

Journal Homepage: -www.journalijar.com

INTERNATIONAL JOURNAL OF

INTERNATIONAL ADCRINAL OF ABILINCED BESEARCH GLAR)

GRANINGED BESEARCH GLARI

GRANINGED BESEARCH

Article DOI: 10.21474/IJAR01/20713 **DOI URL:** http://dx.doi.org/10.21474/IJAR01/20713

RESEARCH ARTICLE

CONTRIBUTION TO THE STUDY OF THE EFFECTS OF VARIOUS ADDITIVES OF ANIMAL ORIGIN ON THE GROWTH OF HYBRID CLARIAS FRY (MALE CLARIAS GARIEPINUS X FEMALE CLARIAS ANGUILLARIS)

NdeyeCodouMbaye, Jean Fall, Mbaye B.A and AllaNdione

Institute of Fishery and Aquaculture, University Cheikh Anta DIOP of Dakar, BP:5005.

Manuscript Info

Manuscript History

Received: 09 February 2025 Final Accepted: 13 March 2025

Published: April 2025

Kev words:-

Cow Blood Meal, Chicken Viscera, Black Soldier Fly Larvae, Growth, Feed Efficiency, Cost

Abstract

Aquaculture in Senegal faces constraints such as the high cost of fish feed, representing a brake on the sectors development. In this study, alternative local meals cow blood meal chicken viscera meal and black soldier fly larvae meal were used as feed additives for hybrid fry of Clariasanguillarisand Clariasgariepinus. Four experimental diets R1, R2, R3 and R4 were formulated, in addition to an industrial control diet (R0), to assess their effects on growth, feed efficiency and production cost. 300 fry of initial mean weight of 1.56 g were distributed in 15 tanks of 50 liters. After 45 days of experimentation, the best zootechnical performance was obtained with diet R4, containing the combination of the three additives. This diet showed a relative mean weight gain (RMWG) of 521.58% and a feed conversion ratio (FCR) of 1.44, similar to the control diet. Economically, the R4 diet had the lowest feed cost of 666.41 FCFA/kg and fish cost of 958.94 FCFA/kg, while maintaining a high survival rate (95%). These results demonstrate that the use of local ingredients as additives is a sustainable and costeffective alternative for improving the productivity and competitiveness of aquaculture in Senegal.

"© 2025 by the Author(s). Published by IJAR under CC BY 4.0. Unrestricted use allowed with credit to the author."

Introduction:-

Fishery plays a considerable socio-economic role in Senegal. However, given the difficulties associated with effective fisheries management and the growing demand for fisheries resources, Senegal has been faced for several decades with the effects of overfishing of its ecosystems, climate change and the collapse of fisheries, impacting local consumption of fisheries production, food security and economic growth (Balde, 2019). Faced with this situation, the Senegalese government has been committed since 2007 to promote aquaculture as an alternative way of coping with dwindling fisheries resources (Ndiaye, 2019).

Aquaculture plays a crucial role in food security and economic development, particularly in developing countries. However, one of the major obstacles to its expansion remains the high cost of fish feed, representing between 40% and 60% of total production costs (Siddhuraju and Becker, 2003). This dependence on imported feed limits farm profitability and hinders the development of high-potential species such as Clariasgariepinusand Clariasanguillaris,

renowned for their rapid growth and adaptability. In this context, the use of local by-products to formulate less costly diets represents a sustainable and economically viable alternative.

In Senegal, products such as chicken viscera, cow blood and black soldier fly larvae are often under-exploited. Slaughterhouses discharge their effluents, including blood, into drainage channels without pre-treatment, contributing to significant environmental pollution. These by-products, which are rich in essential nutrients, nonetheless offer potential for valorization. Blood, for example, contains high-quality proteins, notably albumin and hemoglobin, and its recovery reduces the biochemical oxygen demand (BOD) of wastewater, thus limiting the degradation of aquatic ecosystems (WHO, 2023). Chicken viscera, also rich in proteins, fats and amino acids, are often regarded as waste, although they can be processed into meal for animal feed. Black soldier fly larvae, on the other hand, are rich in protein and essential fatty acids, and have the advantage of being produced from recycled organic matter, making them an ecological and sustainable source.

The main objective of this study was to evaluate the effects of these ingredients, used as feed additives, on the growth of catfish hybrid fry of male Clariasgariepinus and female Clariasanguillaris. More specifically, it aims to:

- 1. add value to cow blood, broiler viscera and black soldier fly larvae by incorporating them as additives in Clarias hybrid feeds;
- 2. determine the zootechnical performance of Clarias hybrids;
- 3. determine the nutritional contribution of foodstuffs on the flesh of Clarias hybrids;
- 4. determine variations in rearing water temperature and pH during experimentation;
- 5. determine the production cost of feed and fish.

Material and Methods

Methods

Processing of some ingredients used in this study

Various methods were used in this study. These included processing of raw materials into finished products, feed formulation and manufacture, rearing conditions and determination of zootechnical parameters of Clarias hybrid fry.

Blood meal production

Fresh blood obtained from the SOGAS slaughter house in Dakar was placed in a 20L plastic container and transported to the IUPA aquaculture station. To sterilize the fresh blood, it was heat-treated by boiling at 120 degrees Celsius for a quarter of an hour, then sun-dried for 48 hours and finely ground for use in diets. The blood meal was stored in glass jars until use.

Black soldier fly meal production

Black soldier fly larvae were harvested when they reached an optimal stage of development, generally after 7 to 14 days of rearing.

After harvesting, the larvae were immobilized with hot water. Once treated, they were spread out in the sun to dry and passed through a press to extract the oil. Finally, the dried and defatted larvae were ground to obtain a fine flour.

