

Journal Homepage: -www.journalijar.com

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH (IJAR)

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH (IJAR)
1003 2505-5607
Justical hamogogy: Inquivos justicalija com
Justical (10) 31, 217-111-181

Article DOI:10.21474/IJAR01/19908 **DOI URL:** http://dx.doi.org/10.21474/IJAR01/19908

RESEARCH ARTICLE

Effect of Arbuscular Mycorrhizal Fungi on Salt Stress: A Review

Bhosale K.S.¹, Mankar G.D.¹ and Kenjale P.A.²

- 1. Department of Botany, Nowrosjee Wadia College, Pune 411 001, Maharashtra, India.
- 2. Department of Botany, Tuljaram Chaturchand College of Arts, Commerce and Science, Baramati 413102 Maharashtra, India.

.....

Manuscript Info

Manuscript History

Received: 11 September 2024 Final Accepted: 21 October 2024 Published: November 2024

Key words:-

Arbuscular Mycorrhizal Fungi, Microflora, Salt Stress, Sustainable, Symbiotic

Abstract

Biodiversity is essential to sustaining the ecosystem and preserving ecological balance. However, the growing risk of salt stress brought on by extensive irrigation and chemical fertilizer use has made biodiversity protection extremely difficult and lowers crop productivity. Arbuscular Mycorrhizal (AM) fungi are symbiotic soil microorganisms, plays the important role in relieving the argumentative effects of salt stress in plants. This comprehensive review produces current knowledge related to the complex association between AM fungi and salinity in terrestrial ecosystems. It investigates how AM fungi improve plant's resistance to salt stress. Furthermore, the review investigates the remarkable biodiversity of AM fungi from different plant associations worldwide, emphasizing the significance of choosing appropriate AM fungi species aimed ateffective ecosystem restoration and agricultural practices in saline environments. This research offers important insights into tying AM fungi's capacity to counteract saltinduced biodiversity loss and boost agricultural yield in saline soils by elucidating the intricate relationships between AM fungi and salt stress. This review highlights the need for further research to unravel the complexities of this symbiotic relationship and develop sustainable strategies for countering the problems of salt stress and crop productivity.

.....

Copyright, IJAR, 2024,. All rights reserved.

Introduction:-

Soil microflora includes diverse bacteria and fungi, some pathogenic and others nonpathogenic. Among the nonpathogenic microorganisms, AM fungi play a crucial role in making symbiotic relationships with plant roots (St Arnaud andElsen 2005). According to Gupta et al. (2021), these fungi are crucial for enhancing plant growth and resistance to environmental stress, especially salt stress. The rapid increase of soil salinity due to improper agricultural practices and Overuse of chemical fertilisers has caused soil fertility to decline, which poses problems for crop productivity and biodiversity.

There are many spores of AM fungi found in soil, out of them few species show mutualistic interaction with crops. Mutualistic relation is of two types; ectomycorrhiza's and endomycorrhiza's. Ectomycorrhiza is not associated with plant roots and endomycorrhiza is associated with plant roots by forming and penetrating mycelium between cells

968

and inside the cell (Roy and Choudhury 2022). The intricate relationships between AM fungus and plants under salt stress, are highlighted in this paper, along with how AM fungi can be used to mitigate the negative effects of salt stress on plant growth.

Effect of Salt Stress on Plants

Nowadays soil degradation remains a major issue in the field of agriculture and it is increasing steadily all over the world (Lanfranco, L. et al., 2016). This problem is because of improper irrigation, and excess use of chemical fertilizers (Savci, S. 2012). It causes loss in the microflora of soil and soil fertility; increasing salinity stress. Increased abiotic stress like water stress, salt stress, abrupt changes in temperature basically the mainly due to changes in climatic conditions. Such kinds of stressed conditions affected organic contents of soil. Soil contains active microorganisms of different kinds assisting plant growth in various ways. The soil from rhizosphere consist of more than a trillion species present out of them microbiota and microflora are more dominant in this layer of soil (Sharma et al., 2021). From the fungal group; AM fungi are among the most significant and economically significant organisms in the fungus group (Smith and Smith 2011).

