



RESEARCH ARTICLE

BIOCONVERSION OF LIGNOCELLULOSIC BIOMASS TO ETHANOL USING DIFFERENT MICROORGANISMS

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Abstract

Lignocellulosic material that includes hemicellulose, cellulose and lignin (lignocellulosic complex) is present in the plant cells. The hydrolysis process of the lignocellulose biomass into glucose in the presence of lignocellulolytic enzymes is an area of concern in the production process of cellulosic biofuel. Microorganisms like fungi have the ability for degrading the plant cell wall by an enzyme set which acts in coordination. This moves in a direction to release glucose freely. Another challenge is the modification in the plant cell architecture. Along with this, the capacity of microorganisms in degradation by the modification of the genomes is also one of the challenges. The advantage of the biological process of pre-treatment for degradation of the lignocellulosic materials is because of its effective enzymatic system. There are two types of enzymatic systems which is of extracellular nature in fungi. These are hydrolytic and ligninolytic systems. Hydrolases are produced by hydrolytic system which degrades the polysaccharide and produces sugar. The exclusive oxidative advantage and the extracellular ligninolytic system degrades the components of lignin and also opens the rings of phenyl. The reducing sugars are then converted in ethanol production with the use of various fermentative microorganisms. In this paper, the bioconversion of lignocellulosic biomass to ethanol using different microorganisms is discussed along with other relevant aspects.

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Introduction:-

Petroleum is a very important natural resource and it is depleting at a rapid rate as its consumption is high around the world (Pop 2018). The development of alternative sources of energy is the area of increasing study as the it may provide options for replacement of fossil fuel used for transportation. (Alper and Stephanopoulos 2009; Gupta et al. 2009). Biofuel has been found to have many advantages like less pollution, low on toxicity, biodegradability. It also produces lesser pollutants which are air-borne than those produced by current options of fuel such as petroleum. Ethanol with the mentioned benefits has an advantage of being easily integrated in the present vehicles (Campbell et al. 2019; McCarthy and Tiemann, 1998). Lignocellulosic biomass denotes a huge amount of renewal resources which are carbon neutral for the Bioethanol production (Ragauskas et al. 2006). Lignin, hemicellulose

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and cellulose are the main components of lignocellulosic biomass (Yu, Paterson, Blamey & Millan 2017; Kuhad et al. 1997). A bioconversion process is important for this which has three major steps. The first step is pre-treatment, followed by hydrolysis and finally fermentation. The process of pre-treatment is required for the removal of lignin and hemicellulose fraction. This increases the porosity of materials and also decreases the crystallinity of cellulose. Biological pre-treatment by microorganisms like fungus (brown, white and soft rot fungi) and bacteria has been conducted by researchers. It has been displayed that this increases the productivity of the hydrolysis and the generation of the inhibitors are low and the utilisation of the energy is also shown low. (Wang et al. 2013; Taha et al. 2015; Hatakka 2005). Along with this the biological pre-treatment is also found to be substantially reasonable in terms of cost effectiveness in comparison to other processes of pre-treatment. These biological pre-treatment processes have been comparatively investigated in lesser numbers (Chaturvedi and Verma 2013). In the degradation of lignin, white rot fungi for the soft and brown fungi has been the most efficient which attacks cellulose directly among others in the methods of pre-treatment (Kang et al. 2013; Fernandez et al. 2012; Suhara et al. 2012). In addition to these, the white rot fungi also cause lesser damage to the environment and lesser consumption of energy (Narayanswamy et al. 2013; Kumar et al. 2015). Their enzymatic system is very efficient which is the main reason of their ability for biological pre-treatment in the degradation of the lignocellulosic. The effectiveness of the microorganisms along with the white-rot fungi (WRF) is studied in this paper for the process of biological pre-treatment and further in the process of fermentation for the production of bioethanol.

The ethanol biofuel- a brief history

The modern history has witnessed the usage of ethanol as fuel in various ways (Dagle 2020). It is to be mentioned that the fuel used in the invention of ignition engines was bioethanol. Ethanol was also used as illuminators which was used in 1850s. Approximately ninety million gallons of ethanol production were seen in the United States of America. The changes in its status was seen after the imposition of the tax on the ethanol. This was done to provide assistance in the finances during the civil war. Ethanol was replaced as premier illuminant in 1860s by kerosene which became more affordable and cheaper (Rasmussen 2019; Morris 1993). Subsequently, in the year 1906, the tax on alcohol was lifted. This brought back the inclination in ethanol. In a breakthrough development, Henry Ford designed an automobile car of Model T which ran on ethanol in the year 1908. The production of ethanol was sufficient by the year 1914 and it reached a supply of approximately ten million gallons but petroleum emerged as a fuel in 1919 putting ethanol at the backseat and consequently the production and usage of ethanol as fuel reduced again. Later, The World War II brought back the increase in production of ethanol in the years of early 1940s. It was required for making the synthetic rubber. An annual production of about six hundred gallons of ethanol was produced in the United States of America during this time period (Morris 1993). The demand of ethanol had decreased by the end of the second world war and it continued to reduce for two more decades. This was mostly due to the petroleum imports which were cheaper. This again witnessed changes from the year 1973 due to the oil embargo by the Arab countries. This resulted in the distinct enhancement in the prices of gasoline (Campbell and Laherrere 1998). From the early years of 1970s the shortage and fast depletion of the petroleum has increased the concerns. The crude oil prices have been rising since then. The political instability in different countries around the globe have further added to the concern related to the fuel consumption and prices. With these factors, biofuel is yet again being considered as an alternative biofuel worldwide.