Chicken viscera meal production

Chicken guts obtained fresh from the market were meticulously washed and rinsed. This step was crucial to remove impurities, food residues and contaminants, thus ensuring the cleanliness of the intestines prior to processing. After this stage, they were then subjected to heat treatment by boiling at 100 degrees Celsius for a quarter of an hour. After draining, the viscera were spread out in the sun to dry. Once dried, they were ground into a fine flour and preserved in glass jars.

Diets formulation

Four experimental diets were formulated using the Pearson square method, and an industrial feed was used as a control. Table I provides information on the centesimal composition of the different ingredients used and the composition of the different ingredients in the industrial feed, as well as their bromatological compositions.

TableI:- Centesimal composition of the four formulated diets.

	1			
Ingredients	R1 (viscera)	R2 (viscera + R3 (vis	scera R4 (viscera +	Industrial food
		blood) +BSF)	blood +BSF)	

Fish meal	35	35	35	35	Fish meal,
Soybean meal	10	10	10	10	Soya cake meal,
Peanut meal	10	10	10	10	Wheat meal,
Wheat bran meal	10	10	10	10	Soya oil,
Corn flour	10	10	10	10	Maize meal,
Cassava flour	9	9	9	9	Rice bran,
Chicken viscera	5	2,5	2,5	1,66	Distillers Dried
meal					Grains with
Cow's blood meal	0	2,5	0	1,67	Solubles (DDGS)
BSF larvae meal	0	0	2,5	1,67	and
Fish oil	5	5	5	5	Maize gluten
Premix vitamins ^a	1	1	1	1	
Premix minerals ^b	1	1	1	1	
Yeast	4	4	4	4	
Total	100	100	100	100	
Crude Protein (%)	37,5	37,5	37,5	37,5	37
Crude lipid (%)	11,8	11,37	11,43	11,44	8
Cost of feed (F	657,22	654,72	691,43	666,415	1050
CFA)					

a=vit A 250000 UI; vit D3 250000UI; vit E 5000mg; vit B1 100mg; vit B2 400mg; vit B3(pp) 1000mg; vit B5 pantode Ca2000mg; vit B6 300mg; vit K3 1000g; vit C 5000mg; H biotin 15mg; choline 100g; anti-oxydant (BHT), crushed and calcined attapulgite qs 1000mg;

b=phosphorus 7%; calcium 17%; sodium 1,5%; potassium 4,6%; magnesium 7,5%; manganese 738mg; zinc 3000mg; iron 4000mg; copper 750mg; iodine 5mg; cobalt 208mg; calcined and ground attapulgite qs 1000g; fluorine 1.5% (approximately),

Diets manufacturing

The different proportions of ingredients shown in Table I are formulated to produce 1000g of feed. The raw materials for each feed were sieved, weighed and mixed to homogenize them. Water was then added at a rate of 30% of dry matter. This resulted in a malleable dough mixture. This mixture was passed into a Moulinex mincer, from which it emerged in the form of spaghetti filaments. These filaments then underwent fragmentation, followed by sun-drying for two days. Once dried, they were placed in a mortar, then crushed and sieved to obtain flour, which was then preserved in bags.

Breeding conditions

The hybrids used in this experiment were derived from the artificial reproduction of a male Clariasgariepinus and a female Clariasanguillaris. Three days after hatching, the larvae were fed artemia SEP for 15 days. During the course of the experiment, 300 fry with an initial average weight of 1.56 g were randomly distributed in 15 aquariums of 50-liter, i.e. 20 fry per tank. These fish were acclimatized for one week.

The experiment lasted 45 days. The fish received a daily ration corresponding to 8% of their live weight. This ration was distributed in the morning at 8 a.m. and in the evening at 5 p.m., after temperature and pH measurements had been taken.

Finally, to maintain water quality, the bottom of the rearing tanks was siphoned off eevery morning before feeding. The edges were cleaned and rinsed thoroughly to limit the deposit of any fatty matter left by the feed.

Evaluation of survival growth parameters, feed efficiency and feed costs

Growth checks were carried out early in the morning after every two weeks of the experiment. Using a landing net, the fish were transferred to a plastic pot placed on a scale. After weighing, the fry were returned to the carefully cleaned aquariums. The results obtained were then entered into an Excel spreadsheet to determine zootechnical parameters for assessing fish growth and feed utilization efficiency. The various zootechnical parameters used as indicators in this experiment are:

Specific Growth rate (SGR, %/d)= (ln(final average weight)-ln(initial average weight)/experiment duration X100.

Survival Rate (SR, %)= number of final fish / number of initial fish X100.

Feed Conversion Ratio (FCR) = Feed intake / weight gain .

Absolute Mean Weight Gain (AMWG, g/fish) = average final weight - average initial weight

Relative Mean Weight Gain (RMWG, %)= 100 x ((final average weight - initial average weight)/initial average weight))

Cost of feed (FCFA) = Total costs (raw materials, processing, transport) /Quantity of feed produced per kg Production cost per kg of fish (FCFA)= FCR× Feed cost (per kg)

Data analysis

Analysis of statistical differences between groups was carried out using ANOVA and Duncan test, implemented via SAS software, which is a widely used computer program for statistical data analysis and database management. Created in the 1970s by SAS Institute, it is particularly popular in fields as diverse as social sciences, bio statistics. It enabled us to identify homogeneous groups based on their significant differences. Thanks to SAS' advanced features, the averages were automatically grouped into distinct classes, facilitating interpretation of the results. This method proved effective in highlighting significant differences while controlling the risk of Type I error, in line with the study's objectives

Results and Discussion

Results

Yield of main ingredients used

The yields of the main ingredients used as additives are shown in Table II

TableII:-Yield of main ingredients used as additives.