One of the main abiotic stress which affecting plant growth and productivity is salinity. Increased salt levels of soil disrupt plant water absorption, cause ionic imbalances, and reduce photosynthetic efficiency. The germination of seeds, plant growth, and total crop output are all adversely affected by the high concentration of salt in the soil. Salinity reduces chlorophyll content of crops, resulting in decreased photosynthesis and productivity (Mbarki et al., 2018). Different salts alter the soil's physicochemical characteristics (Li et al., 2019). It also alters water potential of soil, affecting the accessibility of water to plants (Rabie and Almadini, 2005). Salinity of soil is a biggest hazard towards security of food due to lower production. Nearabout 7% of the soil badly affected due to deposition of salts (Ruiz Lozano et al., 2012). Salt contents of soil reduces germination capacity of seed thereby affecting its produce (Ouni et al., 2014). Additionally, it has effect on exhaustion of water in the rhizosphere and reduces capacity to hold water (Sabir et al., 2009). Salt content of soil obstructs plant's capacity to absorb water and disrupt ionic homeostasis (Chakraborty et al., 2018). Furthermore, in stresses various cellular components damage due to reactive oxygen species (Velarde-Buendia AM., 2012). ROS negatively correlats to the plant growth (Ahmad P., 2010). Phosphorous content inversely proportional to salt concentration of soil. Salinity has affected nearabout 25 million acres of soil (Latef and Chaoxing., 2011). If not addressed, the rising salinity in soil could lead to a 30% loss of cultivable land within the next few decades (Porcel et al., 2012).

Effect of AM Fungi on Salt Stress

Applications of non-hazardous organic techniques, such as the use of AM fungus, can mitigate the negative consequences of salt stress (Badda N. et al., 2014). AM fungi are one of the microorganisms that are most well-known for lowering different types of stress. Indeed, AM Fungi potential is being continuously tested for different stresses (Ahangeret al., 2014, Bencherif et al., 2015). By improving the uptake of water and trace elements from the soil, AM fungus can help plants become more resistant to salinity stress (Evelin et al., 2009). Soil microflora with AM fungi and the biofertilizers are known globally to enhance crop yields and protect plants from various stresses. As decomposers, biological controllers, and ecosystem regulators, fungi can serve a variety of purposes in the environment. AM fungi are leading organic amendments to boost the plant growth and guard them from various fungi (Frac et al., 2018). AM fungi assist the associated plant to protect from various pathogens (Lehmann et al., 2017).

AM fungus has been shown to aid in plant's recovery from the negative consequences of salt stress. According to a number of research, plants associated with AM fungus flourish and produce more biomass in saline conditions (Bhosale and Shinde, 2011). This fungi increase nutrient uptake, control osmotic equilibrium, and lessen oxidative stress, which increases plant tolerance to salinity. Plant growth and production are increased by AM fungi, which are known to increase chlorophyll content and photosynthetic efficiency even in the face of salt stress (Majeed & Muhammad 2019). Additionally, AM fungi help to maintain the overall health of the plant by supplying essential nutrients, such as phosphorus, which is required for plant growth under stressful situations (Begum et al., 2019). Association of AM fungi enriched dry biomass of seedlings (Latef & Chaoxing., 2011).

Soybean can undergo a symbiotic interaction with soil microorganisms like rhizobium and AM fungi to fix atmospheric nitrogen as well as to supply various micronutrients. Nitrogen along with phosphorus are ulmost important amendments for growth of plants (Marschner, 1995). We use more synthetic chemical fertilizers to increase the yield of crops. Such use of synthetic fertilizers results in lethal issues related to the environment. Only