Bioethanol Generations: First and Second

Bioethanol First generation

The bioethanol of first generation was produced by the sugar fermentation like fruit juices, sugarcane juices, sugar beet juices, molasses, and other similar sugar sources. In addition to these, starchy feedstocks like potato, corn and wheat were also utilised for the same (Antoniet al. 2007; Baig et al. 2019). The methods used for the production of ethanol in the first generation utilized the enzymatic digestion methods for releasing sugars from the stored starches. This was followed by processes of the sugar fermentation, distillation method and finally drying. As indicated by the Energy Information Administration (EIA) 2008, the bioethanol of the first-generation had a substantial part in the formation of the policy drivers and the related infrastructure which was needed for the renewal fuels to support the transport in the global market. The International Energy Agency (IEA) 2008, however has expressed various concerns over the related drawbacks of the bioethanol of first generation. Some of these concerns were, impact on environment, Competition between food vs fuel, Multi-feedstock flexibility and similar other factors.

Bioethanol: Second generations

It is, now, understood that the bioethanol production in the first-generation had an approach which was unsustainable (Branco et al. 2019). The increasing criticisms also brought forward the usage of crops which were non-

food for second-generation bioethanol production. This production was from the lignocellulosic biomass which comprised of the residual from the parts of the crops which were non-food parts. The other crops which were not utilized for the purposes of food along with the industrial, municipal and construction waste. It is expected to decrease the net emission of carbon, enhance the efficiency of the energy and also decrease the dependency on energy. It would also help to overcome the limitations of biofuels from the first-generation (Antizar et al. 2008). Along with these the other benefits can be seen in the long-term sustainability and the nature of ethanol which is renewable. Due to these advantages the switching of existing options to cellulosic ethanol is recommended (IEA, 2008). There is scope for much work in the areas of improvement of the technology for the second-generation biofuel. Along with this the reduction of the production cost and further improvement in the reliability and performance in the conversion process are also to be looked at, moving ahead further.

Plant Biomass

Plant biomass is a material that is present in abundance on the planet around the globe. These are available as a natural resource which are renewable (Prather et al. 2020). Lignocellulose contains 3 main polymers which are lignin, cellulose and hemicellulose (Figure 1). These are together called as lignocellulose. Their chemical properties make them a substrate for extensive usage in the products related to biotechnological processes (Kuhad and Singh, 1993; Kuhad et al. 1997; Kuhad et al. 2007).

Cellulose

Cellulose is a glucan polymer of D-glucopyranose units, that are linked with each other by β -1,4-glucosidic bonds (Mansora et al. 2019). The wood cellulose has an average degree of polymerization (DP) of at least 9,000–10,000 and possibly as high as 15,000. An average DP of 10,000 corresponds to the linear chain length of approximately 5 μ m in wood. An approximate molecular weight for cellulose ranges from about 10,000 to 150,000 Dalton (Goring and Timell 1962).

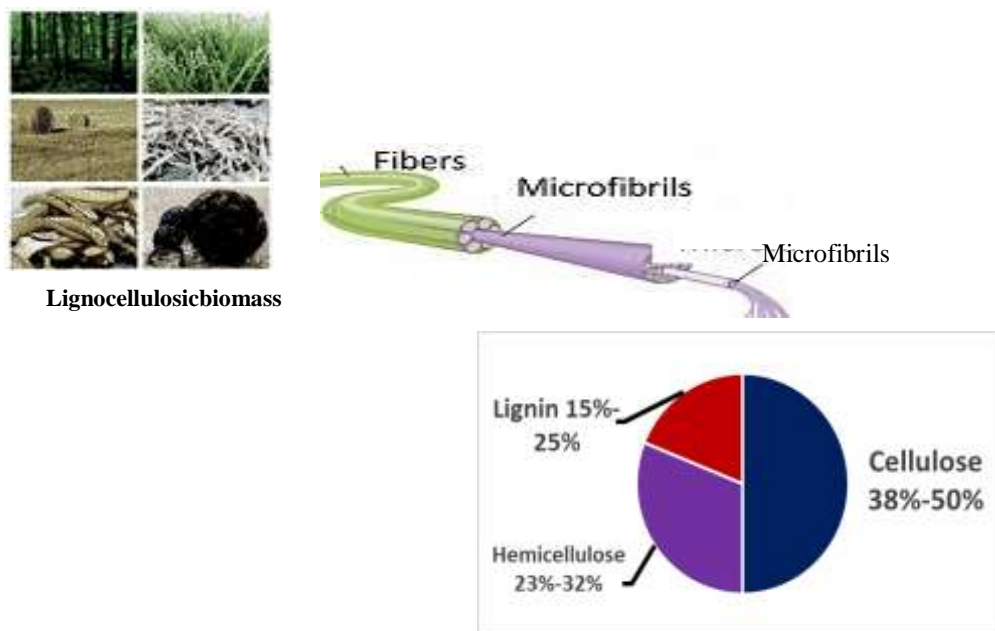


Figure1:- LignocellulosicBiomass components.