Ingredients	Initial quantity	Final quantity (g)	Yield (%)
Cow's blood	17 L	7000	41,48
Black soldier fly larvae	1.70 kg	465	27,35
Chicken viscera	20 kg	800	4

Table II shows a significant variation in yields between processed ingredients. Cow's blood shows the highest yield (41.18%), reflecting efficient processing with moderate losses, making it an economically advantageous option. Black soldier fly larvae, with a yield of 27.35%, show losses linked to their high-water content, but remain attractive due to their high nutritional value and ecological impact. Chicken intestines, on the other hand, had a very low yield (4%), indicating significant losses during the process, probably due to a high proportion of unusable material. These results suggest that cow's blood and fly larvae should be preferred, while chicken intestine processing needs to be optimized or alternative uses considered to reduce losses and improve recovery

Biochemical composition of mainingredients

The various ingredients used, in particular cow blood meal, chicken intestine meal and black soldier fly larvae meal, were sent to LANAC for biochemical analysis using the AOAC, 1995 method. The results of the biochemical composition are presented in Table III.

Table III:Biochemical composition of blood meal from Black Soldier Fly (BSF) larvae and chicken viscera meal

Biochemical composition	Blood meal	BSF meal	Chicken viscerameal
Fat content (%)	1,97	14,32	19,33
Protein (%)	93,81	32,37	61,25

Water quality parameters (Temperature and pH variations)

TableIV: Changes in temperature and pH during experiment.

Parameters	R0	R1	R2	R3	R4
Temperature	28,61±0,005 ^a	28,42±00°	28,45±0,01°	28,45±0,03°	28,50±0,04 ^b
pН	7,83±0,005 ^a	$8,04\pm0,02^{b}$	7,99±0,01°	$7,96\pm0,02^{d}$	$7,93\pm0,01^{e}$

Table IV shows the results of statistical analyses of water parameters, namely temperature and pH, for five different feeding diets (R0, R1, R2, R3, R4). Values shownincludemeans and their standard deviations, while letters (a, b, c, d, e) show significant differences between treatments.

For temperature, diet R0 (28.61°C) had a significantly higher temperature than all other diets. Diets R1, R2, and R3 (28.42; 28.45; 28.45°C) had statistically similar temperatures. Diet R4 (28.50°C) was slightly higher than R1, R2, and R3, but still significantly different from R0 and the others.

In terms of pH, diet R0 (7.83) had the lowest pH and was significantly different from the other diets. Diet R1 (8.04) showed the highest pH and was significantly different from all other treatments. Diets R2 (7.99) and R3 (7.96) had close but statistically distinct values. Diet R4 (7.93) has the lowest pH after R0, but remains significantly different from the others.

Zootechnical parameters

During the experiment, various zootechnical parameters were studied to determine fish growth in each diet. These results are shown in Table V.

TableV:Growth, feed efficiency and survival parameters of catfish hybrids.

Parameters	R0	R1	R2	R3	R4
IMW g	1,56 ^a	1,56 ^a	1,56 ^a	1,56 ^a	1,56 ^a
FMW g	10,62±0,25 ^a	5,65±0,15 ^d	7,85±0,23°	5,82±0,38 ^d	9,69±0,11 ^a
AMWG g	9,07±0,25 ^a	6,09±3,34 ^{ba}	$6,09\pm0,49^{ba}$	4,27±0,39 ^b	8,13±0,11 ^{ba}
RMWG%	582,76±16,81 ^a	$262,46\pm17,59^{d}$	390,73±31,89°	274,56±25,44 ^d	521,58±7,36 ^{ba}
FCR	$1,45\pm0,05^{b}$	2,41±0,20°	$1,66\pm0,18^{b}$	2,28±0,36 ^a	$1,44\pm0,06^{b}$
SCR%/J	4,27±0,06 ^a	2,86±0,05°	3,53±0,14 ^b	$2,93 \pm 0,15^{c}$	4,06±0,03°
SR%	93,33±5,77 ^a	93,33±2,88 ^a	90±0,10 ^a	85±13,23 ^a	95±5 ^a

NB: The letters a, b, c and d show the differences between the results of the statistical analyses.

TheIMW value was constant at 1.56 g for all diets (R0 to R4). This indicates that all fish groups had a homogeneous initial weight, which is essential to ensure that the differences observed in the other parameters are not due to variations in initial weight.

With regard to growth parameters (FMW, AMWG, RMWG, SGR), no statistically significant differences were observed between fish fed diets R0 and R4. Diets R1 and R3 showed no significant differences between them, but were both significantly lower than R0 and R4. For SGR, no significant differences were observed between fish fed diets R0 and R4, nor between R1 and R3. On the other hand, fish fed the R2 diet showed statistical differences from R0, R4, as well as R1 and R3.

In terms of feed efficiency (FCR), fish fed with R4 and R0 diets recorded the best FCR with values of 1.44 and 1.45 respectively, followed by those fed with R2 diet (1.66). However, these three diets had no significant statistical difference and were better than R1 (2.41) and R3 (2.28), which had no significant difference. On the other hand, fish fed diets R0, R2, R4 and those fed diets R1(2.41), R3(2.28) had FCR that were statistically different.

With regard to survival rates, the results obtained in this study do not show a significant difference for fish fed the R0, R1, R2, R3 and R4 diets. Fish fed the R3 diet had the lowest survival rate, with a value of 85%.

Economic analysis

TableVI: Economic analysis of tested diets.

Parameters	R0	R1	R2	R3	R4
Cost of Kg of feed	1050	657,22	654,72	691,43	666,41
(FCFA)					
Cost of Kg of fish	1523,91±50,2	1585,86±137,7	1087,76±119,7	1579,70±250,3	958,94±39,5
(FCFA)	6 ^a	2 ^a	6 ^b	3 ^a	8 ^b

NB: The letters a and b show the differences between the results of the statistical analyses.

Table VI shows the economic analysis of the diets tested. For feed costs, diet R0 (industrial feed) has the highest cost (1050 FCFA), while diets R1, R2, R3 and R4 have significantly lower costs (around 657 to 691 FCFA).