half of the additional N and around a quarter of the P fertiliser can be absorbed by the plants (Adesemoye and Kloepper, 2009, Garnett, 2009). Soybean shows association of Rhizobium as well as AM fungi simultaneously, which are beneficial both Phosphorous along with the Nitrogen (Meena et al., 2018). Both water and nutrients from the soil are captured by the hyphae of AM fungus. The symbiosis between AM fungus and plants is especially crucial for plants that grow in soils with low P availability. (Ibrahim, 2011; Abdul-Fattah, 1997). According to Ghorchiani et al. (2018), Abdel-Fattah and Asrar (2012), Sheng et al. (2013), and Anand et al. (2022), it accelerated the rate of plant development and raised the P concentration in plant tissues. AM fungi shows great biodiversity in the association with different plants as shown in following table, which helps plant to alleviate different stresses and enhance growth and development in crop.

Table 1:- 'Biodiversity of arbuscular mycorrhiza fungi from different plant association worldwide'.

| Sr. | | | |
|-----|---------------------------|------------------------|---------------------------------|
| No. | AMF | Plant association | References |
| | Acaulospora denticulate | | |
| 1 | Entrophospora kentinensis | Date palm | Sghir et al., 2014. |
| 2 | Acaulospora spinosa | Tea | Singh et al., 2008. |
| 3 | Claroideoglomus hanlinii | Plantago lanceolata | Błaszkowski et al., 2015. |
| | Diversispora aurantia | | |
| | Funneliformis coronatus | | Estrada et al., 2013. |
| | Paraglomus occultum | | |
| 4 | Scutellospora calospora | Asteriscus maritimus | |
| 5 | Diversispora clara | Asteriscus maritimus | Estrada et al., 2012. |
| 6 | Diversispor acelata | Glycine max | Gamper et al., 2009. |
| | Entrophospora colombiana | | |
| | Glomus hoi | | |
| 7 | Glomus tenebrosum | fruit orchards | Ragupathy and Mahadevan., 1993. |
| | Gigaspora calospora | | |
| | Sclerocystis coremioides | | |
| 8 | Scutellospora pellucida | Litchi | Sharma et al., 2009. |
| 9 | Gigaspora margarita | Puspalum nototum | Douds and Schenck, 1991. |
| 10 | Otospora bareai | Thymus granatensis | Palenzuela et al., 2008. |
| 11 | Paraglomus majewskii | Plantago lanceolata | Błaszkowski et al., 2012. |
| 12 | Rhizoglomus melanum | Isoetes lacustris | Sudovaet al., 2015. |
| 13 | Sclerocystis sinuosa | Rhododendron lepidotum | Chaurasia et al., 2005. |
| 14 | Scutellispora nigra | Rhododendron barbatum | Mbogne et al., 2015. |

Mechanisms by Which AM Fungi Alleviate Salt Stress Enhanced Nutrient Uptake

Numerous research investigating the role of AMF in protection against salt stress have found that the symbiosis often results in increased nutrient intake, buildup of osmoregulatory chemicals, an increase in photosynthetic rate, and an increase in water usage efficacy. These results imply that AMF's capacity to reduce salt stress is a result of a confluence of molecular, physiological, biochemical, and dietary impacts. Accordingly, it was found that mycorrhization increased the host plant's fitness by promoting its development (Porcel et al., 2012). Several researchers claim that plants infected with AM fungus grow more effectively than those that are not under salt stress (Al-Karaki 2000; Cantrell and Linderman 2001). By boosting the uptake of phosphorus and other immobile nutrients from the soil, AM fungi considerably improve the nutritional status of plants. An essential component of plant growth, phosphorus is frequently scarce in saline soils. Crop development and stress tolerance are improved by AM fungi because they boost phosphorus uptake (Begum et al., 2019, Abdel-Fattah, 1997). Furthermore, plants and AM fungus have a symbiotic interaction that makes it easier for the plants to absorb water and trace elements that are essential for their survival in salt stress (Evelin et al., 2009).