Bioconversion – ethanol from lignocellulosic biomass

The bioconversion to obtain ethanol from lignocelluloses is comprised of 2 major processes (Singh et al. 2018). The first process is the hydrolysis of lignocellulosic carbohydrate. It converts the lignin to the reducing sugars for further fermentation. The second process is fermentation of the reduced sugar to the bio-ethanol. The initiation of the process of hydrolysis is generally catalysed by the cellulase enzymes whereas the process of fermentation is by yeasts or bacteria. Hydrolysis of cellulose can be affected by many factors. Some of these factors may include the crystallinity of cellulose fibre, the porosity (accessible surface area) of the waste material and the content of hemicellulose and lignin (Mosier et al. 2005; Kuhad et al. 2011a). The access of cellulase enzymes is difficult due to

the presence of lignin and hemicellulose in the lignocellulosic materials. This reduces the hydrolysis efficiency (Himmelet al.2007). The hydrolysis efficiency is significantly improved due to the process of pre-treatment of lignocellulosic biomass carried out before hydrolysis. This is because of the hemicellulose and lignin is removed and the crystallinity is also reduced in the process. Along with this, the porosity is also increased (McMillan,1994; Mosier et al. 2005; Kuhad et al., 2011 a).

Pre-treatment of lignocellulosic biomass

The advantages of the strategies of pre-treatment process are well received and recommended from a long time (Mosier et al.2005; Sanchez and Cardona,2008; Kuhad et al.2011a). There are certain requirements which are to be met by the strategies for the pre-treatment (Jung & Kim 2017). One of the requirements for the process of pre-treatment is improving the sugar formation or the capacity to further form the sugars by the enzymatic hydrolysis. Next requirement for the process is avoiding the loss or degradation of the carbohydrate. Similarly, avoiding the hydrolysis formation and the by-products that inhibit fermentation is another requirement. In addition to this, the cost-effectiveness of the process is also required to be looked at. There are many biological, chemical, physio – chemical and physical processes that is being used for the pre-treatment of the lignocellulosic materials.

Bioconversion - Hydrolysis and Fermentation

The process of pre-treatment is necessary to enhance the process of hydrolysis of lignocelluloses and subsequently ethanol production. Biological delignification of the materials from plants has several advantages over the pre-treatment methods conducted by mechanical and chemical methods (Kocher et al. 2017). These advantages include lesser reactor resistance on pressure and corrosion, mild conditions of the reaction, lesser side reactions, fewer energy demand, avoidance of the usage of the toxic and corrosive chemicals and higher product yield. (Lee, 1997; Kuhar et al., 2008; Sanchez, 2009). The delignification from microbes seems to be the most feasible method for an improved output of the depolymerization of cellulose and hemicellulose. One of the most effective microorganisms for the pre-treatment carried out biologically are the white-rot fungi (WRF) due to their property of degrading the lignin in a more extensive and in a rapid way in comparison to the other groups of microorganisms. (Eriksson,1993; Kuhad et al.1997; Keller et al., 2003; Kuhad et al.2007). Few WRF are reported to selectively degrade lignin. The ability to some WRF have the opportunity to be utilised for the delignification of the plant materials without having much impact on the cellulose (Kuhad et al.2008; Gupta et al.2011b). Therefore, it is to be mentioned that the selected WRF that degrades lignin and has relatively lower xylanase and cellulase activities can be of advantage in the effective delignification. This can also have an advantage further in reducing the energy and chemical inputs for enzymatic or chemical hydrolysis of the related material.

There are two types of extracellular enzymatic systems of fungi. These are ligninolytic and hydrolytic systems. Hydrolases are produced by the hydrolytic system (Asif et al. 2017). These are responsible for producing sugar by degrading polysaccharides. This exclusive character of being oxidative and the degrading of lignin component by the extracellular ligninolytic system opens the rings of phenyl (Lundell et al. 2010). The fungi WRF produces one or more of such cellular enzymes. These are important for the lignin degradation. They produce major hydrolytic enzymes like xylanase, amylases, and others. Heme contains manganese peroxidase (MSP), Laccase contains copper (Lac), lignin peroxidase (LiP), and also aryl alcohol oxidase (AAO) (Rosales et al. 2002; Elisashvili et al. 2006). These significant enzymes of the fungi WRF are very important in the productive and efficient bioconversion of plant residues (Radhika et al. 2013; Songulashvili et al. 2006).

Some studies have mentioned that WRF is effective in the pre-treatment of the plant biomass on the cellulose hydrolysis. As mentioned by Hatakka(1983), the percentage of conversion of wheat straw to reducing sugar after the pretreatment with *Pleurotus ostreatus* for a time duration of five weeks were found to be thirty-five. Taniguchi et al., (2005) also reported the similar rate of conversion in the material obtained from rice straw when pre-treated with the microorganism *P.ostreatus* for a time span of sixty days. Similarly, Keller et al., (2003) also recorded an improvement from three to five times in the digestibility of the enzymatic cellulose in the corn-stover pre-treatment with *Coriolus versicolor* in a time span over thirty days. The longer incubation time periods have impacted the fungal pre-treatments negatively (Mishra 2018). There is a requirement for more tests on the basidiomycetous fungi to validate for economizing the microbial pre-treatment of the lignocellulosic materials, for improving the hydrolysis of carbohydrates into reducing sugar and for improving the yield of ethanol (Kuhad et al., 2011a).