Production costs per Kg of fish vary between groups, with R4 (958.94 FCFA) and R2 (1087.76 FCFA) showing the lowest costs and no significant statistical difference. Diets R0, R1, and R3 have the highest costs, exceeding 1500 FCFA and showing no significant statistical difference. However, R4, R2 and R0, R1, R3 are statistically different.

Fish flesh analysis

TableVII: Results of bromatological analysis of fish flesh.

Parameters	Initial fish	R0	R1	R2	R3	R4
Crude	$16,81\pm0,10^{a}$	17,09±0,46 ^a	16,82±0,10 ^a	16,58±0,20°	$16,74\pm0,84^{a}$	17,05±0,76 ^a
Protein (%)						
Crude Fat	$3,94\pm0,19^{b}$	$6,40\pm0,43^{a}$	$4,31\pm0,54^{b}$	$4,22\pm0,19^{b}$	$4,57\pm0,15^{b}$	5,70±0,24 ^a
(%)						
Ash (%)	3.94±0.14 ^{bac}	3.49 ± 0.26^{dc}	4.25±0.19 ^{ab}	4,53±0,13 ^a	3.76 ± 0.22^{dc}	$3,28\pm0,31^{d}$

NB: The letters a, b, c and d show the differences between the results of the statistical analyses and

Bromatological results show that crude protein remains constant between groups, with values ranging from 16.58% to 17.09%, with no significant difference. On the other hand, Crude fat contentvaries significantly: group R1 has the highest content (6.40%), while the other groups (R0, R2, R3 and R4) have lower values, ranging from 3.94% to 4.57%, with no significant statistical difference. In terms of ash, groups R2 and R3 stand out with higher contents (4.25% and 4.53%, respectively), indicating better mineralization. Conversely, groups R4 (3.28%) and R1 (3.49%) show the lowest values, statistically different.

Discussion

Temperatures recorded during the experiment ranged from 26.6°C to 30.1°C. These results are in line with those of Alofaet al. (2016), who argued that Clarias can withstand temperatures as low as 6°C and as high as 50°C. However, a temperature range of 28 to 30° C is considered optimal for the growth of Clariasgariepinusand Clariasanguillaris(Adebayo et al.,2008).

The pH ranged from 7.11 to 8.94. These results fall within the range reported by Geoffroy et al. (2019), who summarized the literature on biological and zootechnical parameters of the African catfish Clariasgariepinusand argue that Clariasgariepinussurvives and thrives best in water with a pH between 6 and 9.

In the present study, the highest growth was observed in diet R4 (a mixture of blood meal, chicken viscera meal and black soldier fly larvae meal).

Mbaye (2023) has shown that blood meal and black soldier fly larvae meal represent promising alternative protein sources in animal feed. Protein-rich bloodmeal and black soldier fly larvae meal were evaluated for their nutritional benefits, but it was observed that the higher the fishmeal substitution, the higher the FCR. In terms of feed efficiency (FCR), fish fed the diet containing 5% bloodmeal recorded the best FCR. The best growth performance was also noted in fish fed the diet (5% bloodmeal). These results are similar to those of Olukunleetal.(2002), who reported that optimal growth and nutrient utilization in hybrid Clarias (male Clariasgariepinusx female Heterobranchuslongifilis) were achieved with 5% sun-dried bloodmeal.

Similarly, fly larvae meal, recognized for its durability and nutritional value, has shown significant potential as an additive. Study have suggested that the optimal incorporation rate for this insect meal should not exceed 30%, due to its chitin content (Gougbedji, 2022).

In this study, the R1 diet containing 5% chicken viscera meal had the highest FCR, followed by the R3 diet containing 2.5% chicken viscera meal and 2.5% black soldier fly meal. These results concur with those of Amtuletal. (2012) who worked on replacing fish meal with poultry by-product meal (chicken intestine) as a protein source in carp fry diets. Diets containing chicken intestine meal, even at the lowest level, significantly limited growth performance and feed efficiency. Values deteriorated as the incorporation rate of chicken intestine meal increased. This was due to the ash and carbohydrate content of the gut meal, producing a faster intestinal transit rate, leading to increased feed intake, associated with poor growth performance and feed efficiency. Similar results were reported by Ronyaiet al. (2002) and Fagbenroetal. (2004), who revealed that deficiency in one or more amino acids limits protein synthesis and affects fish growth

The R2 diet containing 2.5% bloodmeal and 2.5% chicken viscera meal has a FCR of 1.66, and these results are in line with those of Milliamena (2002) and Guo et al. (2007) who obtained better results when using bloodmeal to balance lysine content by replacing fishmeal with poultry byproducts (including broiler viscera) used alone or in combination with other ingredients of animal origin. According to Hu et al. (2008a), methionine and lysine are present in small quantities in chicken viscera (1.43 and 3.51% respectively). This could account for the low growth achieved in fish diets fed chicken viscera. The use of cow blood meal can correct lysine deficiency due to its richness in this amino acid (Hertrampf and Piedad-Pascual, 2000 and Alofa et al., 2016). However, methionine deficiency remains. Although poultry by-products such as chicken viscera have proven to be nutritionally adequate protein sources for many fish species (Fowler, 1991; Robainaetal.,1997; Abdel-Warith et al., 2001; Wang etal., 2006), they were deficient in essential amino acids such aslysine, methionine that limited their utilization (Glencross et al., 2007). Therefore, it will be beneficial to adjust the level of essential amino acids in chicken viscera through supplementation. Hu et al.(2008b) provided 0.49% lysine and 0.14% methionine as supplementation in diets containing chicken viscera with improved growth performance in Carassius auratusgibelio. Similar trends in results were obtained by Rawles et al. (2006) in the Morone chrysops x Morone saxatilis hybrid fed a broiler viscera diet supplemented with methionine (0.63%) and lysine (1.55%)