Osmotic Regulation

Plants experience water stress when salinity upsets their osmotic balance. By controlling osmotic balance and enhancing water intake, AM fungus assist plants in maintaining water relations. AM fungus hyphae help plants adapt to saltwater environments by penetrating the soil and aiding in water absorption (Mayak et al., 2004). One of

the most serious outcomes is the loss of intracellular water due to salinity stress. By stopping water loss from the cytoplasm of the cell, compatible organic solutes that are held in the cytoplasm of plant cells help to promote plant development during salt stress. Several chemicals have been postulated to perform this function in halophytes, including proline, betaines, sugars (primarily fructose, sucrose, and glucose), and complex sugars (trehalose, raffinose, and fructans) (Porcel et al., 2012).

The phytohormone ABA controls plant growth and development and has a major effect on how plants respond to abiotic stresses such as salt stress. Mycorrhizal connection with plant roots has been shown to change the ABA levels in the host plant (Duan et al., 1996).

Reduction of Oxidative Stress

Many of the degenerative reactions connected to different biotic, abiotic, and xenobiotic stresses are mediated by Reactive Oxygen Species (ROS) (Porcel et al., 2012). Superoxide, hydroxyl radicals, hydrogen peroxide, and singlet oxygen are all included in the term ROS. ROS, which can harm plant cells, are frequently produced as a result of salt stress. AM fungi reduce the oxidative stress by modulating ROS levels and improving the antioxidant responses in plants. Because it lessens cellular damage and enhances general plant health, this defence system is essential for plant survival under salt stress (Ahmad, 2010; Velarde-Buendia et al., 2012). Numerous research have shown that AM symbiosis increases the activities of antioxidant enzymes, which helps plants deal with salt stress (Zhong et al., 2007).

Improved Photosynthesis

The loss of chlorophyll content is the main cause of the decrease in photosynthetic efficiency, which is one of the major effects of salinity. When plants are under salt stress, AM fungus increase their chlorophyll concentration and photosynthetic activity. Enhanced photosynthesis leads to better growth and biomass production, even in saline soils (Kong et al., 2020). By improving the plant's ability to capture carbon and utilize it for growth, AM fungi ensure that the plant maintains productivity under stress conditions. Salt stress causes physiological dryness and photosynthetic inhibition, which lower crop output (Pitman and Lauchli 2002). However, relatively little study has been done on how AM fungus inoculation affects photosynthesis, specifically the photochemical properties of leaves under salt stress (Sheng et al., 2008).

Summary:-

Research on the impact of AM fungi under salt stress has shed light on the possible advantages of these symbiotic partnerships in reducing the negative effects of salt on plants. By improving nutrient uptake, mycorrhizal fungi have been shown to dramatically increase a plant's tolerance to salt stress. Furthermore, under salinised conditions, AM fungi have demonstrated potential in sustaining plant growth, biomass output, and general plant health.

However, it's important to remember that the effectiveness of mycorrhizal supplements in reducing salt stress might differ based on a number of variables, including the type of plant or fungus and the particular environmental circumstances. To maximise mycorrhiza's agricultural use and comprehend the mechanisms underlying their role in salt stress resistance, more research is required.

In conclusion, research on the impact of AM fungus on salt stress demonstrates the potential of these helpful fungi, which play a crucial role in ecosystem management and sustainable agriculture, especially in saline conditions. Connecting the advantages of AM fungus offers a promising way to increase crop productivity and environmental resilience as we continue to deal with salinity in a changing climate.

Acknowledgements:-

With profound appreciation, we would like to thank everyone who helped us finish this study on the effect of Arbuscular Mycorrhizal (AM) fungi on salt stress. Our work would not have been possible without the support, guidance, and assistance of the Department of botany of T. C. College Baramati. Department of biotechnology of NowrosjeeWadia College Pune. I also would like to thank SARTHI Pune, for their financial support through CSMNRF to Mr. Kenjale Prasad Anil.

Future Research Directions:-

The specificity of AM fungal-plant interactions in saline environments needs to be investigated. Examining the physiological and molecular processes that underlie the salt tolerance mediated by AM fungus. Formulating plans for the extensive use of AM fungus in saline farming. The diversity of AM fungus in saline salt must be understood.