There are studies conducted in different combinations of other technologies of pre-treatment along with biological method of pre-treatment. (Itoh et al., 2003; Balan et al., 2008). Itoh et al. (2003) indicated ethanol is produced by

simultaneous saccharification and fermentation (SSF) from the chips of beech wood after the pre-treatment by bio-organosolvation from the process of ethanolysis with WRF, *Coriolus versicolor*, *Ceriporiopsis subvermispora*, *Pleurotus ostreatus* and *Dichomitus squalens*.

The ethanol yield produced was found to be 0.294 g/g pulp of ethanolysis and an amount of 0.176 g/g of wood chips were obtained with the pre-treatment by *C. subvermispora* for a time period of eight weeks. This output was 1.6 times more in comparison to the amount received in absence of the treatments with fungus. This combination in the processes enabled the separation of hemicelluloses, celluloses and lignin with the usage of WRF, ethanol and water only. The biological methods of pre-treatments curtailed about 15% electricity that was required for the process of ethanolysis. Balan et al., (2008) indicated in their studies that the impact of treatment by fungus on rice straw followed by the pre-treatment method of AFEX and hydrolysis by enzymatic process. The study reported that the treatment on the rice straw by WRF, followed by AFEX showed distinctively larger conversions of xylan and glucan. Similarly, it is found that the process of SSF with *Lantana camara* and *Prosopis juliflora* which is followed by hydrolysis by acid is observed that the fungal treatment distinctly decreases the inhibitors produced and further decreases the detoxifying agent (Gupta et al., 2011b).

The biomass, rich in cellulose, which is pre-treated is followed by the process of hydrolysis. This is conducted with a combination of cellulolytic enzymes which are β -glucosidase, endocellulase and exocellulase. These enzymes act together and hydrolyse the polymers of cellulose to simpler sugars called glucose (Zhanget al. 2006; Kuhadet al. 2011a). The process of hydrolysis by enzymatic action has shown enhanced results for the further process of fermentation due to the non-formation of the degradation parts of glucose. The process, however is not cost effective (Sanchez and Cardona 2008). Many methods for the better output and other improvements are being tried from past years for decreasing the cost of enzymes, re-usage of the enzymes, increased production of the enzymes and use of genetically engineered advanced systems (Zhanget al. 2006; Kuhadet al. 2011a). It is to be mentioned that the need for searching a better and more competent solution still persists.

As lignocellulose has both pentose and hexose sugars, a pre-requisite is there for the efficient fermentation of both the sugars for cost-effectiveness of the bioethanol production. There are many microorganisms which are known for the fermentation of the hexose sugar whereas there are limited numbers of microbes for the fermentation of pentose sugar. *Pachysolentannophilus*, *Pichiastipitis* and *Candida shehatae* are some of the most common microbes for the fermentation of pentose sugar (Abbietal., 1996a, b; Kuhadet al. 2011a). The reports suggest that the mentioned yeasts have not been very promising with the output received and there are increased steps in the direction of the utilising all the sugars in the hydrolysates by engaging both the genetic manipulations and also the improvement in the approaches towards the process implementation. At present, the common commercially available cellulases are produced from the micro-organisms *Trichoderma reesei*, *Phanerochaete* and *Aspergillus* (Adney et al. 2003). These are mostly used for the description of the mixture the cellulolytic enzymes. The coordinated action of which is needed for enhanced breakdown to monomeric units of its substrates.

The actions of endoglucanases (endo-1, 4- β -glucanases, EGs) in combination is involved in the action of cellulases. These can hydrolyse the internal bonds in the regions of cellulose amorphous with the release of newer ends of terminals. This attacks randomly the internal linkages of β 1,4-linkages cellobiohydrolase (exo-1, 4 β -glucanases, Cbhs). These act on the endoglucanase which generates the chain ends. Amorphous cellulose is degraded by both the enzymes with few exceptions. The only enzymes which efficiently degrades the crystalline cellulose are Cbhs. Cellobiose molecules are released by Cbhs and EGs. This gives the units of cellobiose from the non-reducing ends of the β 3-glucosidase and glucon. This hydrolyses glucose from cellobiose. Hydrolysis can be carried out on the pre-treated substrate with the microorganism and this can be followed by fermentation to produce ethanol which is utilised as biofuel. The use of biomass in producing the bioethanol has hope in providing an indigenously produced, renewable energy source which may be viable option for the fuels that are based on petroleum.

