of the approach. Temperature probes and shelf temperature are used in conjunction with Pirani gauge and capacitance manometer measurements to arrive at the drying give-up point. As a result of the prior secondary drying step, MS has been shown to be quite sensitive to a median cake moisture of 3%. However, it might be used to prevent silicon oil and helium gas leaks in a freeze-dryer. Due to the past restrictions associated with this equipment during batch freeze-drying, TDLAS and

MS will most likely employ the continuous drying technologies to calculate the RMC for each individual vial and bulk item. When it comes to spin-freeze drying, the scientists find that the NIR-CI and 4D Micro-Computed X-ray tomography and imaging capability packages work well. The connection between Karl Fisher analysis and NIR spectroscopy may help predict the RMC post. All NIR, MIR, and Raman spectroscopies, which had previously functioned as capacity pats, may be employed in-line or at-line with this approach to analyse all dried vials or bulk objects manufactured by various drying methods. They are used for process control to assure the quality of infant formula and dairy component powders. NIR and Raman spectroscopy have been used in conjunction to analyse the protein's stability. Despite the fact that most vials transit through different drying chambers during continuous freeze-drying of suspended vials, it is feasible to analyse vials using an NIR or Raman probe. It is also possible to use a probe to load powder into unit dosage forms while the product is being dried using a variety of methods, including spray drying and spray-freeze drying. These PATs have a substantial impact on the market and may reduce the amount of time needed for batch release testing. One of the few powder-based goods that doesn't need CQA is particle length. Spray-drying particle length distributions have been measured using in-line and at-line laser diffraction. The length of dry powder particles was measured using a variety of PATs, but only SFV and FBRM proved to be reliable.

Conclusion: -

The gold standard for drying biopharmaceutical goods is still batch freeze-drying at the moment. Solid biopharmaceuticals may now be produced safely, effectively, and efficiently using a variety of drying techniques that have been shown to be more promising in recent years. CPPs, comprising temperature, shear, and other parameters such as these, impact the production of CQAs and are the fundamental criteria for choosing drying processes, even while opportunity techniques allow continuous output at low operating costs. A variety of drying technologies, including spin-freeze-drying, spray-freeze-drying, spray-drying, and MicroglassificationTM, have shown their capacity to increase the stability of inhaled biopharmaceuticals and certain proteins. With CPPs and the proper selection of formulation additives for the drying technique and the product, ensuring the product is vital. It is also unknown how biopharmaceuticals with unique excipients in a strong country work at the molecular level. It is possible to quickly analyse large amounts of data using some of the finest characterization methods and PATs. The vast majority of other drying technologies, on the other hand, may provide large advantages when combined with PATs, but their economic feasibility needs more examination. There are a few possible drying techniques that reduce the complexity associated with validating a few entire end-unit processes better than others when it comes to scaling up, packaging, and validation elements. In the biopharmaceutical business, the industrial scale-up techniques for different drying technologies have been tried out and shown to work.

References:-

- 1. Declerck, P.J., 2012. Biologicals and biosimilars: a review of the science and its implications. Generics BiosimilarsInitiat. J. 1, 13–16. https://doi.org/10.5639/ gabij.2012.0101.005.
- Gervasi, V., Dall Agnol, R., Cullen, S., McCoy, T., Vucen, S., Crean, A., 2018. Parenteral protein formulations: An overview of approved products within the European Union. Eur. J. Pharm. Biopharm. 131, 8–24. https://doi.org/10.1016/j.ejpb.2018.07.011.
- Martagan, T., Akcay, A., Koek, M., Adan, I., 2020. Optimal production decisions in biopharmaceutical fill-andfinish operations. IISE Trans. 53, 149–163. https://doi.org/10.1080/24725854.2020.1770902.
- 4. Tang, X., Pikal, M.J., 2004. Design of Freeze-Drying Processes for Pharmaceuticals: Practical Advice. Pharm. Res. https://doi.org/10.1023/B: PHAM.0000016234.73023.75.
- 5. Rambhatla, S., Pikal, M.J., 2003. Heat and mass transfer scale-up issues during freezedrying, I: Atypical radiation and the edge vial effect. AAPS PharmSciTech 2003 42 4, 22–31. https://doi.org/10.1208/PT040214.
- Pisano, R., 2019. Alternative methods of controlling nucleation in freeze drying. In: Ward, K.R., Matejtschuk, P. (Eds.), Lyophilization of Pharmaceuticals and Biologicals: New Technologies and Approaches. Springer, Methods in Pharmacology and Toxicology, pp. 79–111. https://doi.org/10.1007/978-1-4939-8928-7_4.
- 7. Al-Hussein, A., Gieseler, H., 2012. The effect of mannitol crystallization in mannitolsucrose systems on LDH stability during freeze-drying. J. Pharm. Sci. https://doi.org/10.1002/jps.23173.
- Wang, B., McCoy, T.R., 2015. Advances in Process Analytical Technology in FreezeDrying. In: Varshney, D., Singh, M. (Eds.), Lyophilized Biologics and Vaccines. Springer, New York, pp. 157–178. https://doi.org/10.1007/978-1-4939-2383-0_8.
- 9. Wang, W., Roberts, C.J., 2018. Protein aggregation Mechanisms, detection, and control. Int. J. Pharm. 550, 251–268. https://doi.org/10.1016/j.