References:-

- 1. Abdel-Fattah, G. M. (1997). Functional activity of VA-mycorrhiza (*Glomus mosseae*) in the growth and productivity of soybean plants grown in sterilized soil. Folia Microbiologica, 42, 495-502.
- 2. Abdel-Fattah, G. M., &Asrar, A. W. A. (2012). Arbuscular mycorrhizal fungal application to improve growth and tolerance of wheat (*Triticum aestivum* L.) plants grown in saline soil. Acta Physiologiae Plantarum, 34, 267-277.
- 3. Adesemoye, A. O., & Kloepper, J. W. (2009). Plant–microbes interactions in enhanced fertilizer-use efficiency. Applied microbiology and biotechnology, 85, 1-12.
- 4. Ahanger, M. A., Hashem, A., Abd-Allah, E. F., & Ahmad, P. (2014). Arbuscular mycorrhiza in crop improvement under environmental stress. In Emerging technologies and management of crop stress tolerance (pp. 69-95). Academic Press.
- 5. Ahmad, P. (2010). Growth and antioxidant responses in mustard (*Brassica juncea* L.) plants subjected to combined effect of gibberellic acid and salinity. Archives of Agronomy and Soil Science, 56(5), 575-588.
- Al-Karaki, G. N. (2000). Growth of mycorrhizal tomato and mineral acquisition under salt stress. Mycorrhiza, 10, 51-54.
- 7. Anand, K., Pandey, G. K., Kaur, T., Pericak, O., Olson, C., Mohan, R., & Yadav, A. N. (2022). Arbuscular mycorrhizal fungi as a potential biofertilizers for agricultural sustainability. Journal of Applied Biology and Biotechnology, 10(1), 90-107.
- 8. Badda, N., Aggarwal, A., Kadian, N., & Sharma, N. (2014). Influence of arbuscular mycorrhizal fungi and different salinity levels on growth enhancement and nutrient uptake of *Gossypium arboretum* L. Kavaka, 43, 14-21.
- 9. Begum, N., Qin, C., Ahanger, M. A., Raza, S., Khan, M. I., Ashraf, M., & Zhang, L. (2019). Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance. Frontiers in plant science, 10, 1068
- 10. Bencherif, K., Boutekrabt, A., Fontaine, J., Laruelle, F., Dalpe, Y., & Sahraoui, A. L. H. (2015). Impact of soil salinity on arbuscular mycorrhizal fungi biodiversity and microflora biomass associated with *Tamarix articulate* Vahll rhizosphere in arid and semi-arid Algerian areas. Science of the Total Environment, 533, 488-494.
- 11. Bhosale, K. S., & Shinde, B. P., (2011). Influence of arbuscularmycorrhizal fungi on proline and chlorophyll content in *Zingiber officinale*Rosc grown under water stress. Indian Journal of Fundamental and Applied Life Sciences, 1(3), 172-176.
- 12. Błaszkowski, J., Furrazola, E., Chwat, G., Góralska, A., Lukács, A. F., &Kovács, G. M. (2015). Three new arbuscular mycorrhizal *Diversispora* species in *Glomeromycota*. Mycological Progress, 14, 1-12.
- 13. Błaszkowski, J., Kovács, G. M., Gáspár, B. K., Balázs, T. K., Buscot, F., & Ryszka, P. (2012). The arbuscular mycorrhizal *Paraglomus majewskii* sp. nov.represents a distinct basal lineage in *Glomeromycota*. Mycologia, 104(1), 148-156.
- 14. Cantrell IC, Linderman RG. (2001). Preinoculation of lettuce and onion with VA mycorrhizal fungi reduces deleterious effects of soil salinity. Plant Soil 233:269–281.
- 15. Chakraborty, K., Basak, N., Bhaduri, D., Ray, S., Vijayan, J., Chattopadhyay, K., & Sarkar, R. K. (2018). Ionic basis of salt tolerance in plants: nutrient homeostasis and oxidative stress tolerance. Plant nutrients and abiotic stress tolerance, 325-362.
- Chaurasia, B., Pandey, A., &Palni, L. M. S. (2005). Distribution, colonization and diversity of arbuscular mycorrhizal fungi associated with central Himalayan rhododendrons. Forest Ecology and Management, 207(3), 315-324.
- 17. Douds Jr, D. D., & Schenck, N. C. (1991). Germination and hyphal growth of VAM fungi during and after storage in soil at five matric potentials. Soil Biology and Biochemistry, 23(2), 177-183.
- 18. Duan XG, Neuman DS, Reiber JM, Green CD, Saxton AM, Augé RM, (1996). Mycorrhizal influence on hydraulic and hormonal factors implicated in the control of stomatal conductance during drought. J Exp Bot 47:1541–1550.