Conclusion:-

There are lignocellulosic materials in the plant cells and these lignocellulosic materials are made up of lignin, cellulose and hemicellulose. The process of pre-treatment of the lignocellulosic biomass before the process of hydrolysis distinctly enhances the hydrolysis efficiency because of the lignin and hemicellulose removal and cellulose reduction. The microorganism degrades the cell walls of the plants by an enzyme set, especially fungi. There are two types of extra-cellular enzymatic systems in fungi. These are hydrolytic and ligninolytic systems. Hydrolases are produced by the hydrolytic system and the lignin component is degraded by the ligninolytic system.

The ligninolytic system also opens the phenyl rings. The white-rot fungi (WRF), among other microorganisms, are effective for the pre-treatment done by biological methods. This is due to the reason that the lignin is degraded more rapidly and extensively in comparison to the other related microorganisms. The activities of the lignin-degrading fungi WRF has relatively lower cellulase and xylanase activities. This is advantageous for an efficient process of delignification. This consequently leads to the reduction of energy and chemical inputs for enzymatic or chemical hydrolysis of the related substrate.

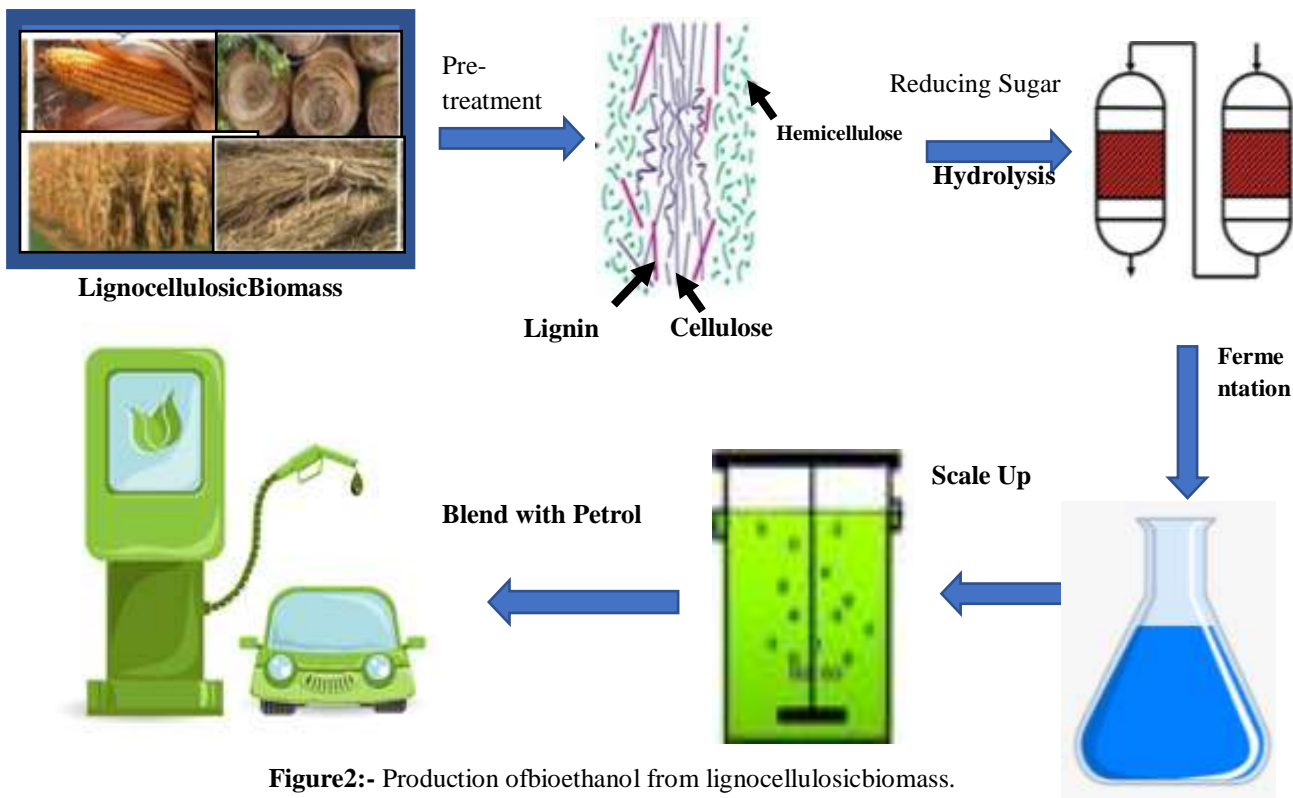


Figure2:- Production of bioethanol from lignocellulosic biomass.