The results of this study show that the diet containing a mixture of chicken viscera meal, cow blood meal and black soldier fly meal, with a total incorporation rate of 5%, offers the best growth performance for catfish hybrid fry of Clariasgariepinusand Clariasanguillaris. This success can be attributed to the nutritional complementarity of the three ingredients: chicken viscera meal, rich in digestible proteins and deficient in amino acids including lysine and methionine according to Wang et al. (2006), cow blood meal which provides iron-rich proteins beneficial to hematopoiesis (Olukunleet al., 2002) and black soldier fly meal, a sustainable source of essential fatty acids and balanced proteins (Gougbedji, 2022; Henry et al., 2015). The synergistic effect observed with this blend, highlighted by Santiago and Lovell (1988) and Youssouf and Sedro (2020), probably results from the combination of diversified nutritional inputs that optimize nutrient digestibility and assimilation. By keeping the incorporation rate low (≤ 5%), negative effects such as reduced palatability or digestibility were avoided, as noted by El Sayed et al. (1998). These results also confirm the findings of similar studydone by Youssouf and Sédro (2020) who reported that diets combining different animal and insect protein sources improve growth performance and feed conversion in fish.

The results in Table VIII highlight the economic efficiency of treatments R4 and R2, characterized by significantly lower production costs per Kg of fish, respectively 958.94 and 1087.76 F CFA compared to R0, R1 and R3, where these costs exceed 1500 FCFA. These data corroborate the studies by Tacon and Metian (2015), who emphasized the importance of optimized feed formulation to reduce production costs in aquaculture, by favoring accessible and digestible ingredients. Furthermore, the discrepancy observed between treatments may be explained by variations in feed conversion rate (FCR), a key parameter influenced by feed composition and rearing conditions, as suggested by Boyd and Tucker (2012). Thus, the results reinforce the idea that the use of adapted formulations (R4 and R2) can improve profitability while reducing the economic and environmental impact of fish production. Results for overall fish body composition showed similar protein content in all treatments.

However, lipid content increased, probably due to the level of chicken viscera meal in the diets. These results are in agreement with Goda et al.(2007), Giri et al.(2010) and Sugumaran et al.(2015), who reported that replacing fish meal with poultry viscera meal in the diets did not affect the body protein content, but increased the whole-body lipid content of the fish.

Conclusion

This study is part of an innovative approach to evaluate the effects of cow blood meal, chicken intestine meal and black soldier fly larvae meal as additives in the feed of hybrid fry of Clariasgariepinusand Clariasanguillaris. The results obtained show that these ingredients, used together at a total incorporation rate of 5%, offer significantly better growth performance and feed efficiency than the other diets tested.

Diets containing the mixture of these three ingredients were well accepted by the fish, with no pathological symptoms or negative impacts on their health. Environmental conditions, notably temperature and pH, were favorable throughout the experiment. These observations underline the potential of these raw materials as viable and sustainable alternatives to conventional fishmeal.

In order to consolidate the results obtained in the present study, it would be relevant to analyze the impact of the mixture on the fish immune system and disease resistance, and to study the long-term effects of these additives on the health and overall performance of fish in different production systems.

References:-

- 1. Abdel-Warith AA, Russell PM, Davies SJ. Inclusion of a commercial poultry by-product meal as a protein replacement of fish meal in practical diets for African catfish Clariasgariepinus (Burchell 1822). Aquaculture Research. 2001; 32:296-306.
- 2. Adebayo, O. T., Fagbenro, O. A., & Popoola, O. M. (2008). Effects of temperature on hatching and larval survival of Clariasgariepinus (Burchell, 1822). Journal of Fisheries and Aquatic Science, 3(4), 302-307
- 3. Alofa, C.S., & Abou, Y. (2020). A comparison between chicken viscera and housefly maggot cultured from this by products for Nile tilapia Diets: Growth Performance, Feed Utilization and Whole Body Composition. Asian Journal of Fisheries and Aquatic Research, 5(3), 1-12. https://doi.org/10.9734/ajfar/2019/v5i330075
- 4. Alofa, C.S., Oke V., & Abou, Y. (2016). Effect of replacement of fish meal with broiler chicken viscera on growth, feed utilization and production of African catfish Clariasgariepinus(Burchell, 1822). International Journal of Fisheries and AquaticStudies, 4(6), 182-186.
- 5. Amtul, B.T., and Amna, B., 2012. Replacement of Fish Meal with Poultry by-Product Meal (Chicken Intestine) as a Protein Source in Grass Carp Fry Diet. Pakistan J. Zool. vol. 44(5), pp. 1373-1381, 2012.
- 6. Baldé, B.S., 2019. Dynamics of small pelagic fish (Sardinella aurita and Ethmalosa fimbriata) in Senegal in a context of climate change: bioecological diagnosis and synthesis (PhD thesis). Université Cheikh Anta Diop, Dakar, Senegal.
- 7. Barragan-Fonseca, K.B.; Dicke, M.; van Loon J.J.A., 2017. Nutritional value of the black soldier fly (Hermetiaillucens L.) and its suitability as animal feed a review. J. Insects as Food Feed 3(2), 105-120.
- 8. Bishop, C. D., Angus, R. A. and Watts, S. A. (1995). The use of feather meal as a replacement for fish meal in the diet of Oreochromis niloticusfry. BioresourceTechnology, 54, 291-295.
- Bloukounon-Goubalan A.Y., Saïdou A., Chrysostome C.A.A.M., Kenis M., Amadji G.L., Igué A.M. & Mensah G.A., 2020. Physical and Chemical Properties of the Agroprocessing By-products Decomposed by Larvae of Musca domestica and Hermetiaillucens. Waste and Biomass Valorization 11(6), 2735-2743.
- 10. Booth, Donald C., and Craig Sheppard. "Oviposition of the black soldier fly, Hermetiaillucens (Diptera: Stratiomyidae): eggs, masses, timing, and site characteristics." Environmentalentomology 13.2 (1984): 421-423.
- 11. Boyd, C. E., & Tucker, C. S. (2012). Pond aquaculture water quality management. Springer Science & Business Media, p 699
- 12. Caruso D., Devic E., Subamia I.W., Talamond P. &Baras E., 2014. Technical handbook of domestication and production of diptera Black Soldier Fly (BSF), IPB Press, Bogor, Indonesia, 141.
- 13. Cuvelier C., Cabaraux J., Dufrasne I., Hornick J. &Istasse L., 2004. Fattyacids: nomenclature and dietary sources. Ann. MédecineVétérinaire 133-140.
- 14. Debode J., De Tender C., Soltaninejad S., Van Malderghem C., Haegeman A., Van der Linden I., Cottyn B., Heyndrickx M. &Maes M., 2016. Chitin mixed in potting soil alters lettuce growth, the survival of zoonotic bacteria on the leaves and associated rhizosphere microbiology. Front. Microbiol. 7(APR), 1-15.
- 15. Dehaumont P. (1982). Les problèmes posés par la valorisation du sang d'abattoirs d'animaux de boucherie. R.T.V.A.,183: 23-32.
- 16. Diener S., Zurbrügg C. &Tockner K., 2009. Conversion of organic material by black soldier fly larvae: establishing optimal feeding rates. Waste Manag. Res. 27(6), 603-610.
- 17. Diener S., Zurbrugg C., Roa Gutiérrez F., Nguyen H.D., Morel A., Koottatep T. &Tockner K., 2011. Black soldier fly larvae for organic waste treatment prospects and constraints. In: WasteSafe 2011 2nd International Conference on Solid Waste Management in Developing Countries 1315 February 2011 Khulna Bangladesh. 978-984.
- 18. El-Sayed, A. F. M. (1998). Total replacement of fish meal with animal protein sources in Nile tilapia, Oreochromis niloticus(L.), feeds. Aquaculture Research, 29, 275-280;
- 19. Ewald N., Vidakovic A., Langeland M., Kiessling A., Sampels S. &Lalander C., Fagbenro OA, Balogun AM, Fasakin EA, Bello-Olusoji OA. 1998b. Dietary lysine requirement of the African catfish Clariasgariepinus. Journal of Applied Aquaculture; 8(2):71-77.
- 20. Fagbenro OA. 2004. Soybean meal replacement by arugula (Eruca sativa Miller) seed meal as protein feedstuff in diets for African Catfish, Clariasgariepinus(Burchell 1822), fingerlings. Aquaculture Resource. 35:917 923.
- 21. FAO, 2020. The State of World Fisheries and Aquaculture. Roma, Italy, ISBN 97892-5132692-3 224p.
- 22. Fly Produces a Foodstuff High in Omega-3 Fatty Acids. J. World Aquac. Soc. 38(2), 309313.