- 19. Estrada, B., Beltrán-Hermoso, M., Palenzuela, J., Iwase, K., Ruiz-Lozano, J. M., Barea, J. M., & Oehl, F. (2013). Diversity of arbuscular mycorrhizal fungi in the rhizosphere of *Asteriscus maritimus* (L.) Less., a representative plant species in arid and saline Mediterranean ecosystems. Journal of arid environments, 97, 170-175.
- 20. Estrada, B., Palenzuela, J., Barea, J. M., Manuel Ruiz-Lozano, J., Alves da Silva, G., & Oehl, F. (2012). *Diversispora clara* (Glomeromycetes)—a new species from saline dunes in the Natural Park Cabo de Gata (Spain). Mycotaxon, 118(1), 73-81.
- 21. Evelin, H., Kapoor, R., &Giri, B. (2009). Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. Annals of botany, 104(7), 1263-1280.
- 22. Frąc, M., Hannula, S. E., Bełka, M., & Jędryczka, M. (2018). Fungal biodiversity and their role in soil health. Frontiers in microbiology, 9, 707.
- 23. Gamper, H. A., Walker, C., & Schüßler, A. (2009). *Diversispora celata* sp. nov: molecular ecology and phylotaxonomy of an inconspicuous arbuscular mycorrhizal fungus. New Phytologist, 182(2), 495-506.
- 24. Garnett, T., Conn, V., & Kaiser, B. N. (2009). Root based approaches to improving nitrogen use efficiency in plants. Plant, cell & environment, 32(9), 1272-1283.
- 25. Ghorchiani, M., Etesami, H., &Alikhani, H. A. (2018). Improvement of growth and yield of maize under water stress by co-inoculating an arbuscular mycorrhizal fungus and a plant growth promoting rhizobacterium together with phosphate fertilizers. Agriculture, Ecosystems & Environment, 258, 59-70.
- 26. Gupta, S., Schillaci, M., Walker, R., Smith, P. M., Watt, M., & Roessner, U. (2021). Alleviation of salinity stress in plants by endophytic plant-fungal symbiosis: Current knowledge, perspectives and future directions. Plant and Soil, 461, 219-244.
- 27. Horst Marschner. (1995). Mineral nutrition of higher plants. Academic press.
- 28. Ibrahim, A. H., Abdel-Fattah, G. M., Eman, F. M., Abd El-Aziz, M. H., & Shohr, A. E. (2011). Arbuscular mycorrhizal fungi and spermine alleviate the adverse effects of salinity stress on electrolyte leakage and productivity of wheat plants. Phyton, 51(2), 261-76.
- 29. Kong, L., Gong, X., Zhang, X., Zhang, W., Sun, J., & Chen, B. (2020). Effects of arbuscular mycorrhizal fungi on photosynthesis, ion balance of tomato plants under saline-alkali soil condition. Journal of Plant Nutrition, 43(5), 682-698.
- 30. Lanfranco, L., Bonfante, P., & Genre, A. (2016). The mutualistic interaction between plants and arbuscular mycorrhizal fungi. Microbiology spectrum, 4(6), 10-1128.
- 31. Latef, A. A. H. A., & Chaoxing, H. (2011). Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. Scientia Horticulturae, 127(3), 228-233.
- 32. Lehmann, A., Leifheit, E. F., & Rillig, M. C. (2017). Mycorrhizas and soil aggregation. In Mycorrhizal mediation of soil (pp. 241-262). Elsevier.
- 33. Li, X., Ren, J., Zhao, K., & Liang, Z. (2019). Correlation between spectral characteristics and physicochemical parameters of soda-saline soils in different states. Remote Sensing, 11(4), 388.
- 34. Majeed, A., & Muhammad, Z. (2019). Salinity: a major agricultural problem causes, impacts on crop productivity and management strategies. Plant abiotic stress tolerance: Agronomic, molecular and biotechnological approaches, 83-99.
- 35. Mayak, S., Tirosh, T., & Glick, B. R. (2004). Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. Plant physiology and Biochemistry, 42(6), 565-572.
- 36. Mbarki, S., Sytar, O., Cerda, A., Zivcak, M., Rastogi, A., He, X., & Brestic, M. (2018). Strategies to mitigate the salt stress effects on photosynthetic apparatus and productivity of crop plants. Salinity Responses and Tolerance in Plants, Volume 1: Targeting Sensory, Transport and Signaling Mechanisms, 85-136.
- 37. Mbogne, J. T., Temegne, C. N., Hougnandan, P., Youmbi, E., Tonfack, L. B., & Ntsomboh-Ntsefong, G. (2015). Biodiversity of arbuscular mycorrhizal fungi of pumpkins (*Cucurbita* spp.) under the influence of fertilizers in ferralitic soils of Cameroon and Benin. Journal of Applied Biology and Biotechnology, 3(5), 001-010.
- 38. Meena, R. S., Vijayakumar, V., Yadav, G. S., &Mitran, T. (2018). Response and interaction of *Bradyrhizobium japonicum* and arbuscularmycorrhizal fungi in the soybean rhizosphere. Plant Growth Regulation, 84, 207-223.
- 39. Ouni, Y., Ghnaya, T., Montemurro, F., Abdelly, C., & Lakhdar, A. (2014). The role of humic substances in mitigating the harmful effects of soil salinity and improve plant productivity. International Journal of Plant Production, 8(3), 353-374.