- 10. Broadwin, S.M., 1965. Centrifugal freeze-drying apparatus. US3203108A.
- 11. Corver, J.A.W.M., 2012. Method and System for Freeze-Drying Injectable Compositions, In Particular Pharmaceutical Compositions. US10670336B2.
- 12. Becker, W., 1957. Gefriertrocknungsverfahren. DE967120.
- De Meyer, L., Van Bockstal, P.J., Corver, J., Vervaet, C., Remon, J.P., De Beer, T., 2015. Evaluation of spin freezing versus conventional freezing as part of a continuous pharmaceutical freeze-drying concept for unit doses. Int. J. Pharm. 496, 75–85. https://doi.org/10.1016/j.ijpharm.2015.05.025.
- Vanbillemont, B., Lammens, J., Goethals, W., Vervaet, C., Boone, M.N., De Beer, T., 2020b. 4D Micro-Computed X-ray Tomography as a Tool to Determine Critical Process and Product Information of Spin Freeze-Dried Unit Doses. Pharmaceutics 12, 430. https://doi.org/10.3390/pharmaceutics12050430.
- Lammens, J., Mortier, S.T.F.C., De Meyer, L., Vanbillemont, B., Van Bockstal, P.J., Van Herck, S., Corver, J., Nopens, I., Vanhoorne, V., De Geest, B.G., De Beer, T., Vervaet, C., 2018. The relevance of shear, sedimentation and diffusion during spin freezing, as potential first step of a continuous freeze-drying process for unit doses. Int. J. Pharm. https://doi.org/10.1016/j.ijpharm.2018.01.009.
- 16. Trout, B.L., Pisano, R., Capozzi, L.C., 2018. Freeze-drying methods and related products. WO2018204484A1.
- Pisano R., 2019. Alternative methods of controlling nucleation in freeze drying. In: Ward, K.R., Matejtschuk, P. (Eds.), Lyophilization of Pharmaceuticals and Biologicals: New Technologies and Approaches. Springer, Methods in Pharmacology and Toxicology, pp. 79–111. https://doi.org/10.1007/978-1-4939-8928-7_4
- Capozzi, L.C., Trout, B.L., Pisano, R., 2019. From Batch to Continuous: Freeze-Drying of Suspended Vials for Pharmaceuticals in Unit-Doses. Ind. Eng. Chem. Res. 58, 1635–1649. https://doi.org/10.1021/acs.iecr.8b02886.
- 19. Van Der Wel, P.G., 2012. Stirred freeze drying. EP1601919A2.
- Luy, B., Stamato, H., 2020. Spray Freeze Drying. In: Ohtake, S., Iztutsu, K.-I., LechugaBallesteros, D. (Eds.), Drying Technologies for Biotechnology and Pharmaceutical Applications. Wiley, pp. 217–237. https://doi.org/10.1002/9783527802104.ch8.
- Duerkop, M., Berger, E., Dürauer, A., Jungbauer, A., 2018. Impact of Cavitation, High Shear Stress and Air/Liquid Interfaces on Protein Aggregation. Biotechnol. J. 13, 1800062. https://doi.org/10.1002/biot.201800062.
- 22. Struschka, M., Plitzko, M., Gebhard, T., Luy, B., 2016. Rotary drum for use in a vacuum freeze-dryer. US9347707B2.
- Sebasti[~] ao, I.B., Bhatnagar, B., Tchessalov, S., Ohtake, S., Plitzko, M., Luy, B., Alexenko, A., 2019b. Bulk Dynamic Spray Freeze-Drying Part 2: Model-Based Parametric Study for Spray-Freezing Process Characterization. J. Pharm. Sci. 108, 2075–2085. https://doi.org/10.1016/j.xphs.2019.01.011.
- 24. Lowe D., Mehta M., Govindan G., Gupta K., 2018. Spray Freeze-Drying Technology: Enabling Flexibility of Supply Chain and Drug-Product Presentation for Biologics BioProcessInternationalBioProcess International. Bioprocess Int. URL https://bio processintl.com/manufacturing/supply-chain/spray-freeze-drying-technology-en abling-flexibility-of-supply-chain-and-drug-product-presentation-for-biologics/
- 25. DeMarco, Francis, Renzi, Ernesto, 2015. Bulk Freeze-Drying Using Spray Freezing and Stirred Drying. US9052138B2.
- 26. IMA Life, 2019. LYNFINITY: Continuous Aseptic Spray Freeze Drying. Process, Technology and Product Characterization. URL: https://ima.it/pharma/pape r/lynfinity-continuous-aseptic-spray-freeze-drying-process-technology-and-pro duct-characterization/
- Siow, C.R.S., Heng, P.W.S., Chan, L.W., 2018. Bulk Freeze-Drying Milling: a Versatile Method of Developing Highly Porous Cushioning Excipients for Compacted MultipleUnit Pellet Systems (MUPS). AAPS PharmSciTech 19, 845–857. https://doi.org/10.1208/s12249-017-0899-6.
- 28. Celik M., Wendell, S.C., 2010. Spray Drying and Pharmaceutical Applications, in: Parikh, D.M. (Ed.), Handbook of Pharmaceutical Granulation Technology. Taylor and Francis, pp. 98–125. https://doi.org/10.1201/9780849354953.ch5.
- 29. Roser, B., 1991. Trehalose, a new approach to premium dried foods. Trends Food Sci. Technol. 2, 166–169. https://doi.org/10.1016/0924-2244(91)90671-5.
- 30. Wales, T.E., Engen, J.R., 2006. Hydrogen exchange mass spectrometry for the analysis of protein dynamics. Mass Spectrom. Rev. 25, 158–170. https://doi.org/10.1002/ mas.20064.
- Vhora I., Bardoliwala D., Ranamalla S.R., Javia A., 2019. Parenteral controlled and prolonged drug delivery systems: Therapeutic needs and formulation strategies. Nov. Drug Deliv. Technol. Innov. Strateg. Drug Repositioning 183–260. https://doi.org/ 10.1007/978-981-13-3642-3_7.