References:-

1. Abbi M, Kuhad RC, Singh A (1996a) Bioconversion of pentose sugars to ethanol by free and immobilized cells of *Candida shehatae* NCL-3501 Fermentation behaviour. *Process Biochem* 31:555-560
2. Abbi M, Kuhad RC, Singh A (1996b) Fermentation of xylose and rice straw hydrolysate to ethanol by *Candida shehatae* NCL-3501. *J Ind Microbiol* 17: 20-23
3. Adney WS, Chou YC, Decker SR, Ding SY, Baker JO, Kunkel G, Vinzant TB, Himmel ME (2003) Heterologous expression of *Trichoderma reesei* 1,4-beta-D-glucan cellobiohydrolase (Cel 7A). In *Applications of Enzymes to Lignocellulosics*. Washington, Amer Chemical Soc 855:403-437.
4. Alper H and Stephanopoulos G (2009) Engineering for biofuels: exploiting innate microbial capacity or importing biosynthetic potential? *Nat Rev Microbiol* 7:715-723.
5. Antizar-Ladislao B and Turron-Gomez JL (2008) Second-generation biofuels and local bioenergy systems. *Biofuel Bioprod Biorefin* 2:455-469.
6. Antoni D, Zverlov VV, Schwarz WH (2007) Biofuels from microbes. *Appl Microbiol Biotechnol* 77: 23-35
7. Asif, M. B., Hai, F. I., Hou, J., Price, W. E., & Nghiem, L. D. (2017). Impact of wastewater derived dissolved interfering compounds on growth, enzymatic activity and trace organic contaminant removal of white rot fungi—a critical review. *Journal of environmental management*, 201, 89-109.
8. Baig, K. S., Wu, J., & Turcotte, G. (2019). Future prospects of delignification pretreatments for the lignocellulosic materials to produce second generation bioethanol. *International Journal of Energy Research*, 43(4), 1411-1427.
9. Balan V, Souza LDC, Chundawat SS, Vismeh R, Jones AD, Dale BE (2008) Mushroom spent straw: a potential substrate for an ethanol-based biorefinery. *J Ind Microbiol Biotechnol* 35: 293-301
10. Branco, R. H., Serafim, L. S., & Xavier, A. M. (2019). Second generation bioethanol production: on the use of pulp and paper industry wastes as feedstock. *Fermentation*, 5(1), 4.

11. Campbell CJ and Laherrere JH (1998) The end of cheap oil. *Science* 3:78-83
12. Campbell, G. M., Mustač, N. Č., Alyassin, M., Gomez, L. D., Simister, R., Flint, J., ...& Westwood, N. J. (2019). Integrated processing of sugarcane bagasse: arabinoxylan extraction integrated with ethanol production. *Biochemical Engineering Journal*, 146, 31-40.
13. Chaturvedi V, Verma P (2013) An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. *Biotech* 3: 415-431.
14. Dagle, R. A., Winkelman, A. D., Ramasamy, K. K., Lebarbier Dagle, V., & Weber, R. S. (2020). Ethanol as a renewable building block for fuels and chemicals. *Industrial & Engineering Chemistry Research*, 59(11), 4843-4853.
15. Gupta R, Sharma KK and Kuhad RC (2009) Separate hydrolysis and fermentation (SHF) of Prosopis juliflora, a woody substrate, for the production of cellulosic ethanol by *Saccharomyces cerevisiae* and *Pichia stipitis*-NCIM 3498. *Bioresour Technol* 100:1214-1220.
16. Elisashvili V, Penninckx M, Kachlishvili E, Asatiani M, Kvesitadze G (2006) Use of *Pleurotus dryinus* for lignocellulolytic enzymes production in submerged fermentation of mandarin peels and tree leaves. *Enzyme Microb Technol* 38:998-1004
17. Eriksson KEL (1993) Lignin biodegradation and practical utilization. *J Biotechnol* 30:149-158
18. Eriksson KL, Blanchette RA, Ander P (1990) Microbial and enzymatic degradation of wood and wood components. Springer, Berlin Heidelberg New York.
19. Fang X, Shen Y, Zhao J, Bao X, Qu Y (2010) Status and prospects of lignocellulosic bioethanol production in China. *Bioresour Technol* 101: 4814-4819
20. Fernandez-Fueyo E, Ruiz-Dueñas FJ, Ferreira P, Floudas D, Hibbett DS (2012) Comparative genomics of *Ceriporiopsis subvermispora* and *Phanerochaete chrysosporium* provide insight into selective ligninolysis. *Proc Natl Acad Sci* 109 :5458–5463.
21. Gnansounou E (2010) Production and use of lignocellulosic bioethanol in Europe: Current situation and perspectives. *Bioresour Technol* 101: 4842–50
22. Goring DAI, Timell TE (1962) Molecular weight of native celluloses. *Tappi* 45:454–460
23. Gupta R, Mehta G, Khasa YP, Kuhad RC (2011b) Fungal delignification of lignocellulosic biomass improves the saccharification of celluloses. *Biodegradation* 22:797-804
24. Hatakka AI (1983) Pretreatment of wheat straw by white-rot fungi for enzymatic saccharification of cellulose. *Appl Microbiol Biot* 18: 350–357
25. Hatakka A (2005) Biodegradation of lignin. *Biopolym Online*. 1.
26. Himmel ME, Ding SY, Johnson DK, Adney WS, Nimlos MR, Brady JW, Foust TD (2007) Biomass recalcitrance: Engineering plants and enzymes for biofuels production. *Science* 315: 804-807
27. IEA (2008) World Energy Outlook
28. Itoh H, Wada M, Honda Y, Kuwahara M, Watanabe T (2003) Bioorganosolve pretreatments for simultaneous saccharification and fermentation of beech wood by ethanolysis and white rot fungi. *J Biotechnol* 103: 273–280
29. Jung, Y. H., & Kim, K. H. (2017). Evaluation of the main inhibitors from lignocellulose pretreatment for enzymatic hydrolysis and yeast fermentation. *BioResources*, 12(4), 9348-9356.
30. Kang S, Li X, Fan J, Chang J (2013) Hydrothermal conversion of lignin: A review. *Renew Sustainable Energy Rev* 27: 546-558.
31. Keller FA, Hamillton TE, Nguyen QA (2003) Microbial pretreatment of biomass potential for reducing severity of thermo-chemical biomass pretreatment. *Appl Biochem Biotech* 105:27-41
32. Kim JS, Park SC, Kim JW, Park JC, Park SM, Lee JS (2010) Production of bioethanol from lignocellulose: Status and perspectives in Korea. *Bioresour Technol* 101: 4801–4805
33. Kim S, Dale EB (2004) Global potential bioethanol production from wasted crops and crop residues. *BIOMASS BIOENERG* 26: 361–375
34. Kocher, G. S., Kaur, P., & Taggar, M. S. (2017). An overview of pretreatment processes with special reference to biological pretreatment for rice straw delignification. *Current Biochemical Engineering*, 4(3), 151-163.
35. Kuhad RC, Singh A (1993) Lignocellulose Biotechnology: Current and future prospects. *Current reviews in Biotechnology* 13:151-172
36. Kuhad RC, Gupta R, Khasa YP (2011a). Bioethanol production from lignocellulosics: an overview. In: *Wealth from waste*. 3rd Edition. Banwari Lal and Priyangshu M. Sharma (Editors). TERI Press, New Delhi, India.
37. Kuhad RC, Kuhar S, Kapoor M, Sharma KK, Singh A (2007) Lignocellulolytic microorganisms, their enzymes and possible biotechnologies based on lignocellulolytic microorganisms and their enzymes pp 3-32 In: *Lignocellulose Biotechnology: Future prospects* (edited by) R. C. Kuhad and A. Singh. IK International, New Delhi.