- 40. Palenzuela, J., Ferrol, N., Boller, T., Azcón-Aguilar, C., & Oehl, F. (2008). *Otospora bareai*, a new fungal species in the Glomeromycetes from a dolomitic shrub land in Sierra de Baza National Park (Granada, Spain). Mycologia, 100(2), 296-305.
- 41. Pitman M, Läuchli A. (2002). Global impact of salinity and agricultural ecosystems. In: Läuchli A, Lüttge U (eds) Salinity: environment–plants–molecules. Springer, Netherlands, pp 3–20
- 42. Porcel, R., Aroca, R., & Ruiz-Lozano, J. M. (2012). Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. Agronomy for sustainable development, 32, 181-200.
- 43. Rabie, G. H., & Almadini, A. M. (2005). Role of bioinoculants in development of salt-tolerance of *Vicia faba* plants under salinity stress. African Journal of biotechnology, 4(3), 210.
- 44. Ragupathy, S., & Mahadevan, A. (1993). Distribution of vesicular-arbuscular mycorrhizae in the plants and rhizosphere soils of the tropical plains, Tamil Nadu, India. Mycorrhiza, 3, 123-136.
- 45. Roy, S., & Choudhury, L. (2022). The Symbiotic Relationship between Fungi and Plants. In Sustainable Utilization of Fungi in Agriculture and Industry (pp. 52-74). Bentham Science Publishers.
- 46. Ruiz-Lozano, J. M., Porcel, R., Azcon, C., & Aroca, R. (2012). Regulation by arbuscular mycorrhizae of the integrated physiological response to salinity in plants: new challenges in physiological and molecular studies. Journal of Experimental Botany, 63(11), 4033-4044.
- 47. Sabir, P., Ashraf, M., Hussain, M., & Jamil, A. (2009). Relationship of photosynthetic pigments and water relations with salt tolerance of proso millet (*Panicum miliaceum* L.) accessions. Pak. J. Bot, 41(6), 2957-2964.
- 48. Savci, S. 2012. An agricultural pollutant: chemical fertilizer. International Journal of Environmental Science and Development, 3(1), 73.
- 49. Sghir, F., Touati, J., Chliyeh, M., Ouazzani Touhami, A., Filali-Maltouf, A., El Modafar, C.&Douira, A.(2014). Diversity of arbuscular mycorrhizal fungi in the rhizosphere of date palm tree (*Phoenix dactylifera*) in Tafilalt and Zagora regions (Morocco). Int. J. Pure App. Biosci, 2(6), 1-11.
- 50. Sharma, P., Sharma, M. M. M., Malik, A., Vashisth, M., Singh, D., Kumar, R., & Pandey, V. (2021). Rhizosphere, rhizosphere biology, and rhizospheric engineering. In Plant growth-promoting microbes for sustainable biotic and abiotic stress management (pp. 577-624). Cham: Springer International Publishing.