- 23. Fowler LG. 1991. Poultry by product meal as a dietary protein source in fall Chinook salmon diets, Aquaculture. 99:309-321.
- 24. Giannetto A., Oliva S., CecconLanes C.F., de AraújoPedron F., Savastano D., Baviera C., Parrino V., Lo Paro G., Spanò N.C., Cappello T., Maisano M., Mauceri A. &Fasulo S., 2020. Hermetiaillucens(Diptera: Stratiomydae) larvae and prepupae: Biomass production, fatty acid profile and expression of key genes involved in lipid metabolism. J. Biotechnol. 307: 44-54.
- 25. Gilles S., Dugué R., Slembrouck J., (2001) Manuel pratique de pisciculture du poisson-chat africain (Clarias gariepinus) page
- 26. Giri SS, Sahoo SK, Mohanty SN. 2010. Replacement of by catch fishmeal with dried chicken viscera meal in extruded feeds: effect on growth, nutrient utilisation and carcass composition of catfish Clariasbatrachus(Linn.) fingerlings Aquaculture Int. 18:539-544.
- 27. Glencross BD, Booth M, Allan GL. 2007. A feed is only as good as its ingredients- a review of ingredient evaluation strategies for aquaculture feeds. Aquaculture Nutrition. 3:17-34.
- 28. Goda, A.M., El-Haroun, E.R. & Chowdhury, M.A.K. (2007). Effect of totally or partially replacing fish meal by alternative protein sources on growth of African catfish Clariasgariepinus (Burchell, 1822) reared in concrete thanks. Aquaculture Research, 38, 279-287.
- 29. Goddard, S., Al-Shagaa, G. and Ali, A. (2008). Fisheries by-catch and processing wastemeals as ingredients in diets for Nile tilapia, Oreochromis niloticus. Aquaculture Research, 39, 518-525;
- 30. -Guo, J., Wang, Y. & Bureau, D.P. (2007). Inclusion of rendered animal ingredients as fishmeal substitutes in practical diets for cuneate drum, Nibeamiichthioides (Chu, Lo and Wu). Aquaculture Nutrition, 13, 81-87.
- 31. Henry M., Gasco L., Piccolo G. &Fountoulaki E., 2015. Review on the use of insects in the diet of farmed fish: Past and future. Anim. Feed Sci. Technol. 203 (1), 1-22.
- 32. Hernandez, C., Olvera-Novoa, M. A., Hardy, R. W., Hermosillo, A., Reyes, C. and Gonzalez, B. (2010). Complete replacement of fish meal by porcine and poultry by-product meals in practical diets for fingerling Nile tilapia Oreochromis niloticus: digestibility and growth performance. Aquaculture Nutrition, 16, 44-53.
- 33. Hertrampf JW, Piedad-Pascual F. 2000. Handbook on ingredients for Aquaculture Feeds, Kluwer academic publishers, Dordrecht, Netherlands. 482-483.
- 34. Hoc B., Genva M., Fauconnier M.-L., Lognay G., Francis F. &CaparrosMegido R., (2020). About lipid metabolism in Hermetiaillucens(L. 1758): on the origin of fatty acids in prepupae. Sci. Rep. 10(1), 11916.
- 35. Holmes L., (2010). Role of Abiotic Factors on the Development and Life History of the Black Soldier Fly, Hermetiaillucens(L.)(Diptera: Stratiomyidae). Masters Abstr. Int.
- 36. Holmes L.A., Vanlaerhoven S.L. &Tomberlin J.K., 2012. Relative Humidity Effects on the Life History of Hermetiaillucens(Diptera: Stratiomyidae). Environ. Entomol. 41(4), 971-978.
- 37. Hu M, Wang Y, Wang Q, Zhao M, Xiong B, Qian X (2008a); Evaluation of rendered animal protein ingredients for replacement of fish meal in practical diets for gibel carp, Carassius auratusgibelio (Bloch), Aquaculture research. 39:1474-1482.
- 38. Hu M, Wang Y, Wang Q, Zhao M, Xiong B, Qian X (2008b). Replacement of fish meal by rendered animal protein ingredients with lysine and methionine supplementation to practical diets for gibel carp, Carassius auratusgibelio. Aquaculture. 275:260-265
- 39. Kim W., Bae S., Park H., Park K., Lee S., Choi Y., Han S. & Koh Y., 2010. The Larval Age and Mouth Morphology of the Black Soldier Fly, Hermetiaillucens(Diptera:
- 40. Stratiomyidae). Int. J. Ind. Entomol. 21(2), 185-187.
- 41. Lévêque C. &Paugy D., 2006. Les poissons des eaux continentales africaines: Diversité, écologie, utilisation par l'homme, 59-74.
- 42. Liland N.S., Biancarosa I., Araujo P., Biemans D., Bruckner C.G., Waagbø R., Torstensen B.E. & Lock E.-J., (2017). Modulation of nutrient composition of black soldier fly (Hermetiaillucens) larvae by feeding seaweed-enriched media. PLoS One 12(8), e0183188.
- 43. Liu X., Chen X., Wang H., Yang Q., ur Rehman K., Li W., Cai M., Li Q., Mazza L., Zhang J., Yu Z. & Zheng L., (2017). Dynamic changes of nutrient composition throughout the entire life cycle of black soldier fly. PLoS One 12(8), e0182601.
- 44. Maier T., Leibundgut M., Boehringer D. & Ban N., 2010. Structure and function of eukaryotic fatty acid synthases. Q. Rev. Biophys. 43(3), 373-422.
- 45. Milliamena, O.M. (2002). Replacement of fish meal by animal by-product meals in a practical diet for grow-out culture of grouper Epinepheluscoioides. Aquaculture, 204, 75 84. https://doi.org/10.1016/s0044-8486(01)00629-9