- 32. Wenzel S., Wilbraham D., Fuller R., Getz E.B., Longphre M., 2007. Effect of an interleukin-4 variant on late phase asthmatic response to allergen challenge in asthmatic patients: results of two phase 2a studies. Lancet 370, 1422–1431. https://doi.org/10.1016/S0140-6736(07)61600-6.
- Bowen M., Turok R., Maa Y.F., 2013. Spray Drying of Monoclonal Antibodies: Investigating Powder-Based Biologic Drug Substance Bulk Storage. Dry. Technol. 31, 1441–1450. https://doi.org/10.1080/07373937.2013.796968.
- 34. Albasarah Y.Y., Somavarapu S., Taylor K.M.G., 2010. Stabilizing protein formulations during air-jet nebulization. Int. J. Pharm. 402, 140–145. https://doi.org/10.1016/j. ijpharm.2010.09.042.
- Ganguly A., Stewart J., Rhoden A., Volny M., Saad N., 2018. Mass spectrometry in freeze-drying: Motivations for using a bespoke PAT for laboratory and production environment. Eur. J. Pharm. Biopharm. 127, 298–308. https://doi.org/10.1016/j. ejpb.2018.02.036.
- Garcia A., Mack P., Williams S., Fromen C., Shen T., Tully J., Pillai J., Kuehl P., Napier M., Desimone J.M., Maynor B.W., 2012. Microfabricated Engineered Particle Systems for Respiratory Drug Delivery and Other Pharmaceutical Applications. J. Drug Deliv. 2012 https://doi.org/10.1155/2012/941243.
- 37. Euliss L.E., DuPont J.A., Gratton S., DeSimone J., 2006. Imparting size, shape, and composition control of materials for nanomedicine. Chem. Soc. Rev. 35, 1095–1104. https://doi.org/10.1039/b600913c.
- 38. DeSimone J.M., 2016. Co-opting Moore's law: Therapeutics, vaccines and interfacially active particles manufactured via PRINT®. J. Control. Release 240, 541–543. https://doi.org/10.1016/j.jconrel.2016.07.019.
- Garcia, A., Mack, P., Williams, S., Fromen, C., Shen, T., Tully, J., Pillai, J., Kuehl, P., Napier, M., Desimone, J.M., Maynor, B.W., 2012. Microfabricated Engineered Particle Systems for Respiratory Drug Delivery and Other Pharmaceutical Applications. J. Drug Deliv. 2012 https://doi.org/10.1155/2012/941243.
- Morton, S.W., Herlihy, K.P., Shopsowitz, K.E., Deng, Z.J., Chu, K.S., Bowerman, C.J., Desimone, J.M., Hammond, P.T., 2013. Scalable manufacture of built-to-order nanomedicine: Spray-assisted layer-by-layer functionalization of PRINT nanoparticles. Adv. Mater. 25, 4707–4713. https://doi.org/10.1002/ adma.201302025.
- 41. Allison, S.D., Chang, B., Randolph, T.W., Carpenter, J.F., 1999. Hydrogen bonding between sugar and protein is responsible for inhibition of dehydration-induced protein unfolding. Arch. Biochem. Biophys. 365, 289–298. https://doi.org/10.1006/ abbi.1999.1175.