- 51. Sharma, S. D., Kumar, P., Raj, H., & Bhardwaj, S. K. (2009). Isolation of arbuscular mycorrhizal fungi and *Azotobacter chroococcum* from local litchi orchards and evaluation of their activity in the air-layers system. Scientia horticulturae, 123(1), 117-123.
- 52. Sheng M, Tang M, Chen H, Yang BW, Zhang FF, Huang YH. (2008). Influence of arbuscular mycorrhizae on photosynthesis and waterstatus of maize plants under salt stress. Mycorrhiza 18:287–296.doi:10.1007/s00572-008-0180-7
- 53. Sheng, M., Lalande, R., Hamel, C., & Ziadi, N. (2013). Effect of long-term tillage and mineral phosphorus fertilization on arbuscular mycorrhizal fungi in a humid continental zone of Eastern Canada. Plant and Soil, 369, 599-613.
- 54. Singh, S., Pandey, A., &Palni, L. M. S. (2008). Screening of arbuscular mycorrhizal fungal consortia developed from the rhizospheres of natural and cultivated tea plants for growth promotion in tea [*Camellia sinensis* (L.) O. Kuntze]. Pedobiologia, 52(2), 119-125.
- 55. Smith, F. A., & Smith, S. E. (2011). What is the significance of the arbuscular mycorrhizal colonisation of many economically important crop plants? Plant and Soil, 348, 63-79.
- 56. St Arnaud, M., &Elsen, A. (2005). Interaction of arbuscular mycorrhizal fungi with soil-borne pathogens and non-pathogenic rhizosphere micro-organisms. In In vitro culture of mycorrhizas (pp. 217-231). Berlin, Heidelberg: Springer Berlin Heidelberg.
- 57. Sudová, R., Sýkorová, Z., Rydlová, J., Čtvrtlíková, M., &Oehl, F. (2015). *Rhizoglomus melanum*, a new arbuscular mycorrhizal fungal species associated with submerged plants in freshwater lakeAvsjøen in Norway. Mycological Progress, 14, 1-8.
- 58. Velarde-Buendía, A. M., Shabala, S., Cvikrova, M., Dobrovinskaya, O., & Pottosin, I. (2012). Salt-sensitive and salt-tolerant barley varieties differ in the extent of potentiation of the ROS-induced K⁺ efflux by polyamines. Plant Physiology and Biochemistry, 61, 18-23.
- 59. Zhong QH, Chao XH, Zhi BZ, Zhi RZ, Huai SW. (2007). Changes in antioxidative enzymes and cell membrane osmosis in tomato colonized by arbuscular mycorrhizae under NaCl stress. Colloids Surf B Biointerfaces 59:128–133. doi: 10.1016/j.colsurfb.2007.04.023.