- 46. Ndiaye, N. A., Maiguizo-Diagne, H., Diadhiou, H. D., Ndiaye, W. N., Diedhiou, F., Cournac, L.,&Brehmer, P. (2020). Methanogenic and fertilizing potential of aquaculture waste: towards freshwater farms energy self-sufficiency in the framework of blue growth. Reviews in Aquaculture, 12(3), 1435-1444.
- 47. Newton L., Sheppard C., Watson W.D., Burtle G. & Dove R., 2005. Using the black soldier fly, Hermetiaillucens, as a value-added tool for the management of swine manure. J. KoreanEntomol. Appl. Sci. 36(12), 17 pp.
- 48. Nguyen T.T.X., Tomberlin J.K. &Vanlaerhoven S., 2015. Ability of Black Soldier Fly (Diptera: Stratiomyidae) Larvae to Recycle Food Waste. Environ. Entomol. 44(2), 406410.
- 49. NRC, 2011. Nutrient requirements of fish and shrimp, Washington, DC, 376.
- 50. Olukunle, O.A., Ogunsanmi, A.O., Taiwo, V.O., and Samuel, A.A. (2002). The nutritional value of cow blood meal and its effect on growth performance haematology and plasma enzymes of hybrid catfish. Departments of wildlife and fisheries Management and Veterinary Pathology, university of Ibadan, Ibadan. Nigeria;
- 51. Oonincx D.G.A., Laurent S., Veenenbos M.E. & van Loon J.J.A., (2020). Dietary enrichment of edible insects with omega 3 fatty acids. Insect Sci. 27(3), 500-509.
- 52. Oonincx D.G.A.B., van Broekhoven S., van Huis A. & van Loon J.J.A., (2015). Feed Conversion, Survival and Development, and Composition of Four Insect Species on Diets Composed of Food By-Products. PLoS One 10(12), e0144601.
- 53. Oonincx D.G.A.B., van Itterbeeck J., Heetkamp M.J.W., van den Brand H., van Loon J.J.A. & van Huis A., (2010). An Exploration on Greenhouse Gas and Ammonia Production by Insect Species Suitable for Animal or Human Consumption. PLoS One 5(12), e14445.
- 54. Quilliam R.S., Nuku-Adeku C., Maquart P., Little D., Newton R. & Murray F., (2020). Integrating insect frass biofertilisers into sustainable peri-urban agro-food systems. J. Insects as Food Feed 6(3), 315-322.
- 55. Rawles SD, Riche M, Gaylord TG, Webb J, Freeman DW, Davis M. (2006). Evaluation of poultry by-product meal in commercial diets for hybrid striped bass (Morone chrysops male X M. saxatilis female) in recirculated tank production. Aquaculture, 259:377-389.
- 56. Reyes M., Rodríguez M., Montes J., Barroso F.G., Fabrikov D., Morote E. &SánchezMuros M.J., (2020). Nutritional and Growth Effect of Insect Meal Inclusion on Seabass (Dicentrarchuslabrax) Feeds. Fishes(2), 16-0.
- 57. Robaina L, Moyano FJ, Izquierdo MS, Socorro J, Vergera JM, Montero D. Corn gluten meal and meat and bone meals as protein sources in diets for gilthead seabream Sparus aurata nutritional and histological implications. Aquaculture. 1997; 157:347-359.
- 58. Rónyai A, Csengeri I, Váradi L (2002). Partial substitution of animal protein with full-fat soybean meal and amino acid supplementation in the diet of Siberian sturgeon (Acipenser baerii). J ApplIchthyol.; 18:682-684.
- 59. Sanogo S, Compaore I, Senou I, Somda M B, Ouedraogo R B, Ouattara B And Kabre Ta. (2020). Etude comparée de la structuration des macro invertébrés benthiques de cours d'eau urbain et péri-urbain à l'Ouest du Burkinafaso. International Journal of Development Research Vol. 11, Issue, 01, pp. 4317;
- 60. Scala A., Cammack J.A., Salvia R., Scieuzo C., Franco A., Bufo S.A., Tomberlin J.K. &Falabella P., 2020. Rearing substrate impacts growth and macronutrient composition of Hermetiaillucens(L.) (Diptera: Stratiomyidae) larvaeproduced at an industrial scale. Sci. Rep. 10 (1), 19448.
- 61. Spranghers T., Ottoboni M., Klootwijk C., Ovyn A., Deboosere S., De Meulenaer B., Michiels J., Eeckhout M., De Clercq P. & De Smet S., (2017). Nutritional composition of black soldier fly (Hermetiaillucens) prepupae reared on different organic waste substrates. J. Sci. Food Agric. (8), 2594-2600.
- 62. Stanley-Samuelson D.W., Jurenka R.A., Cripps C., Blomquist G.J. & de Renobales M., 1988. Fatty acids in insects: Composition, metabolism, and biological significance. Arch. InsectBiochem. Physiol. 9(1), 1-33.
- 63. St-Hilaire S., Cranfill K., McGuire M.A., Mosley E.E., Tomberlin J.K., Newton L., Sealey W., Sheppard C. & Irving S., 2007. Fish OffalRecycling by the Black Soldier.....
- 64. St-Hilaire S., Sheppard C., Tomberlin J.K., Irving S., Newton L., McGuire M.A., Mosley E.E., Hardy R.W. & Sealey W. (2007). Fly Prepupae as a Feedstuff for Rainbow Trout, Oncorhynchus mykiss. J. World Aquac. Soc. 38(1), 59-67.
- 65. Sugumaran, E. and Radhakrishnan, M.V. (2015). Feed utilization, growth and carcass composition of catfish Clariasbatrachus (L.) fed on fish meal replaced by dried chicken viscera incorporated diets. International Journal of Research in Fisheries and Aquaculture 5, 143-146.
- 66. Tacon A.G.J. &Metian M. (2008). Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: Trends and future prospects. Aquaculture 285(1-4), 146-158.