38. Kuhad RC, Singh A, Eriksson KEL (1997). Microorganisms and enzymes involved in the degradation of plant fiber cell wall. *AdvBiochemEngBiot* 57:47-125
39. Kuhar S, Nair LM, Kuhad RC (2008) Pretreatment of lignocellulosic material with fungi capable of higher lignin degradation and lower carbohydrate degradation improves substrate acid hydrolysis and the eventual conversion to ethanol. *Can J Microbiol* 54:305-13
40. Kumar G, Bakonyi P, Periyasamy S, Kim SH, Nemestóthy N (2015) Lignocellulose biohydrogen: Practical challenges and recent progress. *RenewSustain Energy Rev* 44: 728-737.
41. Lee J (1997) Biological conversion of lignocellulosic biomass to ethanol. *J Biotechnol* 56:1– 24
42. Li SZ, Chan-Halbrendt C (2009) Ethanol production in China: potential and technologies. *ApplEnergy* 4:4–9
43. Licht FO (2005) World Ethanol and Biofuels Report 3:15
44. Lundell TK, Mäkelä MR, Hildén K (2010) Lignin-modifying enzymes in filamentous basidiomycetes: ecological, functional and phylogenetic review. *Basic Microbiol* 50: 1-16
45. Mansora, A. M., Lima, J. S., Anib, F. N., Hashima, H., &Hoa, W. S. (2019). Characteristics of cellulose, hemicellulose and lignin of MD2 pineapple biomass. *Chem. Eng*, 72, 79-84.
46. Matsumoto N, Sano D, Elder M (2009) Biofuel initiatives in Japan: Strategies, policies and future potential. *ApplEnergy* 86:569–576
47. McCarthy JE and Tiemen M (1998) CRS report for congress MTBE in gasoline clean air and drinking water. <http://www.epa.gov/otaq/consumer/fuels/mtbe/MTBE.pdf>.
48. McMillan JD (1994) Pretreatment of lignocellulosic biomass. *AcsSymSer* 566: 292–324
49. Mishra, S., Singh, P. K., Dash, S., & Pattnaik, R. (2018). Microbial pretreatment of lignocellulosic biomass for enhanced biomethanation and waste management. *3 Biotech*, 8(11), 1-12.
50. Morris D (1993) Ethanol: A 150 Year Struggle Toward a Renewable Future. (Washington: Institute for Local Self-Reliance)
51. Mosier NS, Wyman C, Dale B, Elander R, Lee YY, Holtzapple M, Ladisch MR (2005) Features of promising technologies for pretreatment of lignocellulosic biomass. *BioresourceTechnol* 96: 673–686
52. Mosier NS, Wyman C, Dale B, Elander R, Lee YY, Holtzapple M, Ladisch MR (2005) Features of promising technologies for pretreatment of lignocellulosic biomass. *BioresourceTechnol* 96: 673–686.
53. Mussatto SI, Dragone G, Guimarães PMR, Silva JPA, Carneiro LM, Roberto IC, Vicente A, Domingues L, Teixeira JA (2010). Technological trends, global market and challenges of bio-ethanol production R1. *BiotechnolAdv* 28: 817-30
54. Narayanaswamy N, Dheeran P, Verma S, Kumar S (2013) Biological pre-treatment of lignocellulosic biomass for enzymatic saccharification. Fang Z, editor. In *Pre-treatment Techniques for Biofuels and Biorefineries*. Springer, Germany.
55. Orellana C, Bonalume NR (2006) Brazil and Japan give fuel to ethanol market. *Nat Biotechnol* 24: 232
56. Petrova P, Ivanova V (2010) Perspectives for the production of bioethanol from lignocellulosic materials. *BiotechnolBioeng* 24: 529-546
57. Pop, V. (2018). PETROLEUM-STRATEGIC RESOURCE FOR WORLD ECONOMY. *Studia Universitatis Vasile Goldiș, Arad-Seria Științe Economice*, 28(2), 70-85.
58. Prasad S, Singh A, Joshi HC (2007) Ethanol as an alternative fuel from agricultural, industrial and urban residues. *Resources Conservation and Recycling*
59. Prather, R. M., Castillioni, K., Welti, E. A., Kaspari, M., & Souza, L. (2020). Abiotic factors and plant biomass, not plant diversity, strongly shape grassland arthropods under drought conditions.
60. Prieur-Vernat A, His S (2006). *Biofuels in Europe*. Panoram. Paris: IFP
61. Radhika R, Jebapriya GR, Gnanadoss JJ (2013) Production of cellulase and laccase using *Pleurotus* sp. under submerged and solid-state fermentation. *Int J CurrSci* 6E: 7-13
62. Ragauskas AJ, Williams CK, Davison BH, Britovsek G, Cairney J, Eckert CA, Frederick Jr, WJ, Hallett JP, Leak DJ, Liotta CL, Mielenz JR, Murphy R, Templer R and Tschaplinski T (2006) The path forward for biofuels and biomaterials. *Science* 311: 484-489.
63. Rasmussen, S. C. (2019). From Aqua Vitae to E85: The History of Ethanol as Fuel. *Substantia*, 43-55.
64. Rosales E, Rodriguez Couto S, Sanroman MA (2002) A new uses of food wastes: application to laccase production by *Trametes hirsute*. *BiotechnolLett* 24:701-704
65. Rowell RM, Pettersen R, Han JS, Rowell JS, Tshabalala MA (2005) Cell wall chemistry in: handbook of wood chemistry and wood composites (edited by) R M Rowell. Florida 33431, CRC Press LLC pp 35-62.
66. Sanchez C (2009) Lignocellulosic residues: biodegradation and bioconversion by fungi. *BiotechnolAdv* 27: 185–194