- 42. Duncan, P.B., Needham, D., 2006. Microdroplet dissolution into a second-phase solvent using a micropipet technique: Test of the epstein-plesset model for an aniline-water system. Langmuir 22, 4190–4197. https://doi.org/10.1021/la053314e.
- 43. Aniket, Gaul, D.A., Bitterfield, D.L., Su, J.T., Li, V.M., Singh, I., Morton, J., Needham, D., 2015a. Enzyme dehydration using microglassificationTM preserves the Protein's structure and function. J. Pharm. Sci. 104, 640–651. https://doi.org/10.1002/ jps.24279.
- 44. Nguyen, T. Van, Ta, V.D., 2020. High-quality factor, biological microsphere and microhemisphere lasers fabricated by a single solution process. Opt. Commun. 465, 125647 https://doi.org/10.1016/j.optcom.2020.125647.
- Kammari, R., Topp, E.M., 2020. Effects of Secondary Structure on Solid-State HydrogenDeuterium Exchange in Model α-Helix and β-Sheet Peptides. Mol. Pharm. 17, 3501–3512. https://doi.org/10.1021/acs.molpharmaceut.0c00521.
- ElKassas, K., Chullipalliyalil, K., McAuliffe, M., Vucen, S., Crean, A., 2021. Fluorescence spectroscopy for the determination of reconstitution time of an in-vial lyophilised product. Int. J. Pharm. 597, 120368 https://doi.org/10.1016/j. ijpharm.2021.120368.
- 47. Bjelo`sevi'c M., ZvonarPobirk A., Planin`sek O., AhlinGrabnar P., 2020. Excipients in freeze-dried biopharmaceuticals: Contributions toward formulation stability and lyophilisation cycle optimisation. Int. J. Pharm. https://doi.org/10.1016/j. ijpharm.2020.119029.
- 48. Kolhe P., Amend E., Singh K., 2009. Impact of freezing on pH of buffered solutions and consequences for monoclonal antibody aggregation. Biotechnol. Prog. 26, 727–733. https://doi.org/10.1002/btpr.377.
- 49. Pinto J.T., Faulhammer E., Dieplinger J., Dekner M., Makert C., Nieder M., Paudel A., 2021. Progress in spraydrying of protein pharmaceuticals: Literature analysis of trends in formulation and process attributes. Dry. Technol. 1–32 https://doi.org/10.1080/07373937.2021.1903032.
- 50. Ji S., Thulstrup P.W., Mu H., Hansen S.H., van de Weert M., Rantanen J., Yang M., 2016. Effect of ethanol as a co-solvent on the aerosol performance and stability of spray-dried lysozyme. Int. J. Pharm. https://doi.org/10.1016/j. ijpharm.2016.09.025.
- 51. Rathore, N., Rajan, R.S., 2008. Current Perspectives on Stability of Protein Drug Products during Formulation, Fill and Finish Operations. Biotechnol. Prog. 24, 504–514. https://doi.org/10.1021/bp070462h.