- 67. Tacon, A. G. J. and Jackson, A. J. (1985). Utilization of conventional and unconventional protein sources in practical fish feeds. In Nutrition and Feeding in Fish, Cowey CB, Mackie AM, Bell JG. (eds). Academic Press, London, 119 145.
- 68. Tacon, A. G. J., &Metian, M. (2015). Feed matters: Satisfying the feed demand of aquaculture. Reviews in Fisheries Science & Aquaculture, 23(1), 1-10.
- 69. Teugels G.G. & Thys van den Audenaerde D.F.E., (2003). Cichlidae. In: The Fresh and Brackish Water Fishes of West Africa. Paris, 521-600.
- 70. Teugels, (1986). A systematic revision of the African species of genus ClariasPices; Clariidae). Annales du musée royal d'Afrique centrale 247, 1-19
- 71. Tomberlin J.K., Adler P.H. & Myers H.M., (2009). Development of the Black Soldier Fly (Diptera: Stratiomyidae) in Relation to Temperature. Environ. Entomol. 38(Hogsette 1992), 930-934.
- 72. Tomberlin J.K., Sheppard D.C. & Joyce J.A., (2002). Selected Life-History Traits of Black Soldier Flies (Diptera: Stratiomyidae) Reared on Three Artificial Diets. Ann. Entomol. Soc. Am. 95(3), 379-386.
- 73. Truzzi C., Giorgini E., Annibaldi A., Antonucci M., Illuminati S., Scarponi G., Riolo P., Isidoro N., Conti C., Zarantoniello M., Cipriani R. &Olivotto I., (2020). Fatty acids profile of black soldier fly (Hermetiaillucens): Influence of feeding substrate based on coffeewastesilverskin enriched with microalgae. Anim. FeedSci. Technol. 259(October 2019), 114309.
- 74. Van Huis A. &Tomberlin J.K. (2017). Insects as food and feed: from production to consumption. van Huis, A., Tomberlin, J.K. eds, Wageningen Academic Publishers, The Netherlands, 448.
- 75. Viveen W.J.A.R., C.J.J. Richter, P.G.W.J. Van Oordt, J.A.L. Janssen, E.A. Huisman (1985). Practical manual for fish culture of African catfish (Clariasgariepinus), Netherlands, 128.
- 76. Wang Y, Li K, Han H, Zheng Z, Bureau DP. (2008). Potential of using a blend of rendered animal protein ingredients to replace fish meal in practical diets for malabar grouper (Epinephelusmalabaricus). Aquaculture281:113 117.
- 77. Zarantoniello M., Zimbelli A., Randazzo B., Compagni M.D., Truzzi C., Antonucci M., Riolo P., Loreto N., Osimani A., Milanović V., Giorgini E., Cardinaletti G., Tulli F., Cipriani R., Gioacchini G. &Olivotto I., (2020). Black Soldier Fly (Hermetiaillucens) reared on roasted coffee by-product and Schizochytriumsp. as a sustainable terrestrial ingredient for aquafeeds production. Aquaculture 518, 734659.
- 78. Zhou J.S., Liu S.S., Ji H. & Yu H.B., (2018). Effect of replacing dietary fish meal with black soldier fly larvae meal on growth and fatty acid composition of Jian carp (Cyprinus carpiovar .Jian). Aquac. Nutr. 24(1), 424-433.