67. Sanchez OJ, Cardona CA (2008) Trends in biotechnological production of fuel ethanol from different feedstocks. *Bioresour Technol* 99: 5270-5295
68. Silalertruksa T, Gheewala SH (2009) Environmental sustainability of bio- ethanol production in Thailand. *Energy* 34: 1933-1946
69. Singh, J. K., Vyas, P., Dubey, A., Upadhyaya, C. P., Kothari, R., Tyagi, V. V., & Kumar, A. (2018). Assessment of different pretreatment technologies for efficient bioconversion of lignocellulose to ethanol. *Front. Biosci*, 10(521), 10-2741.
70. Solomon BD, Barnes JR, Ivorsen KE (2007) Grain and cellulosic ethanol: history, economics and energy policy. *Biomass Bioenerg* 31: 416–425
71. Songulashvili G, Elisashvili V, Wasser S, Nevo E, Hadar Y (2006) Laccase and manganese peroxidases activities of *Phellinus robustus* and *Ganoderma adspersum* grown on food industry wastes in submerged fermentation. *Biotechnol. Lett.* 28: 1425-1429
72. Suhara H, Kodama S, Kamei I, Maekawa N, Meguro S (2012) Screening of selective lignin-degrading basidiomycetes and biological pretreatment for enzymatic hydrolysis of bamboo culms. *Int Biodeterior Biodegradation* 75:176–180.
73. Sukumaran RK, Surender VJ, Sindhu R, Binod P, Janu KU, Sajna KV, Rajasree KP, Pandey A (2010) Lignocellulosic ethanol in India: Prospects, challenges and feedstock availability. *Bioresour Technol* 101:4826–4833
74. Taha M, Shahsavari E, Al-Hothaly K, Mouradov A, Smith AT (2015) Enhanced Biological Straw Saccharification Through Coculturing of Lignocellulose-Degrading Microorganisms. *Appl Biochem Biotechnol* 175:3709-3728.
75. Taniguchi M, Suzuki H, Watanabe D, Sakai K, Hoshino K and Tanaka T (2005) Evaluation of pretreatment with *Pleurotus ostreatus* for enzymatic hydrolysis of rice straw. *J Biosci Bioeng* 100:637-643
76. Wang W, Yuan T, Cui B, Dai Y (2013) Investigating lignin and hemicellulose in white rot fungus-pretreated wood that affect enzymatic hydrolysis. *Bioresour Technol* 134: 381-385.
77. Yu, J., Paterson, N., Blamey, J., & Millan, M. (2017). Cellulose, xylan and lignin interactions during pyrolysis of lignocellulosic biomass. *Fuel*, 191, 140-149.
78. Zhang YHP, Himmel ME, Mielenz JR (2006) Outlook for cellulase improvement: Screening and selection strategies. *Biotechnol Adv* 24: 452-481.